Chronic Kidney Disease (CKD) Meets Nonsteroidal Anti-Inflammatory Drugs (NSAIDs): A Risky Interaction of Toxicity and Impaired Clearance

Anirban Banerjee¹, Samrat Bose^{* 2}, Soumita Das³, Shreya Sarkar⁴, Asfakul Islam¹, Shubham Kolay¹, Prasanna Roy⁵, Sreetama Moitra¹, Suvasish Mitra¹, Prasit Roy⁵, Bipasha Pal¹, Ankit Chatterjee⁶, Joyeta Ghosh¹

 ¹ Guru Nanak Institute of Pharmaceutical Sciences and Technology, 157/F Nilgunj Road, Panihati, Kolkata, West Bengal-700114
 ² JIS University, 81 Nilgunj Rd, Agarpara, Kolkata, West Bengal-700109
 ³ NSHM Knowledge Campus, 124 BL Saha Road, West Bengal-700053
 ⁴ Flemming College of Pharmacy, Balarampur, Beralia, West Bengal-700144
 ⁵ Gandhari College (School of Pharmacy), Garhbari, Nazir Bazar, West Bengal-721655
 ⁶ Dr. B. C. Roy College of Pharmacy and Allied Health Sciences, Dr. Meghnad Saha Sarani, Durgapur, West Bengal-713212

Corresponding Author Mr. Samrat Bose* Assistant Professor, Department of Pharmacy, JIS University Email: samratbose09@gmail.com

ABSTRACT

Chronic Kidney Disease (CKD) affects over 850 million individuals globally, posing a significant public health burden with rising morbidity and mortality, marked by progressive nephron loss and impaired renal clearance mechanisms. It substantially alters pharmacokinetics and enhances drug toxicity risk. Nonsteroidal anti-inflammatory drugs (NSAIDs), widely used analgesics, compromise renal hemodynamics through prostaglandin synthesis inhibition, potentiating nephrotoxicity. Evidence from large-scale cohort studies and meta-analyses reveals consistent associations between NSAID exposure and acute kidney injury (AKI), electrolyte disturbances, and accelerated CKD progression. Several guidelines, including the Kidney Disease: Improving Global Outcomes (KDIGO), recommend minimizing or avoiding NSAID use in CKD, especially when the estimated Glomerular Filtration Rate (eGFR) falls below 60 mL/min/1.73 m². While emerging data suggest short-term NSAID use may be permissible under strict supervision in select cases, the prevailing consensus remains conservative. Effective pain management in CKD must balance analgesic efficacy with nephron preservation through a guideline-directed approach. This review critically examines the mechanistic underpinnings and clinical consequences of NSAID use in CKD populations. It evaluates dose-dependent nephrotoxicity, interaction with the renin-angiotensin-aldosterone system (RAAS), and the cumulative burden in patients with comorbid conditions such as hypertension and myocardial infarction.

Keywords: Chronic Kidney Disease (CKD), Nonsteroidal Anti-inflammatory Drugs (NSAIDs), Nephrotoxicity, Clinical Guidelines

ABBREVIATIONS

AA – Arachidonic Acid; ACE – Angiotensin-Converting Enzyme; AKI – Acute Kidney Injury; ALLHAT – Antihypertensive and Lipid-Lowering Treatment to Prevent Heart Attack Trial; Ang II - Angiotensin II; ARB - Angiotensin Receptor Blocker; ATTACK - Aspirin To Target Arterial Events in CKD; CKD – Chronic Kidney Disease; (cTALH) – Cortical Thick Ascending Limb of Henle; cAMP - cyclic adenosine monophosphate; EGF - Epidermal Growth Factor; ER - Endoplasmic Reticulum; ERBP - European Renal Best Practice; ET - Endothelin; COX - Cyclooxygenase; CYP450 - Cytochrome P450 Enzyme System; eGFR - Estimated Glomerular Filtration Rate; ESRD - End-stage Renal Disease; GBD - Global Burden of Disease; hCOX – Human COX; JGA – Juxtaglomerular Apparatus; KDIGO – Kidney Disease: Improving Global Outcomes; MBD – Membrane-Binding Domain; MBF – Medullary Blood Flow; MD – Macula Densa; MI – Myocardial Infarction; MICs – Medullary Interstitial Cells; NSAID - Nonsteroidal Anti-Inflammatory Drug; NF-kB - Nuclear Factor-kB; PGs -Prostaglandins; PGD2 - Prostaglandin D2; PGDS - Prostaglandin D Synthase; PGE2 -Prostaglandin E2; PGES - Prostaglandin E Synthase; PGI2 - Prostacyclin; PGIS - Prostacyclin Synthase; PLA₂ – Phospholipase A₂; PRA – Plasma Renin Activity; TGF – Tubuloglomerular Feedback; TIPS-3 – The International Polycap Study-3;

1. Introduction

CKD is a progressive disorder characterized by a persistent reduction in renal function. It is typically characterized by a glomerular filtration rate (GFR) of <60 mL/min/1.73 m² or albuminuria \geq 30 mg/day or evidence of structural kidney damage persisting for at least three months [1][2][3]. CKD has emerged as a significant global health burden, with an estimated prevalence of 8–16% in the general population, affecting approximately 850 million individuals worldwide [3][4][5]. The growing prevalence of diabetes mellitus, hypertension, obesity, and aging populations primarily drives the increasing incidence of CKD. The Global Burden of Disease (GBD) study highlights CKD as one of the fastest-rising causes of mortality, with end-stage renal disease (ESRD) accounting for a substantial proportion of morbidity and healthcare expenditures [6][7]. The pathophysiology of CKD involves progressive nephron loss, maladaptive glomerular hyperfiltration, interstitial fibrosis, and inflammation, leading to a decline in renal function and systemic complications, including mineral-bone disorders, anemia, and metabolic acidosis [8][9][10].

| Stage of CKD | GFR (mL/min/1.73 m ²) | Description |
|--------------|-----------------------------------|--------------------------------------|
| Stage 1 | >90 | Normal kidney function or elevated |
| Stage 2 | 60-89 | Mild reduction in kidney function |
| Stage 3a | 45-59 | Mild to moderate decline in function |
| Stage 3b | 30-44 | Moderate to severe kidney |
| | | dysfunction |
| Stage 4 | 15-29 | Severe loss of kidney function |
| Stage 5 | <15 | End-stage kidney failure |

Table 1: Stages of Chronic Kidney Disease (CKD) Based on Glomerular Filtration Rate (GFR) [11]

NSAIDs represent one of the most widely prescribed pharmacological classes, utilized extensively for their analgesic, antipyretic, and anti-inflammatory properties [12][13][14]. NSAIDs function primarily by inhibiting the cyclooxygenase (COX) enzyme family, which is responsible for converting arachidonic acid into prostaglandins and thromboxanes—key mediators of inflammation, pain perception, and homeostatic functions [13][15][16]. COX enzymes exist in two primary isoforms: COX-1, which is constitutively expressed and involved in maintaining renal perfusion, gastrointestinal mucosal protection, and platelet aggregation, and COX-2, an inducible isoform predominantly associated with inflammation and pain modulation [17][18][19]. In individuals with normal renal function, prostaglandins play a compensatory role in maintaining GFR by countering the renin-angiotensin-aldosterone system (RAAS), particularly in conditions of volume depletion or renal hypoperfusion [20][21][22]. COX-1 inhibition contributes to adverse events, particularly in the renal system. In patients with compromised kidney function or CKD, prostaglandin synthesis disruption may lead to significant hemodynamic and nephrotoxic consequences [23][24].

Despite their efficacy, NSAIDs pose significant risks in patients with CKD due to their nephrotoxic potential. The reductions in prostaglandin-mediated vasodilation by NSAIDs can precipitate acute kidney injury (AKI), accelerate CKD progression, and contribute to electrolyte imbalances (hyponatremia and hyperkalemia), hypertension, and cardiovascular complications (including heart attack, stroke, and heart failure) [25][26][27].

This review comprehensively examines the interaction between NSAID therapy and CKD, emphasizing the mechanistic basis of NSAID-induced nephrotoxicity, alterations in NSAID pharmacokinetics in CKD patients, and the clinical consequences of NSAID use in this population. By synthesizing data from recent clinical studies, mechanistic investigations, and pharmacokinetic analyses, this review aims to critically appraise NSAID therapy in CKD and inform clinical decision-making for healthcare professionals managing this vulnerable patient population.

2. Molecular Structure of COX Protein

The human COX (hCOX) family comprises two primary isoforms: COX-1 and COX-2. These isoforms share approximately 70% sequence identity but differ significantly in their expression patterns, regulatory mechanisms, and physiological roles [28][18][29]. COX-1 is ubiquitously expressed in most tissues [*Table 2*] and is involved in maintaining renal function, gastric mucosal protection, and platelet aggregation. The inducible isoform is typically undetectable in most tissues under basal conditions and upregulated in response to pro-inflammatory stimuli, cytokines, and growth factors (such as Epidermal Growth Factor, TGF- β 1, and Vascular Endothelial Growth Factor) [18][30][31]. The constitutive COX-2 expression occurs in macula densa (MD), medullary interstitial cells (MICs), and glomerular podocytes, underscoring its crucial role in renal physiology [32][33]. Both isoforms catalyze the oxygenation of arachidonic acid to prostaglandin G₂ (PGG₂), followed by its reduction to PGH₂.

The structural complexity of cyclooxygenase enzymes underpins their functional versatility and interactions with therapeutic agents. hCOX enzymes are homodimers embedded in the endoplasmic reticulum (ER) and nuclear membrane, composed of 576 (consisting of 599 amino acids, with a mature form of 576 residues after signal peptide cleavage) and 581 (contains 604 amino acids, with a mature form of 581 residues, including an 18-residue insertion absent in COX-1) amino acid residues respectively [18][34][35]. Despite their high sequence homology, key sequence variations, including additional glycosylation sites in COX-2, influence their structural dynamics and regulatory mechanisms.

2.1. Domain Organization of COX

Cyclooxygenase enzymes are multi-domain and membrane-bound proteins. The domain architecture is crucial for comprehending these enzymes' catalytic mechanisms, membrane anchoring, and selective drug binding properties. The structural organization of each COX monomer is divided into three primary domains: the Epidermal Growth Factor (EGF)-like domain, the Membrane-Binding Domain (MBD), and the Catalytic Domain [18][36]. The EGF-like domain (residues 34–72) adopts a compact, β -sheet-rich structure, stabilized by disulfide bonds (characteristic of the EGF fold) [18][37][38]. This motif is common in extracellular and membrane-associated proteins, mediating protein-protein interactions. Crystallographic studies reveal that the EGF domain contributes to homodimerization, a prerequisite for enzymatic activity, as it facilitates proper heme incorporation and stabilizes the active site conformation. The C2 axis of symmetry observed in COX dimers is partly maintained by the EGF domain through hydrophobic and electrostatic interactions [39][40]. The EGF domain spatially positions the enzyme relative to the membrane surface. This orientation optimally aligns the MBD and the Catalytic Domain for substrate acquisition and processing [40][41].

The membrane-binding domain (residues 73–116) is an α -helical bundle consisting of 4 amphipathic helices (A–D), oriented parallel to the membrane surface. The helices are arranged such that hydrophobic residues face the lipid bilayer, embedding the enzyme in the membrane, while hydrophilic residues orient toward the aqueous cytoplasmic environment [18][42][43]. The MBD of COX enzymes partially inserts into the ER cytoplasmic leaflet and nuclear envelope membranes. This shallow embedding positions the enzyme optimally to capture AA and other polyunsaturated fatty acids released from phospholipids [40].



Figure 1: Schematic Illustration of the Structural Variations in the Substrate-Binding Channels of Cyclooxygenase-1 (COX-1) and Cyclooxygenase-2 (COX-2). Adopted from [60]

The catalytic domain is a multi-functional region containing a heme cofactor and two distinct active sites responsible for the sequential oxidation of AA to PGH₂, which consists of a sevenhelix bundle homologous to other heme peroxidases [29][44][45]. The heme-binding pocket is essential for enzyme function, held in place by conserved histidine residues (His-336 and 388) that coordinate the iron atom [40][46]. The heme iron undergoes cyclical oxidation-reduction, mediating electron transfer between the peroxidase and active sites. The peroxidase active site is near the heme-binding region, reducing hydroperoxides to alcohols, generating radicals essential for cyclooxygenase activity [40]. The active site of COX enzymes is a long, L-shaped (inverted) hydrophobic channel that extends from the membrane-binding domain to the core of the catalytic domain [*Figure 1; B*]. The active site is lined with crucial amino acid residues that facilitate substrate binding and catalysis, including Arg120, Tyr384, Ser530, Val349, Leu352, and Leu359 [28][18][35][47]. The active site volume of COX-2 is approximately 25% larger than that of COX-1, owing to a valine-to-isoleucine substitution (Val-523 in COX-2, Ile-523 in COX-1). This structural difference accounts for the selectivity of COX-2 inhibitors like celecoxib and rofecoxib [28].

2.2. Dimerization and Functional Assembly

COX is an example of an allosteric and cooperative enzyme that possesses an oligomeric state. It exists as homodimers, each monomer contributing to the overall enzymatic activity. Each subunit contains independent active sites, and functional asymmetry arises during catalysis, where one subunit acts as the "catalytic monomer" and the other as the "allosteric monomer" [48][49][50][51]. The dimer interface spans ~20% of the total protein surface area, involving residues primarily from the EGF-like domain and MBD. Hydrogen bonds and salt bridges stabilize the dimer interface of COX enzymes, and Trp-87 and Arg-120 form a critical hydrophobic clamp, anchoring the subunits [18][40][52][53]. Crystallographic and mutagenesis studies suggest that COX dimers are functionally asymmetric; the catalytic monomer binds AA and performs the cyclooxygenase and peroxidase reactions.

Meanwhile, allosteric monomer regulates catalytic efficiency by modulating heme redox states and substrate affinity through long-range conformational shifts [40][54][55]. This half-site reactivity ensures tight regulation of prostanoid synthesis, preventing excessive prostaglandin production and reducing oxidative stress. This dimerization-dependent regulation provides a sophisticated mechanism for spatiotemporal control of inflammatory responses. It highlights potential therapeutic opportunities for targeting the dimer interface to develop allosteric inhibitors with improved selectivity and safety profiles [40].

2.3. Sequence Variations and Structural Distinctions Defining COX Isoform Specificity

COX-1 is a housekeeping enzyme encoded on chromosome-09, involved in baseline prostaglandin production for renal homeostasis, gastroprotection, and platelet function. COX-2 is an inducible enzyme encoded from chromosome-01, upregulated in response to cytokines and pro-inflammatory stimuli [28][56]. Despite sharing ~70% sequence identity, COX-1 and COX-2 exhibit distinct structural features that underlie their divergent physiological roles and differential sensitivity to inhibitors. Unlike COX-1, COX-2 lacks a 14-residue segment near its N-terminus, which may contribute to its looser dimer interface and higher catalytic flexibility. COX-2 contains an 18-residue insertion in the catalytic domain (missing in COX-1), introducing subtle changes to the cyclooxygenase active site topology [29][57][58]. The N-linked glycosylation site (residue Asn-594) of COX-2 influences membrane interactions and enzyme stability [59][44]. COX-1 possesses an inverted L-shaped hydrophobic channel lined by smaller side chains, COX-2 has an enlarged active site due to Val-523 substitution, which introduces greater flexibility and accommodates bulkier substrates like arachidonic acid derivatives and endocannabinoids [28][35][47].

3. COX Enzymes in Kidney Function and Homeostatic Regulation

COX enzymes are indispensable regulators of renal homeostasis, orchestrating complex physiological processes. These isoforms exhibit distinct expression patterns and functional roles within the kidney, influencing basal and adaptive renal responses. COX-1 is constitutively expressed in renal tissues, including the glomeruli, collecting ducts, and renal vasculature [56][61][62]. Unlike COX-1, COX-2 exhibits dynamic, inducible expression, localized primarily to the MD, cortical thick ascending limb of Henle (cTALH), and MICs [61][63][64]. Despite their distinct expression patterns, COX-1 and COX-2 exhibit functional redundancy. The dynamic interplay between COX-1 and COX-2 ensures the kidney can respond flexibly to acute and chronic physiological stressors.



Figure 2: Illustration of COX-1 and COX-2 Localization in the Kidney: COX-1 (green) and COX-2 (blue) expression in various regions of the nephron, including the macula densa, proximal tubule, Henle's loop, and collecting duct.

| Table 2: Distribution and functional Role | s of COX-1 | and COX-2 in | Various Tissues |
|---|------------|--------------|-----------------|
|---|------------|--------------|-----------------|

| COX Isoform | Location | Function | | | |
|-------------|-------------|---|--|--|--|
| | Platelets | Constitutively expressed in most tissues; plays a role | | | |
| | | in platelet aggregation, gastric protection, ren function, and regulation of vascular tone. | | | |
| | | | | | |
| | Stomach | Involved in protecting the gastric mucosa by | | | |
| | | regulating the production of prostaglandins that | | | |
| | | maintain mucosal blood flow and bicarbon secretion. | | | |
| COX 1 | | | | | |
| COA-1 | Kidney | Participates in the regulation of renal blood flow and | | | |
| | | filtration. | | | |
| | Brain | Involved in homeostatic functions such as | | | |
| | | maintaining blood-brain barrier integrity and | | | |
| | | modulating cerebral blood flow. | | | |
| | Endothelium | Contributes to the regulation of vascular tone and | | | |
| | | platelet aggregation. | | | |

| | Inflammatory | Induced in response to inflammatory stimuli, it plays | | | |
|--------------------------------|-----------------|---|--|--|--|
| | sites (such as | a critical role in pain, fever, and tissue repair | | | |
| | joints, injured | processes. | | | |
| | tissues) | | | | |
| | Brain (induced) | Involved in pain processing, neuroinflammation, and | | | |
| COX-2 | | neurodegenerative diseases. | | | |
| KidneyRegulate(induced)inflamm | | Regulates renal function under stress, including | | | |
| | | inflammation or injury. | | | |
| | Endothelium | Upregulated in response to inflammatory or stress | | | |
| | (induced) | stimuli, contributing to inflammation and blood flow | | | |
| | | regulation. | | | |

3.1. Synthesis of Prostaglandins in the Kidney

PG biosynthesis is a highly regulated, multi-step enzymatic process that plays a fundamental role in physiological homeostasis and inflammatory responses. The COX enzyme family is a central molecule in this cascade, orchestrating AA's pivotal transformation into PGH2 [Figure 3]. This transformation proceeds via a stereospecific radical mechanism, ensuring precise structural fidelity of the final prostanoids [40][60][40][65]. The polyunsaturated ω -6 fatty acid AA (C20) remains esterified (as arachidonate) within membrane phospholipids under resting membrane potential. Its molecular integrity is tightly regulated by phospholipase A₂ (PLA₂), an enzyme that hydrolyzes membrane-bound AA in response to physiological stimuli, including mechanical stress, cytokines, and hormonal signals. The liberated AA serves as the substrate for cyclooxygenases, initiating the prostaglandin synthesis pathway [66][67][68]. The bifunctional heme-containing enzyme COX (COX 1 and 2) catalyzes the oxidation of AA following a tightly orchestrated radical-based mechanism, comprising three fundamental steps: substrate binding, oxygenation, and peroxidase reduction [65][69][70]. AA enters the COX active site via a hydrophobic channel (L-shaped) that accommodates its polyunsaturated tail. The peroxidase domain of COX, containing a heme prosthetic group, facilitates the generation of a tyrosyl radical at Tyr-385 (COX-2 numbering). These radicals abstract the 13pro-S hydrogen from AA, producing a carbon-centered pentadienyl radical, the reactive intermediate for subsequent oxygenation. This step results in the formation of PGG₂ (a hydroperoxide intermediate) [40][18][71][72]. The structural integrity of the active site dictates the stereoselectivity of these transformations. Particularly in COX-2, restricted binding orientations enforce regio- and stereospecificity. The formation of 15S-hydroperoxide at C15 ensures the correct prostanoid configuration [18][73]. The final step in COX-mediated prostaglandin synthesis involves the conversion of PGG₂ to PGH₂ by the peroxidase domain and is mediated by a heme-dependent electron transfer mechanism. Reducing peroxidase reduces the hydroperoxy group (-OOH) at C15 of PGG₂ to a hydroxyl (-OH) moiety. The resulting PGH₂ is the central prostanoid precursor for downstream enzymatic modifications [18][63][18][74].

3.1.1. Diversification of PGH₂

PGH₂ is an inherently unstable intermediate with an exceedingly short half-life. The immediate enzymatic conversion by tissue-specific isomerases and synthases converts PGH₂ into bioactive prostanoids. This enzymatic diversification ensures the precise regulation of prostaglandin-mediated physiological responses. Prostaglandin E Synthase (PGES) catalyzes the conversion of PGH₂ to PGE₂, a crucial inflammatory mediator that modulates vasodilation and immune responses [22][75][76][77]. Prostaglandin D Synthase (PGDS) facilitates the biosynthesis of PGD₂, a lipid mediator implicated in allergic reactions and sleep regulation [78][79][80][81]. Prostacyclin Synthase (PGIS) generates prostacyclin (PGI₂), a potent vasodilator that plays a critical role in endothelial function and the inhibition of platelet aggregation [82][83]. The tissue-specific expression of these enzymes dictates the functional specificity of prostaglandins, maintaining a delicate balance between hemostatic regulation and vascular homeostasis.

3.1.2. Regulatory Dynamics of Prostaglandin Synthesis

Calcium ion (Ca^{2+}) plays a pivotal role in modulating prostaglandin synthesis by influencing the release of AA and its subsequent enzymatic conversion into bioactive prostanoids.



Figure 3: Arachidonic acid metabolism pathway showing COX-1 and COX-2 enzymes, prostaglandin synthesis, and the effects of NSAIDs and inhibitors. Adopted from [77].

Arachidonic acid liberation is the rate-limiting step in prostaglandin synthesis and is catalyzed by cytosolic PLA₂. Ca²⁺ binds to the C2 domain of PLA₂, facilitating its translocation to perinuclear and plasma membranes, enhancing the hydrolysis of membrane phospholipids, and releasing free AA for COX-mediated conversion. The Secretory PLA₂ (sPLA₂; Predominantly active in the renal cortex) exhibits a Ca²⁺-dependent hydrolytic mechanism, contributing to local AA release under inflammatory conditions [84][85][86][87]. Calcium-Independent PLA₂ (iPLA₂) indirectly interacts with Ca²⁺ signaling, especially under ischemic conditions, modulating prostaglandin synthesis in the renal medulla [88][89][90]. Although COX enzymatic activity is Ca²⁺-independent, intracellular Ca²⁺ profoundly influences COX expression and prostaglandin synthesis by regulating AA substrate availability. Under stress conditions (such as ischemia, sodium depletion), predominantly in the macula densa and medullary interstitial cells, COX-2 responds to Ca²⁺-dependent activation pathways [91]. In glomerular mesangial cells, Ca²⁺ influx via L-type channels enhances COX-2 expression, promoting PGE₂-mediated vasodilation and autoregulation of GFR. The increased intracellular Ca²⁺ activates COX-2 in the proximal tubular cells [63][92]. COX-2-derived prostaglandins modulate afferent arteriolar tone, influencing glomerular filtration rates [93][24].

3.2. Regulation of Renal Blood Flow by COX

COX enzymes are integral to fine-tuning renal blood flow (RBF) and glomerular filtration dynamics due to the generation of bioactive prostanoids. PGs function as critical mediators in balancing vasodilatory and vasoconstrictive forces within the renal vasculature [94][95]. Dysregulation of this system, particularly through pharmacological inhibition of COX enzymes, can compromise renal perfusion and contribute to acute and chronic kidney injuries. The primary function of COX-1 is to sustain basal renal physiology by facilitating the synthesis of vasodilatory PGs (such as PGE₂ and PGD₂). COX–1–derived prostanoids mitigate the vasoconstrictive actions of angiotensin II and norepinephrine on afferent arterioles, preserving RBF and GFR under normal physiological conditions [24][94][96]. COX-2 becomes upregulated in response to volume depletion or high renin states, producing vasodilatory prostacyclin (PGI₂) to safeguard RBF under stress conditions [97][98].

3.2.1. Vasodilatory Actions of COX-Derived Prostaglandins

PGI₂, predominantly synthesized by endothelial COX-1 and COX-2, exerts its effects via IP receptors on vascular smooth muscle cells. It activates adenylate cyclase, increasing intracellular cyclic adenosine monophosphate (cAMP), which ultimately leads to vasodilation and ensures adequate perfusion under both physiological and pathophysiological conditions [95][99][100][101]. PGE₂ is produced via both COX isoforms, depending on the renal compartment, and is notably abundant in the thick ascending limb, collecting duct, and glomerular apparatus. Its vasodilatory actions are subtype-specific, predominantly mediated through EP₂ and EP₄ receptors, which similarly activate cAMP pathways [102][103][104]. Notably, PGE₂ modulates afferent arteriolar tone, safeguarding glomerular filtration during hypovolemia, ischemia, or sympathetic overactivity [105][106].

3.2.2. Vasoconstriction Counterbalance: Antagonism of Angiotensin II and Endothelin-1

The renal circulation is subject to complex neurohumoral control, wherein vasoconstrictors such as angiotensin II (Ang II) and endothelin-1 (ET-1) influence vascular tone. COX-derived PGs function as physiological counter-regulators in this highly regulated system [107][108][109][110]. Ang II induces constriction predominantly of the efferent arteriole, but

also affects the afferent arteriole during heightened systemic activation via AT₁ receptor activation [108][111][112][113]. However, COX-derived PGs (especially PGE₂ and PGI₂) mitigate these vasoconstrictive responses by enhancing local vasodilatory signaling [114][115]. Similarly, endothelin-1 (ET-1; a potent paracrine vasoconstrictor) reduces renal cortical and medullary blood flow through ETA receptors, and PGE₂ blunts the vascular and tubular effects of ET-1 [116][117][118].

3.2.3. Adaptation Mechanisms of RBF by COX

COX-2 is particularly inducible among the two isoforms and exhibits a dynamic expression profile within the renal parenchyma. A key site of its expression under physiological and stress-adaptive conditions is the renal MICs, which play a critical role in modulating medullary blood flow (MBF) and sodium homeostasis. Under high dietary sodium intake conditions, COX-2 expression is markedly upregulated in the inner medulla, particularly within the renal papilla. This phenomenon is believed to be a homeostatic mechanism aimed at promoting natriuresis and preventing volume overload [91][119][120][121][122]. However, COX-2 dynamically responds to fluctuations in sodium intake and osmotic stress, whereas COX-1 primarily maintains baseline electrolyte excretion [62][123].

COX-2 plays a cytoprotective role in the renal medulla during water deprivation or high-salt intake by mitigating hypertonic stress. COX-2 upregulation in response to hypertonicity drives PGE₂ synthesis, activating nuclear factor- κ B (NF- κ B) and p38 MAPK pathways. This upregulation triggers the transcription of anti-apoptotic genes and hypertonicity-induced cell death [124][125][126][127][128].

3.3. RAAS Modulation and Renin Release by COX

RAAS is a central hormonal cascade that governs renal perfusion, electrolyte balance, and systemic vascular resistance [129][130][131]. COX-derived PGs emerge as critical renin synthesis and secretion modulators, primarily via their autocrine and paracrine actions within the juxtaglomerular apparatus (JGA). The JGA is strategically composed of juxtaglomerular cells (JG cells; aka Granular cells), MD cells, and extraglomerular mesangial cells, acting as a sensor-effector interface for renal autoregulation [132][133][134]. Among the prostanoids, PGE₂ is the primary effector stimulating renin release from the JG cell [135][136]. This effect is predominantly mediated through EP2 and EP4 receptors (both are coupled to Gs-proteins that activate adenylate cyclase), elevate cAMP levels, and thereby enhance renin gene transcription and exocytosis [137][137][137]. Under physiological stress, such as hypotension, volume depletion, or dietary sodium restriction, tubuloglomerular feedback (TGF) signals via MD lead to upregulation of COX-2 expression, increasing local PGE₂ production. This paracrine signal acts directly on juxtaglomerular cells to stimulate renin secretion, contributing to the compensatory activation of RAAS and restoration of circulatory homeostasis [137][138][139][140].

Evidence from COX-2 knockout mice and pharmacologic inhibition studies underscores the indispensable role of COX-2-derived prostaglandins in renin regulation. Deletion or selective blockade of COX-2 leads to marked suppression of renin mRNA and plasma renin activity (PRA), especially during sodium deprivation. This observation reinforces the notion that COX-2 is a rate-limiting factor in renin biosynthesis under stimulated conditions [62][141][142]. EP receptor knockout models have revealed that EP₄-deficient mice exhibit blunted renin responses, impaired TGF signaling, and alterations in sodium handling, highlighting the PGE₂-EP₄ axis as a molecular determinant of JGA function [143][144][145].

4. Nephrotoxicity and Hemodynamic Consequences of COX Inhibition in Renal Disorders

Renal cyclooxygenases (COX-1 and COX-2) continuously produce PGs that maintain glomerular perfusion and fluid-electrolyte balance. In conditions of reduced circulating volume or renal perfusion, PGs (especially PGE₂ and PGI₂) dilate afferent arterioles to sustain RBF and GFR [24] [25]. COX-derived PGs also modulate tubular transport and counteract potent vasoconstrictors. NSAIDs irreversibly or reversibly inhibit COX, thereby suppressing PG synthesis [17][25]. The four key mechanisms by which COX inhibition exacerbates CKD/AKI are impairment of RBF (GFR regulation), loss of vasoconstrictor buffering, osmotic adaptation, and dysregulated RAAS. COX-2-deficient mice with adenine-induced CKD exhibited exacerbated medullary hypoxia and tubular apoptosis [146].

4.1. **RBF and GFR Regulations**

COX-generated prostaglandins play an indispensable role in setting renal vascular tone. Under normal and volume-depleted states, PGI₂ and PGE₂-mediated vasodilation raise RBF and sustain GFR when circulating volume or adequate perfusion is low [24][105][147]. Genetic and pharmacologic COX inhibition removes this compensatory mechanism. In experimental models, selective COX-2 blockade blunts the afferent arteriolar vasodilation commonly induced by vasoconstrictors, causing a decline in GFR [123][148][149]. Clinically, NSAID use acutely reduces RBF and GFR, especially in volume-contracted states [123][150][151]. Thus, by impeding COX-mediated PG vasodilation, NSAIDs can precipitate renal hypoperfusion and acute kidney injury, especially in already compromised kidneys [149][152]. In CKD, interstitial fibrosis and hypoxia are prevalent [153][154]; COX-2-derived PGE₂ preserves perfusion in the medullary thick ascending limb and collecting ducts. This is achieved by stimulating vasodilation via EP₂/EP₄ receptors and activating cytoprotective pathways such as NF-κB and MAPK [104][155].

4.2. COX Enzymes in Renal Homeostasis: Counter-Regulation of Vasoconstrictive Pathways

COX-derived prostaglandins act as endogenous buffers against renal vasoconstrictors. PGE₂ and PGI₂ blunt the vasoconstrictor and antidiuretic effects of angiotensin II (Ang II) on the afferent arteriole. Under normal physiological conditions, PGs oppose Ang II–induced afferent

constriction, thereby maintaining GFR. PGs also mitigate the impact of endothelin-1 and sympathetic catecholamines in the renal microcirculation. Inhibition of COX unopposed this vasoconstriction. Experimental studies show that COX-2 inhibition dramatically potentiates the hypertensive response to Ang II by reducing medullary blood flow and sodium excretion [114][138][156][157]. Blockade of PG synthesis by NSAIDs removes the "safety valve" against vasoconstriction [158][159][160]. COX inhibition shifts the balance toward Ang II– and endothelin-mediated afferent arteriolar constriction and AKI.

4.3. Sodium and Osmotic Regulation: Adaptive Mechanisms

Cyclooxygenase pathways are crucial for renal salt and osmotic balance. PGE₂ normally acts as a counter-regulatory factor when sodium reabsorption is high, inhibiting the NKCC2 cotransporter in the thick ascending limb and reducing water reabsorption in collecting ducts. [161][162]. Clinically, selective COX-2 inhibitors consistently decrease urinary sodium excretion in the first days of use, and NSAID use can cause sodium retention and peripheral edema [120][163][164][165]. These effects increase blood pressure and extracellular volume, which is counterproductive, especially in CKD or heart failure. By impairing medullary PG production, COX inhibition also diminishes renal medullary blood flow, further hampering the kidney's ability to concentrate or dilute urine in response to osmotic stress [120][166]. In short, NSAIDs cause inappropriate sodium and water retention and reduce renal concentrating ability, contributing to hypertension and volume overload in susceptible patients.

4.4. Suppression of RAAS Activity

CKD progression involves maladaptive activation of the RAAS [167]. Although localized RAAS activation remains essential for preserving perfusion pressure, systemic RAAS overactivity contributes to hypertension and glomerular injury. COX activity supports RAAS function, where NSAID therapy inhibits this critical mechanism, blunting renin release and destabilizing the delicate hemodynamic balance in CKD [168]. CKD stage 3–4 patients, NSAID administration led to significant reductions in plasma renin activity, correlating with episodes of hypotension, AKI, and fluid overload. Animal models show that COX-1 deletion abolishes Ang II–induced hypertension, whereas COX-2 blockade heightens it [169][25]. Thus, COX inhibition disrupts homeostatic RAAS signaling and can precipitate maladaptive volume and hemodynamic effects.

4.5. Evidence-Based Assessment of NSAID-Associated Renal Adverse Effects

The mechanistic disruptions above manifest as significant nephrotoxicity in patients with CKD or cardiovascular comorbidities. Observational studies consistently link NSAID exposure to higher AKI and CKD progression risk. A meta-analysis of CKD patients estimated a pooled odds ratio of ~1.6 for NSAID-associated AKI [169]. In a large cohort of 1.98 million adults with normal baseline renal function, NSAID use was associated with a 1.71-fold higher hazard of incident eGFR <60 mL/min/1.73m² and a 1.93-fold greater hazard of \geq 30% eGFR decline, compared to nonusers [170]. Retrospective analyses report that NSAID use roughly doubles

the odds of AKI or new CKD in elderly and multimorbid populations. For instance, one study found that NSAIDs increased AKI risk by 73% in the general population (OR 1.73) and by 58% in those >65 years (OR 1.58). Combined NSAID and RAAS blocker or diuretic therapy multiplies risk; one analysis showed an OR \approx 2.5 for AKI in CKD patients on diuretics/RAAS inhibitors who also used NSAIDs [169][25]. These epidemiologic findings align with clinical observations: NSAIDs are a leading cause of drug-related hospitalizations for AKI, especially in patients with CKD, heart failure, or liver disease. So, Cyclooxygenase inhibition disrupts multiple protective mechanisms of renal homeostasis.

5. Clinical implications of aspirin therapy in post-myocardial infarction patients with chronic kidney disease: a case-based evaluation

CKD is a well-established accelerator of atherosclerosis and vascular calcification, substantially increasing the incidence and severity of cardiovascular disease. Patients with CKD are at markedly elevated risk for acute coronary syndromes, including myocardial infarction (MI), due to persistent endothelial dysfunction, dysregulated lipid metabolism, and systemic inflammation. Consequently, the post-MI period in CKD patients is clinically precarious, characterized by higher rates of recurrent ischemic events, arrhythmias, and sudden cardiac death [171][172][173][174][175]. Aspirin, an irreversible COX inhibitor, remains the cornerstone of antiplatelet therapy for secondary prevention of atherothrombotic events [176][177]. However, in patients with impaired renal function, the altered pharmacokinetics and hemostatic fragility necessitate a reevaluation of its efficacy and safety profile. The dual burden of cardiovascular vulnerability and increased hemorrhagic risk complicates the clinical utility of sustained-release aspirin in this population.

5.1. Efficacy of Aspirin in CKD for Cardiovascular Event Reduction

The antithrombotic efficacy of aspirin in CKD patients post-MI is nuanced and highly dependent on the stage of renal dysfunction [178][179]. The International Polycap Study-3 (TIPS-3) trial demonstrated that aspirin (75 mg daily) significantly reduced the composite outcome of non-fatal MI, non-fatal stroke, or cardiovascular death, yielding a hazard ratio of 0.57 (95% CI, 0.34–0.94) in patients with eGFR <60 ml/min/1.73 m², supporting its role in secondary prevention within moderate-to-severe CKD [180]. Conversely, post hoc analyses of the ALLHAT (Antihypertensive and Lipid-Lowering Treatment to Prevent Heart Attack Trial) trial and similar studies failed to identify significant reductions in all-cause mortality or MI recurrence in patients with eGFR <60 ml/min/1.73 m², suggesting a diminishing benefit of aspirin in advanced renal insufficiency [181]. This bifurcation in therapeutic outcomes underscores the need to stratify aspirin use based on CKD stage and individual cardiovascular risk profiles.

5.2. Hemorrhagic Complications and Safety Considerations in Antiplatelet Therapy

CKD is characterized by a paradoxical hemostatic milieu—hypercoagulability coexisting with intrinsic bleeding tendencies [182][183], primarily driven by uremia-induced platelet

dysfunction [184][185][186], vascular fragility [187][188], and impaired prostaglandinmediated vasoregulation [61][95]. Aspirin, via COX inhibition, exacerbates bleeding risk by suppressing thromboxane A₂ synthesis. Nonetheless, specific observational cohorts have demonstrated no significant increase in major bleeding events across GFR strata, suggesting that when cautiously prescribed, aspirin may maintain a favorable safety margin in secondary prevention [189][190][191]. Notably, low body weight, concurrent anticoagulant use, and uncontrolled hypertension remain critical modifiers of bleeding risk in the CKD-MI subset [192].

The pharmacodynamic response to aspirin is significantly attenuated in CKD due to platelet hyperreactivity and residual thromboxane generation, a phenomenon termed "Aspirin Resistance" [193][194]. This resistance is mechanistically linked to oxidative stress, enhanced platelet turnover, and inflammation-driven non-COX thromboxane synthesis [195][196][197]. Studies using serum thromboxane B₂ assays [198][199] and platelet aggregometry [200][201] have documented persistently elevated platelet activity despite aspirin therapy in CKD, correlating with higher rates of recurrent ischemic events [202][203][204]. These findings reinforce the hypothesis that standard-dose aspirin may be insufficient for effective platelet inhibition in CKD, particularly in sustained-release formulations where bioavailability may be variable.

5.3. Clinical Recommendations

Despite its therapeutic value, aspirin remains the recommended first-line agent for secondary prevention in post-MI CKD patients, conditional upon stringent bleeding surveillance and appropriate dose titration [205][206][207]. Data from coronary care registries indicate a sharp decline in the use of both aspirin and β -blockers as renal function deteriorates. Among patients with creatinine clearance (CrCl) <46.2 ml/min, only 35% received both agents, in contrast to 63.9% among patients with higher CrCl values [108,[209][210]. Clinical variables such as concurrent heart failure, arrhythmias, and hemodynamic instability at admission contributed to therapeutic omission. The role of aspirin in primary prevention is increasingly contested, especially in those with advanced CKD or low thrombotic risk, where the marginal benefit may not outweigh the elevated bleeding potential [207][211]. Ongoing large-scale randomized controlled trials such as ATTACK (Aspirin To Target Arterial Events in CKD) are expected to provide more granular evidence on the net clinical benefit of aspirin stratified by CKD stage and comorbid burden [212]. In the interim, clinical decision-making should prioritize personalized therapy, integrating metrics such as eGFR, hemoglobin levels, platelet function assays, and bleeding risk scores.

6. Current Guidelines and NSAID Management in CKD

NSAIDs remain a cornerstone in the management of pain and inflammation; however, their utility in patients with CKD is met with substantial caution due to their well-documented nephrotoxic potential. By suppressing the COX enzyme (both COX-1 and COX-2) and prostaglandin-mediated vasodilation, NSAIDs compromise renal hemodynamics, particularly

in the afferent arterioles, leading to diminished GFR [13][17][24][97][11]. Epidemiological data underscore these concerns. Longitudinal studies and pooled meta-analyses have identified a statistically significant association between chronic NSAID use and CKD progression. Contemporary clinical guidelines from nephrology and pharmacology societies-including the Kidney Disease: Improving Global Outcomes (KDIGO) and European Renal Best Practice (ERBP)-advocate for stringent restriction of NSAID use in patients with impaired renal function [213][214][169][215]. NSAID therapy is generally contraindicated in individuals with eGFR below 60 mL/min/1.73 m², where renal autoregulation becomes increasingly dependent on prostaglandin-mediated vasodilation [169][123][23]. In stages 3-5 CKD, where structural and functional nephron loss is pronounced, the use of NSAIDs-both selective (COX-2 inhibitors) and non-selective-is discouraged due to heightened risk of glomerular hypoperfusion, sodium retention, hyperkalemia, and acute-on-chronic kidney injury. Particularly in advanced CKD (eGFR <30 mL/min/1.73 m²) and dialysis-dependent patients, NSAIDs are deemed unsafe. [148][170][216]. In scenarios where NSAID therapy is deemed unavoidable, such as refractory inflammatory conditions, guidelines recommend adopting a minimal effective dose strategy for the shortest possible duration. Topical NSAIDs (such as topical diclofenac 1% gel) are often preferred in CKD patients with localized musculoskeletal pain due to reduced systemic absorption and lower nephrotoxicity risk [217][218][219]. Overthe-counter (OTC) NSAID consumption, particularly in CKD patients unaware of their diagnosis or risks, remains a critical concern.

| Topical Analgesics | Common Pain Conditions | Comments | | | |
|-----------------------|-----------------------------------|---------------------------------------|--|--|--|
| (Formulations) | Tested | | | | |
| Diclofenac (1% gel) | Minor sports injuries, acute | Topical NSAIDs (especially | | | |
| | ankle sprains, knee | diclofenac and ibuprofen) are more | | | |
| | osteoarthritis, and chronic | extensively studied, providing | | | |
| | lateral epicondylitis | short-term relief for acute injuries | | | |
| | | and chronic joint conditions like | | | |
| | | osteoarthritis. | | | |
| Ibuprofen (5% | Chronic knee pain, chronic leg | Effective for managing chronic | | | |
| cream or gel) | ulcers, soft tissue injuries, and | conditions, especially knee pain | | | |
| | acute ankle sprains | and soft tissue injuries. | | | |
| Ketoprofen (2.5% | Soft tissue injuries | Used for relief in soft tissue | | | |
| gel, total daily dose | | injuries. | | | |
| of 250 mg) | | | | | |
| Salicylates (750 mg | Acute and postherpetic | The combination of aspirin and | | | |
| aspirin + diethyl | neuralgia | diethyl ether offers effective relief | | | |
| ether mixture or 75 | | for neuralgic pain. | | | |
| mg/mL of aspirin | | | | | |
| alone) | | | | | |

| Table 3: | : Topical | Analgesics | for I | Managing A | cute and | Chronic | Pain | [218] |
|----------|-----------|------------|-------|------------|----------|---------|------|-------|
|----------|-----------|------------|-------|------------|----------|---------|------|-------|

| Lidocaine (5% | Postherpetic neuralgia, diabetic | Data suggests superior pain control | | | |
|--|----------------------------------|-------------------------------------|--|--|--|
| medicated patch or | neuropathy | for postherpetic neuralgia | | | |
| plaster) | | compared to oral pregabalin. | | | |
| Capsaicin (0.025– | Neuropathic pain, postherpetic | Weak evidence, but has shown | | | |
| 0.075% cream, 8% | neuralgia, acute migraine | improvement in neuropathic pain | | | |
| patch) | | and postherpetic neuralgia. | | | |
| Amitriptyline (1– | Neuropathic pain | Used for neuropathic pain | | | |
| 5% cream) | | management. | | | |
| Glyceryl trinitrate | Lateral epicondylitis, chronic | Poor data, but has reported | | | |
| (0.72 mg/day) | noninsertional Achilles | improvement in wound healing. | | | |
| | tendinopathy, post- | | | | |
| | hemorrhoidectomy | | | | |
| Others (opioids, | Chronic knee pain, vulvar | Scant data, but used for various | | | |
| menthol, | lichen scleroatrophicus, | chronic pain conditions. | | | |
| pimecrolimus, superficial burns, chronic leg | | | | | |
| phenytoin) | ulcers | | | | |

6.1. NSAID Interactions and Comorbidities in CKD

The synergistic nephrotoxicity occurs when NSAIDs are co-administered with other medications frequently prescribed in CKD, such as RAAS inhibitors, diuretics [220][129], and calcineurin inhibitors [221][222], resulting in hemodynamic instability and electrolyte imbalances. The simultaneous use of an ACE inhibitor or ARB, a loop or thiazide diuretic, and an NSAID (so-called triple whammy interaction) has been consistently implicated in precipitating AKI [222][223][224]. Furthermore, NSAID-induced sodium retention can exacerbate hypertension, congestive heart failure, and volume overload, all of which are common comorbidities in the CKD population [225][226][227]. In patients with hepatorenal syndrome, cirrhosis, or right-sided heart failure, renal perfusion is highly prostaglandin-dependent. NSAIDs can provoke sudden declines in GFR, precipitating hepatorenal decompensation or exacerbating fluid retention and pulmonary edema [228][229][230]. So, appropriate surveillance and proactive education are critical in mitigating NSAID-associated nephrotoxicity in CKD populations.

7. Discussion and Conclusion

The intersection of chronic kidney disease and nonsteroidal anti-inflammatory drug pharmacology presents a critical clinical dilemma underscored by intricate biochemical, pharmacokinetic, and pathophysiological interdependencies. The widespread utilization of NSAIDs, often without medical supervision, aggravates renal vulnerability, especially in patients with underlying renal dysfunction or comorbidities, such as hypertension and cardiac issues. The nephrotoxic potential of NSAIDs arises primarily from their inhibition of cyclooxygenase enzymes, which are instrumental in the biosynthesis of vasodilatory prostaglandins critical for maintaining renal perfusion. Under normal physiological conditions,

these prostaglandins exert a counter-regulatory effect on vasoconstrictive agents (like Ang II), thereby preserving the glomerular filtration rate. However, in CKD patients, where renal autoregulation is already compromised, NSAID-induced suppression of prostaglandin synthesis severely impairs afferent arteriolar dilation, precipitating acute kidney injury, accelerating fibrotic remodeling, and facilitating a downward trajectory toward end-stage renal disease. The renal expression of COX-2 in the macula densa and glomerular podocytes suggests that selective COX-2 inhibitors are not devoid of renal side effects and may similarly disrupt natriuretic and vasodilatory pathways. From a clinical perspective, the indiscriminate use of NSAIDs in CKD remains a pressing concern, often exacerbated by over-the-counter availability and inadequate risk communication. The evidence collectively advocates for heightened pharmacovigilance, therapeutic individualization, and robust patient education. Regular renal function monitoring, risk stratification based on CKD staging, and exploring alternative analgesic regimens—such as acetaminophen, topical agents, or low-dose opioids—are prudent strategies to mitigate iatrogenic renal injury.

In conclusion, NSAID use in CKD embodies a paradigmatic example of risk amplification through pathophysiological synergy. The interplay between impaired renal excretion, disrupted prostaglandin-mediated hemodynamics, and systemic drug accumulation renders NSAID therapy a double-edged sword in this population. In patients with chronic kidney disease, NSAID use represents a precarious therapeutic choice where the compounded risk of nephrotoxicity often outweighs analgesic benefit.

Acknowledgements

The authors sincerely acknowledge the academic and institutional support received during the development of this review. While no individuals or organizations were directly involved in the writing or editing process, the authors are grateful for access to scientific literature and open-access databases that facilitated this evidence-based synthesis.

Funding

The authors declare that this review article did not receive specific grants or financial support from public, institutional, commercial, or not-for-profit funding agencies.

Ethical Declaration

The authors declare that the use of Artificial Intelligence (AI) tools in the preparation of this manuscript adhered strictly to the ethical standards outlined by the Committee on Publication Ethics (COPE) and the International Committee of Medical Journal Editors (ICMJE). AI technologies were employed solely for improving language clarity, grammar, and structural coherence, without contributing to generating novel scientific content, data fabrication, or citation manipulation. Conceptualization, analysis, and interpretations presented in this review represent the original intellectual contributions of the authors. The manuscript was critically

reviewed and finalized by the authors to ensure integrity, transparency, and accountability by international ethical publishing standards.

References

- P. Romagnani et al., "Chronic kidney disease.," Nat. Rev. Dis. Prim., vol. 3, p. 17088, Nov. 2017, doi: 10.1038/nrdp.2017.88.
- [2] C. Charles and A. H. Ferris, "Chronic Kidney Disease," Prim. Care Clin. Off. Pract., vol. 47, no. 4, pp. 585–595, 2020, doi: https://doi.org/10.1016/j.pop.2020.08.001.
- [3] T. K. Chen, D. H. Knicely, and M. E. Grams, "Chronic Kidney Disease Diagnosis and Management: A Review.," JAMA, vol. 322, no. 13, pp. 1294–1304, Oct. 2019, doi: 10.1001/jama.2019.14745.
- [4] V. Jha et al., "Chronic kidney disease: global dimension and perspectives.," Lancet (London, England), vol. 382, no. 9888, pp. 260–272, Jul. 2013, doi: 10.1016/S0140-6736(13)60687-X.
- [5] K.-U. Eckardt et al., "Trends and perspectives for improving quality of chronic kidney disease care: conclusions from a Kidney Disease: Improving Global Outcomes (KDIGO) Controversies Conference," Kidney Int., vol. 104, no. 5, pp. 888–903, Nov. 2023, doi: 10.1016/j.kint.2023.05.013.
- [6] K. Xie et al., "Global, regional, and national burden of chronic kidney disease, 1990-2021: a systematic analysis for the global burden of disease study 2021.," Front. Endocrinol. (Lausanne)., vol. 16, p. 1526482, 2025, doi: 10.3389/fendo.2025.1526482.
- [7] C. P. Kovesdy, "Epidemiology of chronic kidney disease: an update 2022.," Kidney Int. Suppl., vol. 12, no. 1, pp. 7–11, Apr. 2022, doi: 10.1016/j.kisu.2021.11.003.
- [8] S. R. Vaidya and N. R. Aeddula, "Chronic Kidney Disease.," Treasure Island (FL), 2025.
- [9] S. Altamura, D. Pietropaoli, F. Lombardi, R. Del Pinto, and C. Ferri, "An Overview of Chronic Kidney Disease Pathophysiology: The Impact of Gut Dysbiosis and Oral Disease.," Biomedicines, vol. 11, no. 11, Nov. 2023, doi: 10.3390/biomedicines11113033.
- [10] J. M. López-Novoa, C. Martínez-Salgado, A. B. Rodríguez-Peña, and F. J. López-Hernández, "Common pathophysiological mechanisms of chronic kidney disease: therapeutic perspectives.," Pharmacol. Ther., vol. 128, no. 1, pp. 61–81, Oct. 2010, doi: 10.1016/j.pharmthera.2010.05.006.
- [11] A. Džidić-Krivić, E. K. Sher, J. Kusturica, E. K. Farhat, A. Nawaz, and F. Sher, "Unveiling drug induced nephrotoxicity using novel biomarkers and cutting-edge preventive strategies.," Chem. Biol. Interact., vol. 388, p. 110838, Jan. 2024, doi: 10.1016/j.cbi.2023.110838.
- [12] A. Abdulla et al., "Guidance on the management of pain in older people.," Age Ageing, vol. 42 Suppl 1, pp. i1-57, Mar. 2013, doi: 10.1093/ageing/afs200.
- [13] I. Ghlichloo and V. Gerriets, "Nonsteroidal Anti-Inflammatory Drugs (NSAIDs).," Treasure Island (FL), 2025.
- [14] W. J. Phillips and B. L. Currier, "Analgesic pharmacology: II. Specific analgesics.," J. Am. Acad. Orthop. Surg., vol. 12, no. 4, pp. 221–233, 2004, doi: 10.5435/00124635-200407000-00003.

- [15] J. R. Vane and R. M. Botting, "Mechanism of action of nonsteroidal anti-inflammatory drugs.," Am. J. Med., vol. 104, no. 3A, pp. 2S-8S; discussion 21S-22S, Mar. 1998, doi: 10.1016/s0002-9343(97)00203-9.
- [16] J. R. Vane, "Inhibition of prostaglandin synthesis as a mechanism of action for aspirinlike drugs.," Nat. New Biol., vol. 231, no. 25, pp. 232–235, Jun. 1971, doi: 10.1038/newbio231232a0.
- [17] O. Qureshi and A. Dua, "COX Inhibitors.," Treasure Island (FL), 2025.
- [18] C. A. Rouzer and L. J. Marnett, "Cyclooxygenases: Structural and functional insights," J. Lipid Res., vol. 50, no. SUPPL., pp. 29–34, 2009, doi: 10.1194/jlr.R800042-JLR200.
- [19] J. A. Mitchell et al., "Cyclooxygenases and the cardiovascular system.," Pharmacol. Ther., vol. 217, p. 107624, Jan. 2021, doi: 10.1016/j.pharmthera.2020.107624.
- [20] K. Malik and A. Dua, "Prostaglandins.," Treasure Island (FL), 2025.
- [21] E. J. Pharmaceutical, V. Kushwaha, P. Agrawal, H. Vekaria, and A. Das, "PROSTAGLANDINS : AN OVERVIEW," no. December, 2023.
- [22] M. Murakami, Y. Nakatani, T. Tanioka, and I. Kudo, "Prostaglandin E synthase.," Prostaglandins Other Lipid Mediat., vol. 68–69, pp. 383–399, Aug. 2002, doi: 10.1016/s0090-6980(02)00043-6.
- [23] G. N. C. Lucas, A. C. C. Leitão, R. L. Alencar, R. M. F. Xavier, E. D. F. Daher, and G. B. da Silva Junior, "Pathophysiological aspects of nephropathy caused by non-steroidal anti-inflammatory drugs.," J. Bras. Nefrol., vol. 41, no. 1, pp. 124–130, 2019, doi: 10.1590/2175-8239-JBN-2018-0107.
- [24] G.-H. Kim, "Renal effects of prostaglandins and cyclooxygenase-2 inhibitors.," Electrolyte Blood Press., vol. 6, no. 1, pp. 35–41, Jun. 2008, doi: 10.5049/EBP.2008.6.1.35.
- [25] S. Drożdżal et al., "Kidney damage from nonsteroidal anti-inflammatory drugs-Myth or truth? Review of selected literature.," Pharmacol. Res. Perspect., vol. 9, no. 4, p. e00817, Aug. 2021, doi: 10.1002/prp2.817.
- [26] S. Shimada, C. Yang, V. Kumar, D. L. Mattson, and A. W. Cowley, "Acute Increase of Renal Perfusion Pressure Causes Rapid Activation of mTORC1 (Mechanistic Target Of Rapamycin Complex 1) and Leukocyte Infiltration," Hypertension, vol. 79, no. 6, pp. 1180–1189, Jun. 2022, doi: 10.1161/HYPERTENSIONAHA.121.18643.
- [27] W. B. White, "Cardiovascular Effects of the Cyclooxygenase Inhibitors," Hypertension, vol. 49, no. 3, pp. 408–418, Mar. 2007, doi: 10.1161/01.HYP.0000258106.74139.25.
- [28] Y. Faki and A. Er, "Different chemical structures and physiological/pathological roles of cyclooxygenases," Rambam Maimonides Med. J., vol. 12, no. 1, pp. 1–13, 2021, doi: 10.5041/RMMJ.10426.
- [29] W. L. Smith, D. L. DeWitt, and R. M. Garavito, "Cyclooxygenases: Structural, Cellular, and Molecular Biology," Annu. Rev. Biochem., vol. 69, no. Volume 69, 2000, pp. 145– 182, 2000, doi: https://doi.org/10.1146/annurev.biochem.69.1.145.
- [30] M. Miciaccia et al., "Three-dimensional structure of human cyclooxygenase (hCOX)-1," Sci. Rep., vol. 11, no. 1, p. 4312, 2021, doi: 10.1038/s41598-021-83438-z.
- [31] F. A. Fitzpatrick, "Cyclooxygenase enzymes: regulation and function," Curr. Pharm. Des., vol. 10, no. 6, p. 577—588, 2004, doi: 10.2174/1381612043453144.

- [32] R. Nørregaard, T. H. Kwon, and J. Frøkiær, "Physiology and pathophysiology of cyclooxygenase-2 and prostaglandin E2 in the kidney," Kidney Res. Clin. Pract., vol. 34, no. 4, pp. 194–200, 2015, doi: 10.1016/j.krcp.2015.10.004.
- [33] R. C. Harris, "Cyclooxygenase-2 inhibition and renal physiology," Am. J. Cardiol., vol. 89, no. 6 SUPPL. 1, pp. 10–17, 2002, doi: 10.1016/S0002-9149(02)02232-4.
- [34] W. K. Ju and A. H. Neufeld, "Cellular localization of cyclooxygenase-1 and cyclooxygenase-2 in the normal mouse, rat, and human retina," J. Comp. Neurol., vol. 452, no. 4, pp. 392–399, 2002, doi: 10.1002/cne.10400.
- [35] N. Chandrasekharan and D. L. Simmons, "Protein family review The Smads," Genome Biol., vol. 5, no. 9, p. 241, 2004.
- [36] D. Picot, P. J. Loll, and R. M. Garavito, "The X-ray crystal structure of the membrane protein prostaglandin H2 synthase-1," Nature, vol. 367, no. 6460, pp. 243–249, 1994, doi: 10.1038/367243a0.
- [37] C. Ma, Y. Tsukamoto, and H. Takeuchi, "Generation of Properly Folded Epidermal Growth Factor-Like (EGF) RepeatsEpidermal growth factor-like (EGF) repeats and GlycosyltransferasesGlycosyltransferases Enables In Vitro O-GlycosylationOglycosylation," in Notch Signaling Research: Methods and Protocols, D. Jia, Ed., New York, NY: Springer US, 2022, pp. 27–38. doi: 10.1007/978-1-0716-2201-8_3.
- [38] B. Bersch, J. F. Hernandez, D. Marion, and G. J. Arlaud, "Solution structure of the epidermal growth factor (EGF)-like module of human complement protease C1r, an atypical member of the EGF family," Biochemistry, vol. 37, no. 5, pp. 1204–1214, 1998, doi: 10.1021/bi971851v.
- [39] E. Kovacs, J. A. Zorn, Y. Huang, T. Barros, and J. Kuriyan, "A structural perspective on the regulation of the epidermal growth factor receptor.," Annu. Rev. Biochem., vol. 84, pp. 739–764, 2015, doi: 10.1146/annurev-biochem-060614-034402.
- [40] C. A. Rouzer and L. J. Marnett, "Structural and Chemical Biology of the Interaction of Cyclooxygenase with Substrates and Non-Steroidal Anti-Inflammatory Drugs.," Chem. Rev., vol. 120, no. 15, pp. 7592–7641, Aug. 2020, doi: 10.1021/acs.chemrev.0c00215.
- [41] J. Dong et al., "The membrane-anchoring domain of epidermal growth factor receptor ligands dictates their ability to operate in juxtacrine mode.," Mol. Biol. Cell, vol. 16, no. 6, pp. 2984–2998, Jun. 2005, doi: 10.1091/mbc.e04-11-0994.
- [42] M. N. Romanelli, "Chapter 6.1 Cyclooxygenase," in Metalloenzymes, C. T. Supuran and W. A. Donald, Eds., Academic Press, 2024, pp. 431–447. doi: https://doi.org/10.1016/B978-0-12-823974-2.00010-3.
- [43] C. Pasero and M. McCaffery, "Selective COX-2 inhibitors," Am. J. Nurs., vol. 101, no. 4, pp. 655–683, 2001, doi: 10.1097/00000446-200104000-00024.
- [44] U. R. Mbonye et al., "Two Distinct Pathways for Cyclooxygenase-2 Protein Degradation *," J. Biol. Chem., vol. 283, no. 13, pp. 8611–8623, Mar. 2008, doi: 10.1074/jbc.M710137200.
- [45] R. M. Garavito, M. G. Malkowski, and D. L. DeWitt, "The structures of prostaglandin endoperoxide H synthases-1 and -2.," Prostaglandins Other Lipid Mediat., vol. 68–69, pp. 129–152, Aug. 2002, doi: 10.1016/s0090-6980(02)00026-6.

- [46] I. Song, "The Structural Environment Around the Heme-Binding Site in Cyclooxygenase and Its Implication," Biophys. J., vol. 120, no. 3, p. 119a, Feb. 2021, doi: 10.1016/j.bpj.2020.11.933.
- [47] M. Ahmadi, S. Bekeschus, K.-D. Weltmann, T. von Woedtke, and K. Wende, "Nonsteroidal anti-inflammatory drugs: recent advances in the use of synthetic COX-2 inhibitors.," RSC Med. Chem., vol. 13, no. 5, pp. 471–496, May 2022, doi: 10.1039/d1md00280e.
- [48] M. M. Mitchener et al., "Competition and allostery govern substrate selectivity of cyclooxygenase-2.," Proc. Natl. Acad. Sci. U. S. A., vol. 112, no. 40, pp. 12366–12371, Oct. 2015, doi: 10.1073/pnas.1507307112.
- [49] N. V. Bhagavan and C.-E. Ha, "Enzymes and Enzyme Regulation," Essentials Med. Biochem., pp. 63–84, 2015, doi: 10.1016/b978-0-12-416687-5.00006-3.
- [50] Y. Li et al., "Oligomeric interactions maintain active-site structure in a noncooperative enzyme family.," EMBO J., vol. 41, no. 17, p. e108368, Sep. 2022, doi: 10.15252/embj.2021108368.
- [51] C. Kropp, A. Bruckmann, and P. Babinger, "Controlling Enzymatic Activity by Modulating the Oligomerization State via Chemical Rescue and Optical Control.," Chembiochem, vol. 23, no. 5, p. e202100490, Mar. 2022, doi: 10.1002/cbic.202100490.
- [52] S. N. Kudalkar, C. A. Rouzer, and L. J. Marnett, "The Peroxidase and Cyclooxygenase Activity of Prostaglandin H Synthase," in Heme Peroxidases, The Royal Society of Chemistry, 2015. doi: 10.1039/9781782622628-00245.
- [53] A. J. Vecchio, B. J. Orlando, R. Nandagiri, and M. G. Malkowski, "Investigating substrate promiscuity in cyclooxygenase-2 the role of Arg-120 and residues lining the hydrophobic groove," J. Biol. Chem., vol. 287, no. 29, pp. 24619–24630, 2012, doi: 10.1074/jbc.M112.372243.
- [54] H. Zou et al., "Human cyclooxygenase-1 activity and its responses to COX inhibitors are allosterically regulated by nonsubstrate fatty acids.," J. Lipid Res., vol. 53, no. 7, pp. 1336–1347, Jul. 2012, doi: 10.1194/jlr.M026856.
- [55] L. Dong, A. J. Vecchio, N. P. Sharma, B. J. Jurban, M. G. Malkowski, and W. L. Smith, "Human cyclooxygenase-2 is a sequence homodimer that functions as a conformational heterodimer," J. Biol. Chem., vol. 286, no. 21, pp. 19035–19046, 2011, doi: 10.1074/jbc.M111.231969.
- [56] N. Zidar, K. Odar, D. Glavac, M. Jerse, T. Zupanc, and D. Stajer, "Cyclooxygenase in normal human tissues--is COX-1 really a constitutive isoform, and COX-2 an inducible isoform?," J. Cell. Mol. Med., vol. 13, no. 9B, pp. 3753–3763, Sep. 2009, doi: 10.1111/j.1582-4934.2008.00430.x.
- [57] R. M. Garavito and A. M. Mulichak, "The structure of mammalian cyclooxygenases.," Annu. Rev. Biophys. Biomol. Struct., vol. 32, pp. 183–206, 2003, doi: 10.1146/annurev.biophys.32.110601.141906.
- [58] W. L. Smith and D. L. Dewitt, "Prostaglandin endoperoxide H synthases-1 and -2.," Adv. Immunol., vol. 62, pp. 167–215, 1996, doi: 10.1016/s0065-2776(08)60430-7.
- [59] A. Alexanian and A. Sorokin, "Cyclooxygenase 2: protein-protein interactions and posttranslational modifications.," Physiol. Genomics, vol. 49, no. 11, pp. 667–681, Nov. 2017, doi: 10.1152/physiolgenomics.00086.2017.

- [60] Y. Faki and A. Er, "Different Chemical Structures and Physiological/Pathological Roles of Cyclooxygenases.," Rambam Maimonides Med. J., vol. 12, no. 1, Jan. 2021, doi: 10.5041/RMMJ.10426.
- [61] M. D. Breyer and R. C. Harris, "Cyclooxygenase 2 and the kidney," Curr. Opin. Nephrol. Hypertens., vol. 10, no. 1, p. 89—98, Jan. 2001, doi: 10.1097/00041552-200101000-00014.
- [62] H.-F. Cheng and R. C. Harris, "Cyclooxygenases, the Kidney, and Hypertension," Hypertension, vol. 43, no. 3, pp. 525–530, Mar. 2004, doi: 10.1161/01.HYP.0000116221.27079.ea.
- [63] R. Nørregaard, T.-H. Kwon, and J. Frøkiær, "Physiology and pathophysiology of cyclooxygenase-2 and prostaglandin E2 in the kidney.," Kidney Res. Clin. Pract., vol. 34, no. 4, pp. 194–200, Dec. 2015, doi: 10.1016/j.krcp.2015.10.004.
- [64] M. López-Parra et al., "Cyclooxygenase-1 derived prostaglandins are involved in the maintenance of renal function in rats with cirrhosis and ascites.," Br. J. Pharmacol., vol. 135, no. 4, pp. 891–900, Feb. 2002, doi: 10.1038/sj.bjp.0704528.
- [65] K. Tsuge, T. Inazumi, A. Shimamoto, and Y. Sugimoto, "Molecular mechanisms underlying prostaglandin E2-exacerbated inflammation and immune diseases.," Int. Immunol., vol. 31, no. 9, pp. 597–606, Aug. 2019, doi: 10.1093/intimm/dxz021.
- [66] Y. Jang, M. Kim, and S. W. Hwang, "Molecular mechanisms underlying the actions of arachidonic acid-derived prostaglandins on peripheral nociception," J. Neuroinflammation, vol. 17, no. 1, p. 30, 2020, doi: 10.1186/s12974-020-1703-1.
- [67] S. A. Martin, A. R. Brash, and R. C. Murphy, "The discovery and early structural studies of arachidonic acid," J. Lipid Res., vol. 57, no. 7, pp. 1126–1132, Jul. 2016, doi: 10.1194/jlr.R068072.
- [68] M. Murakami and I. Kudo, "Phospholipase A2," J. Biochem., vol. 131, no. 3, pp. 285–292, Mar. 2002, doi: 10.1093/oxfordjournals.jbchem.a003101.
- [69] Y. Zhang, Y. Liu, J. Sun, W. Zhang, Z. Guo, and Q. Ma, "Arachidonic acid metabolism in health and disease.," MedComm, vol. 4, no. 5, p. e363, Oct. 2023, doi: 10.1002/mco2.363.
- [70] B. Wang et al., "Metabolism pathways of arachidonic acids: mechanisms and potential therapeutic targets," Signal Transduct. Target. Ther., vol. 6, no. 1, p. 94, 2021, doi: 10.1038/s41392-020-00443-w.
- [71] A. A. Hajeyah, W. J. Griffiths, Y. Wang, A. J. Finch, and V. B. O'Donnell, "The Biosynthesis of Enzymatically Oxidized Lipids," Front. Endocrinol. (Lausanne)., vol. 11, no. November, pp. 1–32, 2020, doi: 10.3389/fendo.2020.591819.
- [72] P. Needleman, "Characterization of the reaction sequence involved in phospholipid labeling and deacylation and prostaglandin synthesis and actions," J. Allergy Clin. Immunol., vol. 62, no. 2, pp. 96–102, 1978, doi: https://doi.org/10.1016/0091-6749(78)90085-4.
- [73] A. Zarghi and S. Arfaei, "Selective COX-2 Inhibitors: A Review of Their Structure-Activity Relationships.," Iran. J. Pharm. Res. IJPR, vol. 10, no. 4, pp. 655–683, 2011.
- [74] J. Liu, S. A. Seibold, C. J. Rieke, I. Song, R. I. Cukier, and W. L. Smith, "Prostaglandin Endoperoxide H Synthases: PEROXIDASE HYDROPEROXIDE SPECIFICITY AND

CYCLOOXYGENASE ACTIVATION*," J. Biol. Chem., vol. 282, no. 25, pp. 18233–18244, 2007, doi: https://doi.org/10.1074/jbc.M701235200.

- [75] I. Kudo and M. Murakami, "Prostaglandin E synthase, a terminal enzyme for prostaglandin E2 biosynthesis," J. Biochem. Mol. Biol., vol. 38, no. 6, pp. 633–638, 2005, doi: 10.5483/bmbrep.2005.38.6.633.
- [76] B. Samuelsson, R. Morgenstern, and P.-J. Jakobsson, "Membrane prostaglandin E synthase-1: a novel therapeutic target.," Pharmacol. Rev., vol. 59, no. 3, pp. 207–224, Sep. 2007, doi: 10.1124/pr.59.3.1.
- [77] K. Lee, S. H. Lee, and T. H. Kim, "The Biology of Prostaglandins and Their Role as a Target for Allergic Airway Disease Therapy.," Int. J. Mol. Sci., vol. 21, no. 5, Mar. 2020, doi: 10.3390/ijms21051851.
- [78] S. Rittchen and A. Heinemann, "Therapeutic Potential of Hematopoietic Prostaglandin D(2) Synthase in Allergic Inflammation.," Cells, vol. 8, no. 6, Jun. 2019, doi: 10.3390/cells8060619.
- [79] R. Pettipher, "The roles of the prostaglandin D 2 receptors DP 1 and CRTH2 in promoting allergic responses," Br. J. Pharmacol., vol. 153, no. SUPPL. 1, pp. 191–199, 2008, doi: 10.1038/sj.bjp.0707488.
- [80] Y. Urade, "Biochemical and Structural Characteristics, Gene Regulation, Physiological, Pathological and Clinical Features of Lipocalin-Type Prostaglandin D(2) Synthase as a Multifunctional Lipocalin.," Front. Physiol., vol. 12, p. 718002, 2021, doi: 10.3389/fphys.2021.718002.
- [81] A. S. Ahmad, H. Ottallah, C. B. Maciel, M. Strickland, and S. Doré, "Role of the L-PGDS-PGD2-DP1 receptor axis in sleep regulation and neurologic outcomes.," Sleep, vol. 42, no. 6, Jun. 2019, doi: 10.1093/sleep/zsz073.
- [82] Y. Cao, Y. Guan, Y.-Y. Xu, and C.-M. Hao, "Endothelial prostacyclin protects the kidney from ischemia-reperfusion injury," Pflügers Arch. - Eur. J. Physiol., vol. 471, no. 4, pp. 543–555, 2019, doi: 10.1007/s00424-018-2229-6.
- [83] I. M. Lang and S. P. Gaine, "Recent advances in targeting the prostacyclin pathway in pulmonary arterial hypertension.," Eur. Respir. Rev. an Off. J. Eur. Respir. Soc., vol. 24, no. 138, pp. 630–641, Dec. 2015, doi: 10.1183/16000617.0067-2015.
- [84] J. V Bonventre and M. Swidler, "Calcium dependency of prostaglandin E2 production in rat glomerular mesangial cells. Evidence that protein kinase C modulates the Ca2+dependent activation of phospholipase A2.," J. Clin. Invest., vol. 82, no. 1, pp. 168–176, Jul. 1988, doi: 10.1172/JCI113566.
- [85] H. Juan, "Role of calcium in prostaglandin E release induced by bradykinin and the ionophore A 23187," Naunyn. Schmiedebergs. Arch. Pharmacol., vol. 307, no. 2, pp. 177–183, 1979, doi: 10.1007/BF00498461.
- [86] B. B. Boyanovsky and N. R. Webb, "Biology of Secretory Phospholipase A2," Cardiovasc. Drugs Ther., vol. 23, no. 1, pp. 61–72, 2009, doi: 10.1007/s10557-008-6134-7.
- [87] M. Triggiani, F. Granata, G. Giannattasio, and G. Marone, "Secretory phospholipases A2 in inflammatory and allergic diseases: not just enzymes.," J. Allergy Clin. Immunol., vol. 116, no. 5, pp. 1000–1006, Nov. 2005, doi: 10.1016/j.jaci.2005.08.011.

- [88] S. Ramanadham, T. Ali, J. W. Ashley, R. N. Bone, W. D. Hancock, and X. Lei, "Calciumindependent phospholipases A2 and their roles in biological processes and diseases.," J. Lipid Res., vol. 56, no. 9, pp. 1643–1668, Sep. 2015, doi: 10.1194/jlr.R058701.
- [89] X. Lei, S. E. Barbour, and S. Ramanadham, "Group VIA Ca2+-independent phospholipase A2 (iPLA2beta) and its role in beta-cell programmed cell death.," Biochimie, vol. 92, no. 6, pp. 627–637, Jun. 2010, doi: 10.1016/j.biochi.2010.01.005.
- [90] L. Zhang, B. L. Peterson, and B. S. Cummings, "The effect of inhibition of Ca2+independent phospholipase A2 on chemotherapeutic-induced death and phospholipid profiles in renal cells.," Biochem. Pharmacol., vol. 70, no. 11, pp. 1697–1706, Nov. 2005, doi: 10.1016/j.bcp.2005.09.008.
- [91] W. He et al., "Increased dietary sodium induces COX2 expression by activating NFκB in renal medullary interstitial cells.," Pflugers Arch., vol. 466, no. 2, pp. 357–367, Feb. 2014, doi: 10.1007/s00424-013-1328-7.
- [92] H. Shi et al., "Cadmium induces epithelial-mesenchymal transition and migration of renal cancer cells by increasing PGE2 through a cAMP/PKA-COX2 dependent mechanism," Ecotoxicol. Environ. Saf., vol. 207, p. 111480, 2021, doi: 10.1016/j.ecoenv.2020.111480.
- [93] R. D. Rudic et al., "COX-2-derived prostacyclin modulates vascular remodeling.," Circ. Res., vol. 96, no. 12, pp. 1240–1247, Jun. 2005, doi: 10.1161/01.RES.0000170888.11669.28.
- [94] F. Electrolyte and R. Physiol, "Prostaglandins," no. 168, 2025.
- [95] Y. Li et al., "Prostaglandins in the pathogenesis of kidney diseases.," Oncotarget, vol. 9, no. 41, pp. 26586–26602, May 2018, doi: 10.18632/oncotarget.25005.
- [96] L. G. Navar, E. W. Inscho, S. A. Majid, J. D. Imig, L. M. Harrison-Bernard, and K. D. Mitchell, "Paracrine regulation of the renal microcirculation.," Physiol. Rev., vol. 76, no. 2, pp. 425–536, Apr. 1996, doi: 10.1152/physrev.1996.76.2.425.
- [97] N. S. Kirkby et al., "Cyclooxygenase-2 Selectively Controls Renal Blood Flow Through a Novel PPARβ/δ-Dependent Vasodilator Pathway.," Hypertens. (Dallas, Tex. 1979), vol. 71, no. 2, pp. 297–305, Feb. 2018, doi: 10.1161/HYPERTENSIONAHA.117.09906.
- [98] P. Ferreira et al., "Renal Function Underpins the Cyclooxygenase-2: Asymmetric Dimethylarginine Axis in Mouse and Man.," Kidney Int. reports, vol. 8, no. 6, pp. 1231– 1238, Jun. 2023, doi: 10.1016/j.ekir.2023.03.014.
- [99] Z. Li et al., "Prostacyclin facilitates vascular smooth muscle cell phenotypic transformation via activating TP receptors when IP receptors are deficient.," Acta Physiol. (Oxf)., vol. 231, no. 2, p. e13555, Feb. 2021, doi: 10.1111/apha.13555.
- [100] J. Stitham, E. J. Arehart, S. R. Gleim, K. L. Douville, and J. Hwa, "Human prostacyclin receptor structure and function from naturally-occurring and synthetic mutations.," Prostaglandins Other Lipid Mediat., vol. 82, no. 1–4, pp. 95–108, Jan. 2007, doi: 10.1016/j.prostaglandins.2006.05.010.
- [101] M. D. Breyer and R. M. Breyer, "Prostaglandin E receptors and the kidney.," Am. J. Physiol. Renal Physiol., vol. 279, no. 1, pp. F12-23, Jul. 2000, doi: 10.1152/ajprenal.2000.279.1.F12.

- [102] L. Tang, K. Loutzenhiser, and R. Loutzenhiser, "Biphasic actions of prostaglandin E(2) on the renal afferent arteriole : role of EP(3) and EP(4) receptors.," Circ. Res., vol. 86, no. 6, pp. 663–670, Mar. 2000, doi: 10.1161/01.res.86.6.663.
- [103] T. Yang and Y. Du, "Distinct roles of central and peripheral prostaglandin E2 and EP subtypes in blood pressure regulation.," Am. J. Hypertens., vol. 25, no. 10, pp. 1042– 1049, Oct. 2012, doi: 10.1038/ajh.2012.67.
- [104] L. Wang, Y. Wu, Z. Jia, J. Yu, and S. Huang, "Roles of EP Receptors in the Regulation of Fluid Balance and Blood Pressure.," Front. Endocrinol. (Lausanne)., vol. 13, p. 875425, 2022, doi: 10.3389/fendo.2022.875425.
- [105] R. Dalal, Z. S. Bruss, and J. S. Sehdev, "Physiology, Renal Blood Flow and Filtration.," Treasure Island (FL), 2025.
- [106] J. Wang, M. Liu, X. Zhang, G. Yang, and L. Chen, "Physiological and pathophysiological implications of PGE(2) and the PGE(2) synthases in the kidney.," Prostaglandins Other Lipid Mediat., vol. 134, pp. 1–6, Jan. 2018, doi: 10.1016/j.prostaglandins.2017.10.006.
- [107] Z. Guan, J. P. VanBeusecum, and E. W. Inscho, "Endothelin and the renal microcirculation.," Semin. Nephrol., vol. 35, no. 2, pp. 145–155, Mar. 2015, doi: 10.1016/j.semnephrol.2015.02.004.
- [108] Y.-J. Lin et al., "Angiotensin II enhances endothelin-1-induced vasoconstriction through upregulating endothelin type A receptor.," Biochem. Biophys. Res. Commun., vol. 451, no. 2, pp. 263–269, Aug. 2014, doi: 10.1016/j.bbrc.2014.07.119.
- [109] C. Mangana, M. Lorigo, and E. Cairrao, "Implications of Endothelial Cell-Mediated Dysfunctions in Vasomotor Tone Regulation," Biologics, vol. 1, no. 2, pp. 231–251, 2021, doi: 10.3390/biologics1020015.
- [110] G. L. Nádasy, A. Balla, G. Dörnyei, L. Hunyady, and M. Szekeres, "Direct Vascular Effects of Angiotensin II (A Systematic Short Review).," Int. J. Mol. Sci., vol. 26, no. 1, Dec. 2024, doi: 10.3390/ijms26010113.
- [111] M. A. Ayoub, "Angiotensin II type 1 receptor heterodimers in the kidney," Curr. Opin. Endocr. Metab. Res., vol. 16, pp. 96–101, 2021, doi: https://doi.org/10.1016/j.coemr.2020.09.008.
- [112] M. Colin, C. Delaitre, S. Foulquier, and F. Dupuis, "The AT(1)/AT(2) Receptor Equilibrium Is a Cornerstone of the Regulation of the Renin Angiotensin System beyond the Cardiovascular System.," Molecules, vol. 28, no. 14, Jul. 2023, doi: 10.3390/molecules28145481.
- [113] K. D. Burns and N. Li, "The role of angiotensin II-stimulated renal tubular transport in hypertension.," Curr. Hypertens. Rep., vol. 5, no. 2, pp. 165–171, Apr. 2003, doi: 10.1007/s11906-003-0074-1.
- [114] T. Green, A. A. Gonzalez, K. D. Mitchell, and L. G. Navar, "The complex interplay between cyclooxygenase-2 and angiotensin II in regulating kidney function.," Curr. Opin. Nephrol. Hypertens., vol. 21, no. 1, pp. 7–14, Jan. 2012, doi: 10.1097/MNH.0b013e32834d9d75.
- [115] C. Gomez-Alamillo, L. A. Juncos, A. Cases, J. A. Haas, and J. C. Romero, "Interactions Between Vasoconstrictors and Vasodilators in Regulating Hemodynamics of Distinct Vascular Beds," Hypertension, vol. 42, no. 4, pp. 831–836, 2003, doi: 10.1161/01.HYP.0000088854.04562.DA.

- [116] C. De Miguel, J. S. Speed, M. Kasztan, E. Y. Gohar, and D. M. Pollock, "Endothelin-1 and the kidney: new perspectives and recent findings.," Curr. Opin. Nephrol. Hypertens., vol. 25, no. 1, pp. 35–41, Jan. 2016, doi: 10.1097/MNH.00000000000185.
- [117] X. Ma, Y. Liang, W. Chen, L. Zheng, H. Lin, and T. Zhou, "The role of endothelin receptor antagonists in kidney disease.," Ren. Fail., vol. 47, no. 1, p. 2465810, Dec. 2025, doi: 10.1080/0886022X.2025.2465810.
- [118] N. Vignon-Zellweger, S. Heiden, T. Miyauchi, and N. Emoto, "Endothelin and endothelin receptors in the renal and cardiovascular systems.," Life Sci., vol. 91, no. 13–14, pp. 490– 500, Oct. 2012, doi: 10.1016/j.lfs.2012.03.026.
- [119] R. C. Harris, "Physiologic and pathophysiologic roles of cyclooxygenase-2 in the kidney.," Trans. Am. Clin. Climatol. Assoc., vol. 124, pp. 139–151, 2013.
- [120] M.-Z. Zhang, S. Wang, Y. Wang, Y. Zhang, C. Ming Hao, and R. C. Harris, "Renal Medullary Interstitial COX-2 (Cyclooxygenase-2) Is Essential in Preventing Salt-Sensitive Hypertension and Maintaining Renal Inner Medulla/Papilla Structural Integrity.," Hypertens. (Dallas, Tex. 1979), vol. 72, no. 5, pp. 1172–1179, Nov. 2018, doi: 10.1161/HYPERTENSIONAHA.118.11694.
- [121] C. M. Hao, M. Kömhoff, Y. Guan, R. Redha, and M. D. Breyer, "Selective targeting of cyclooxygenase-2 reveals its role in renal medullary interstitial cell survival.," Am. J. Physiol., vol. 277, no. 3, pp. F352-9, Sep. 1999, doi: 10.1152/ajprenal.1999.277.3.F352.
- [122] C. M. Hao, F. Yull, T. Blackwell, M. Kömhoff, L. S. Davis, and M. D. Breyer, "Dehydration activates an NF-kappaB-driven, COX2-dependent survival mechanism in renal medullary interstitial cells.," J. Clin. Invest., vol. 106, no. 8, pp. 973–982, Oct. 2000, doi: 10.1172/JCI9956.
- [123] W. H. Hörl, "Nonsteroidal Anti-Inflammatory Drugs and the Kidney.," Pharmaceuticals (Basel)., vol. 3, no. 7, pp. 2291–2321, Jul. 2010, doi: 10.3390/ph3072291.
- [124] C. Sobolewski, C. Cerella, M. Dicato, L. Ghibelli, and M. Diederich, "The role of cyclooxygenase-2 in cell proliferation and cell death in human malignancies.," Int. J. Cell Biol., vol. 2010, p. 215158, 2010, doi: 10.1155/2010/215158.
- [125] J.-L. Luo, H. Kamata, and M. Karin, "IKK/NF-kappaB signaling: balancing life and death--a new approach to cancer therapy.," J. Clin. Invest., vol. 115, no. 10, pp. 2625– 2632, Oct. 2005, doi: 10.1172/JCI26322.
- [126] V. Singh, D. Gupta, and R. Arora, "NF-kB as a key player in regulation of cellular radiation responses and identification of radiation countermeasures.," Discov. (Craiova, Rom., vol. 3, no. 1, p. e35, Mar. 2015, doi: 10.15190/d.2015.27.
- [127] M. S. Phong, R. D. Van Horn, S. Li, G. Tucker-Kellogg, U. Surana, and X. S. Ye, "p38 mitogen-activated protein kinase promotes cell survival in response to DNA damage but is not required for the G(2) DNA damage checkpoint in human cancer cells.," Mol. Cell. Biol., vol. 30, no. 15, pp. 3816–3826, Aug. 2010, doi: 10.1128/MCB.00949-09.
- [128] J. Yue and J. M. López, "Understanding MAPK Signaling Pathways in Apoptosis.," Int. J. Mol. Sci., vol. 21, no. 7, Mar. 2020, doi: 10.3390/ijms21072346.
- [129] J. H. Fountain, J. Kaur, and S. L. Lappin, "Physiology, Renin Angiotensin System.," Treasure Island (FL), 2025.

- [130] S. Patel, A. Rauf, H. Khan, and T. Abu-Izneid, "Renin-angiotensin-aldosterone (RAAS): The ubiquitous system for homeostasis and pathologies.," Biomed. Pharmacother., vol. 94, pp. 317–325, Oct. 2017, doi: 10.1016/j.biopha.2017.07.091.
- [131] P. C. Underwood and G. K. Adler, "The renin angiotensin aldosterone system and insulin resistance in humans.," Curr. Hypertens. Rep., vol. 15, no. 1, pp. 59–70, Feb. 2013, doi: 10.1007/s11906-012-0323-2.
- [132] E. Higashihara, T. Harada, and H. Fukuhara, "Juxtaglomerular apparatus-mediated homeostatic mechanisms: therapeutic implication for chronic kidney disease.," Expert Opin. Pharmacother., vol. 25, no. 7, pp. 819–832, May 2024, doi: 10.1080/14656566.2024.2357188.
- [133] M. L. S. Sequeira-Lopez and R. A. Gomez, "Renin Cells, the Kidney, and Hypertension.," Circ. Res., vol. 128, no. 7, pp. 887–907, Apr. 2021, doi: 10.1161/CIRCRESAHA.121.318064.
- [134] I. Kosovic et al., "Connexin Signaling in the Juxtaglomerular Apparatus (JGA) of Developing, Postnatal Healthy and Nephrotic Human Kidneys.," Int. J. Mol. Sci., vol. 21, no. 21, Nov. 2020, doi: 10.3390/ijms21218349.
- [135] A. A. Gonzalez, N. Salinas-Parra, D. Leach, L. G. Navar, and M. C. Prieto, "PGE(2) upregulates renin through E-prostanoid receptor 1 via PKC/cAMP/CREB pathway in M-1 cells.," Am. J. Physiol. Renal Physiol., vol. 313, no. 4, pp. F1038–F1049, Oct. 2017, doi: 10.1152/ajprenal.00194.2017.
- [136] H. Francois and T. M. Coffman, "Prostanoids and blood pressure: which way is up?," J. Clin. Invest., vol. 114, no. 6, pp. 757–759, Sep. 2004, doi: 10.1172/JCI22929.
- [137] U. N. Das, "Arachidonic Acid as Mechanotransducer of Renin Cell Baroreceptor.," Nutrients, vol. 14, no. 4, Feb. 2022, doi: 10.3390/nu14040749.
- [138] S. S. Quadri, S. A. Culver, C. Li, and H. M. Siragy, "Interaction of the renin angiotensin and cox systems in the kidney.," Front. Biosci. (Schol. Ed)., vol. 8, no. 2, pp. 215–226, Jun. 2016, doi: 10.2741/s459.
- [139] Y. Yang and G. Xu, "Update on Pathogenesis of Glomerular Hyperfiltration in Early Diabetic Kidney Disease.," Front. Endocrinol. (Lausanne)., vol. 13, p. 872918, 2022, doi: 10.3389/fendo.2022.872918.
- [140] M. S. Goligorsky, "Glomerular microcirculation: Implications for diabetes, preeclampsia, and kidney injury," Acta Physiol., vol. 239, no. 3, p. e14048, 2023, doi: https://doi.org/10.1111/apha.14048.
- [141] T. Yang, Y. Endo, Y. G. Huang, A. Smart, J. P. Briggs, and J. Schnermann, "Renin expression in COX-2-knockout mice on normal or low-salt diets.," Am. J. Physiol. Renal Physiol., vol. 279, no. 5, pp. F819-25, Nov. 2000, doi: 10.1152/ajprenal.2000.279.5.F819.
- [142] J. L. Wang, H. F. Cheng, and R. C. Harris, "Cyclooxygenase-2 inhibition decreases renin content and lowers blood pressure in a model of renovascular hypertension," Hypertension, vol. 34, no. 1, pp. 96–101, 1999, doi: 10.1161/01.HYP.34.1.96.
- [143] J. Schnermann and J. P. Briggs, "Synthesis and secretion of renin in mice with induced genetic mutations.," Kidney Int., vol. 81, no. 6, pp. 529–538, Mar. 2012, doi: 10.1038/ki.2011.451.

- [144] B. Neubauer et al., "Development of vascular renin expression in the kidney critically depends on the cyclic AMP pathway.," Am. J. Physiol. Renal Physiol., vol. 296, no. 5, pp. F1006-12, May 2009, doi: 10.1152/ajprenal.90448.2008.
- [145] A. Pöschke et al., "The PGE(2)-EP4 receptor is necessary for stimulation of the reninangiotensin-aldosterone system in response to low dietary salt intake in vivo.," Am. J. Physiol. Renal Physiol., vol. 303, no. 10, pp. F1435-42, Nov. 2012, doi: 10.1152/ajprenal.00512.2011.
- [146] L. Nilsson, K. Madsen, S. Krag, J. Frøkiær, B. L. Jensen, and R. Nørregaard, "Disruption of cyclooxygenase type 2 exacerbates apoptosis and renal damage during obstructive nephropathy.," Am. J. Physiol. Renal Physiol., vol. 309, no. 12, pp. F1035-48, Dec. 2015, doi: 10.1152/ajprenal.00253.2015.
- [147] C. E. Swan and R. M. Breyer, "Prostaglandin E2 modulation of blood pressure homeostasis: Studies in rodent models," Prostaglandins Other Lipid Mediat., vol. 96, no. 1, pp. 10–13, 2011, doi: https://doi.org/10.1016/j.prostaglandins.2011.07.001.
- [148] M. Baker and M. A. Perazella, "NSAIDs in CKD: Are They Safe?," Am. J. Kidney Dis., vol. 76, no. 4, pp. 546–557, Oct. 2020, doi: 10.1053/j.ajkd.2020.03.023.
- [149] N. Klomjit and P. Ungprasert, "Acute kidney injury associated with non-steroidal antiinflammatory drugs.," Eur. J. Intern. Med., vol. 101, pp. 21–28, Jul. 2022, doi: 10.1016/j.ejim.2022.05.003.
- [150] J. Clària et al., "Effects of celecoxib and naproxen on renal function in nonazotemic patients with cirrhosis and ascites.," Hepatology, vol. 41, no. 3, pp. 579–587, Mar. 2005, doi: 10.1002/hep.20595.
- [151] K. P. Moore and G. P. Aithal, "Guidelines on the management of ascites in cirrhosis.," Gut, vol. 55 Suppl 6, no. Suppl 6, pp. vi1-12, Oct. 2006, doi: 10.1136/gut.2006.099580.
- [152] M. Dixit, T. Doan, R. Kirschner, and N. Dixit, "Significant Acute Kidney Injury Due to Non-steroidal Anti-inflammatory Drugs: Inpatient Setting.," Pharmaceuticals (Basel)., vol. 3, no. 4, pp. 1279–1285, Apr. 2010, doi: 10.3390/ph3041279.
- [153] R. Huang, P. Fu, and L. Ma, "Kidney fibrosis: from mechanisms to therapeutic medicines.," Signal Transduct. Target. Ther., vol. 8, no. 1, p. 129, Mar. 2023, doi: 10.1038/s41392-023-01379-7.
- [154] B. Wang, Z.-L. Li, Y.-L. Zhang, Y. Wen, Y.-M. Gao, and B.-C. Liu, "Hypoxia and chronic kidney disease.," EBioMedicine, vol. 77, p. 103942, Mar. 2022, doi: 10.1016/j.ebiom.2022.103942.
- [155] T. J. Weber, T. J. Monks, and S. S. Lau, "PGE2-mediated cytoprotection in renal epithelial cells: evidence for a pharmacologically distinct receptor.," Am. J. Physiol., vol. 273, no. 4, pp. F507-15, Oct. 1997, doi: 10.1152/ajprenal.1997.273.4.F507.
- [156] K. Höcherl, D. Endemann, M. C. Kammerl, H. F. Grobecker, and A. Kurtz, "Cyclooxygenase-2 inhibition increases blood pressure in rats.," Br. J. Pharmacol., vol. 136, no. 8, pp. 1117–1126, Aug. 2002, doi: 10.1038/sj.bjp.0704821.
- [157] A. A. Gonzalez, T. Green, C. Luffman, C. R. T. Bourgeois, L. Gabriel Navar, and M. C. Prieto, "Renal medullary cyclooxygenase-2 and (pro)renin receptor expression during angiotensin II-dependent hypertension.," Am. J. Physiol. Renal Physiol., vol. 307, no. 8, pp. F962-70, Oct. 2014, doi: 10.1152/ajprenal.00267.2014.

- [158] F. Aslam, A. Haque, J. Haque, and J. Joseph, "Heart failure in subjects with chronic kidney disease: Best management practices.," World J. Cardiol., vol. 2, no. 5, pp. 112– 117, May 2010, doi: 10.4330/wjc.v2.i5.112.
- [159] A. J. Polichnowski and A. W. J. Cowley, "Pressure-induced renal injury in angiotensin II versus norepinephrine-induced hypertensive rats.," Hypertens. (Dallas, Tex. 1979), vol. 54, no. 6, pp. 1269–1277, Dec. 2009, doi: 10.1161/HYPERTENSIONAHA.109.139287.
- [160] R. Bellomo and D. D. Giantomasso, "Noradrenaline and the kidney: friends or foes?," Crit. Care, vol. 5, no. 6, pp. 294–298, Dec. 2001, doi: 10.1186/cc1052.
- [161] Y. Li, Y. Wei, F. Zheng, Y. Guan, and X. Zhang, "Prostaglandin E2 in the Regulation of Water Transport in Renal Collecting Ducts.," Int. J. Mol. Sci., vol. 18, no. 12, Nov. 2017, doi: 10.3390/ijms18122539.
- [162] R. Nasrallah et al., "PGE(2) EP(1) receptor inhibits vasopressin-dependent water reabsorption and sodium transport in mouse collecting duct.," Lab. Invest., vol. 98, no. 3, pp. 360–370, Mar. 2018, doi: 10.1038/labinvest.2017.133.
- [163] J. Rossat, M. Maillard, J. Nussberger, H. R. Brunner, and M. Burnier, "Renal effects of selective cyclooxygenase-2 inhibition in normotensive salt-depleted subjects.," Clin. Pharmacol. Ther., vol. 66, no. 1, pp. 76–84, Jul. 1999, doi: 10.1016/S0009-9236(99)70056-1.
- [164] D. Brater, R. Harris, J. Redfern, and B. Gertz, "Renal effects of COX-2 selective inhibitors," Am. J. Nephrol., vol. 21, pp. 1–15, Jan. 2001.
- [165] B. Largeau, J.-L. Cracowski, C. Lengellé, B. Sautenet, and A.-P. Jonville-Béra, "Druginduced peripheral oedema: An aetiology-based review.," Br. J. Clin. Pharmacol., vol. 87, no. 8, pp. 3043–3055, Aug. 2021, doi: 10.1111/bcp.14752.
- [166] E. I. Agaba, M. Rohrscheib, and A. H. Tzamaloukas, "The renal concentrating mechanism and the clinical consequences of its loss.," Niger. Med. J., vol. 53, no. 3, pp. 109–115, Jul. 2012, doi: 10.4103/0300-1652.104376.
- [167] M. K. Ames, C. E. Atkins, and B. Pitt, "The renin-angiotensin-aldosterone system and its suppression.," J. Vet. Intern. Med., vol. 33, no. 2, pp. 363–382, Mar. 2019, doi: 10.1111/jvim.15454.
- [168] A. Whelton, "Nephrotoxicity of nonsteroidal anti-inflammatory drugs: physiologic foundations and clinical implications.," Am. J. Med., vol. 106, no. 5B, pp. 13S-24S, May 1999, doi: 10.1016/s0002-9343(99)00113-8.
- [169] B. Guthrie, "Can NSAIDs Be Used Safely for Analgesia in Patients with CKD?: CON.," Kidney360, vol. 1, no. 11, pp. 1189–1191, Nov. 2020, doi: 10.34067/KID.0005112020.
- [170] E. Y. F. Wan et al., "Comparative Risks of Nonsteroidal Anti-Inflammatory Drugs on CKD.," Clin. J. Am. Soc. Nephrol., vol. 16, no. 6, pp. 898–907, Jun. 2021, doi: 10.2215/CJN.18501120.
- [171] J. Jankowski, J. Floege, D. Fliser, M. Böhm, and N. Marx, "Cardiovascular Disease in Chronic Kidney Disease: Pathophysiological Insights and Therapeutic Options.," Circulation, vol. 143, no. 11, pp. 1157–1172, Mar. 2021, doi: 10.1161/CIRCULATIONAHA.120.050686.
- [172] D. Saeed et al., "Navigating the Crossroads: Understanding the Link Between Chronic Kidney Disease and Cardiovascular Health.," Cureus, vol. 15, no. 12, p. e51362, Dec. 2023, doi: 10.7759/cureus.51362.

- [173] P. Wehner and W. Nitardy, "Chronic Kidney Disease and Acute Myocardial Infarction: The Story After 1 Year.," Journal of the American Heart Association, vol. 5, no. 5. England, May 2016. doi: 10.1161/JAHA.116.003626.
- [174] C. Zoccali et al., "Diagnosis of cardiovascular disease in patients with chronic kidney disease.," Nat. Rev. Nephrol., vol. 19, no. 11, pp. 733–746, Nov. 2023, doi: 10.1038/s41581-023-00747-4.
- [175] Y. Zhu et al., "The mechanisms underlying acute myocardial infarction in chronic kidney disease patients undergoing hemodialysis," Biomed. Pharmacother., vol. 177, p. 117050, 2024, doi: https://doi.org/10.1016/j.biopha.2024.117050.
- [176] A. Majithia and D. L. Bhatt, "Novel Antiplatelet Therapies for Atherothrombotic Diseases.," Arterioscler. Thromb. Vasc. Biol., vol. 39, no. 4, pp. 546–557, Apr. 2019, doi: 10.1161/ATVBAHA.118.310955.
- [177] G. Passacquale, P. Sharma, D. Perera, and A. Ferro, "Antiplatelet therapy in cardiovascular disease: Current status and future directions.," Br. J. Clin. Pharmacol., vol. 88, no. 6, pp. 2686–2699, Jun. 2022, doi: 10.1111/bcp.15221.
- [178] K.-H. Lee, S.-M. Ou, Y.-C. Chu, Y.-P. Lin, M.-T. Tsai, and D.-C. Tarng, "Antithrombotic Therapy for Chronic Kidney Disease Patients With Concomitant Atrial Fibrillation and Coronary Artery Disease.," Front. Cardiovasc. Med., vol. 8, p. 751359, 2021, doi: 10.3389/fcvm.2021.751359.
- [179] S. Kumar et al., "Anticoagulation in Concomitant Chronic Kidney Disease and Atrial Fibrillation: JACC Review Topic of the Week.," J. Am. Coll. Cardiol., vol. 74, no. 17, pp. 2204–2215, Oct. 2019, doi: 10.1016/j.jacc.2019.08.1031.
- [180] J. F. E. Mann et al., "Effects of aspirin on cardiovascular outcomes in patients with chronic kidney disease.," Kidney Int., vol. 103, no. 2, pp. 403–410, Feb. 2023, doi: 10.1016/j.kint.2022.09.023.
- [181] N. Desai, B. Wilson, M. Bond, A. Conant, and M. Rahman, "Association between aspirin use and cardiovascular outcomes in ALLHAT participants with and without chronic kidney disease: A post hoc analysis.," J. Clin. Hypertens. (Greenwich)., vol. 23, no. 2, pp. 352–362, Feb. 2021, doi: 10.1111/jch.14091.
- [182] D. Kaw and D. Malhotra, "Platelet dysfunction and end-stage renal disease.," Semin. Dial., vol. 19, no. 4, pp. 317–322, 2006, doi: 10.1111/j.1525-139X.2006.00179.x.
- [183] G. R. Nunns et al., "The hypercoagulability paradox of chronic kidney disease: The role of fibrinogen.," Am. J. Surg., vol. 214, no. 6, pp. 1215–1218, Dec. 2017, doi: 10.1016/j.amjsurg.2017.08.039.
- [184] C. C. F. M. J. Baaten, M. Sternkopf, T. Henning, N. Marx, J. Jankowski, and H. Noels, "Platelet Function in CKD: A Systematic Review and Meta-Analysis.," J. Am. Soc. Nephrol., vol. 32, no. 7, pp. 1583–1598, Jul. 2021, doi: 10.1681/ASN.2020101440.
- [185] P. Boccardo, G. Remuzzi, and M. Galbusera, "Platelet dysfunction in renal failure.," Semin. Thromb. Hemost., vol. 30, no. 5, pp. 579–589, Oct. 2004, doi: 10.1055/s-2004-835678.
- [186] G. Di Minno, J. Martinez, M. L. McKean, J. De La Rosa, J. F. Burke, and S. Murphy, "Platelet dysfunction in uremia. Multifaceted defect partially corrected by dialysis.," Am. J. Med., vol. 79, no. 5, pp. 552–559, Nov. 1985, doi: 10.1016/0002-9343(85)90051-8.

- [187] A. Shah, M. F. Hashmi, and N. R. Aeddula, "Chronic Kidney Disease-Mineral Bone Disorder (CKD-MBD).," Treasure Island (FL), 2025.
- [188] N. J. Paloian and C. M. Giachelli, "A current understanding of vascular calcification in CKD.," Am. J. Physiol. Renal Physiol., vol. 307, no. 8, pp. F891-900, Oct. 2014, doi: 10.1152/ajprenal.00163.2014.
- [189] K. W. Davidson et al., "Aspirin Use to Prevent Cardiovascular Disease: US Preventive Services Task Force Recommendation Statement.," JAMA, vol. 327, no. 16, pp. 1577– 1584, Apr. 2022, doi: 10.1001/jama.2022.4983.
- [190] I. Bellos, S. Marinaki, P. Lagiou, and V. Benetou, "Aspirin for the Primary Prevention of Cardiovascular Diseases in Patients with Chronic Kidney Disease: An Updated Metaanalysis.," Am. J. Cardiovasc. drugs drugs, devices, other Interv., vol. 24, no. 2, pp. 241–253, Mar. 2024, doi: 10.1007/s40256-024-00630-y.
- [191] T. D. Warner, S. Nylander, and C. Whatling, "Anti-platelet therapy: cyclo-oxygenase inhibition and the use of aspirin with particular regard to dual anti-platelet therapy.," Br. J. Clin. Pharmacol., vol. 72, no. 4, pp. 619–633, Oct. 2011, doi: 10.1111/j.1365-2125.2011.03943.x.
- [192] V. Aursulesei and I. I. Costache, "Anticoagulation in chronic kidney disease: from guidelines to clinical practice.," Clin. Cardiol., vol. 42, no. 8, pp. 774–782, Aug. 2019, doi: 10.1002/clc.23196.
- [193] R. Maiti, "Prevalence of Aspirin Resistance in Patients of Chronic Kidney Disease," World J. Nephrol. Urol., vol. 1, pp. 79–84, Jan. 2012, doi: 10.4021/wjnu4w.
- [194] H. Khan, O. Kanny, M. H. Syed, and M. Qadura, "Aspirin Resistance in Vascular Disease: A Review Highlighting the Critical Need for Improved Point-of-Care Testing and Personalized Therapy.," Int. J. Mol. Sci., vol. 23, no. 19, Sep. 2022, doi: 10.3390/ijms231911317.
- [195] J. Guo, J. Wang, and J. Feng, "Aspirin resistance mediated by oxidative stress-induced 8-Isoprostaglandin F2.," J. Clin. Pharm. Ther., vol. 44, no. 5, pp. 823–828, Oct. 2019, doi: 10.1111/jcpt.12838.
- [196] C. N. Floyd and A. Ferro, "Mechanisms of aspirin resistance.," Pharmacol. Ther., vol. 141, no. 1, pp. 69–78, Jan. 2014, doi: 10.1016/j.pharmthera.2013.08.005.
- [197] G. J. Hankey and J. W. Eikelboom, "Aspirin resistance.," Lancet (London, England), vol. 367, no. 9510, pp. 606–617, Feb. 2006, doi: 10.1016/S0140-6736(06)68040-9.
- [198] L. R. Lopez, K. E. Guyer, I. G. D. La Torre, K. R. Pitts, E. Matsuura, and P. R. Ames, "Platelet thromboxane (11-dehydro-Thromboxane B2) and aspirin response in patients with diabetes and coronary artery disease.," World J. Diabetes, vol. 5, no. 2, pp. 115–127, Apr. 2014, doi: 10.4239/wjd.v5.i2.115.
- [199] C. Brun et al., "Aspirin response: Differences in serum thromboxane B2 levels between clinical studies.," Platelets, vol. 27, no. 3, pp. 196–202, 2016, doi: 10.3109/09537104.2015.1072147.
- [200] C. P. M. Hayward et al., "Diagnostic utility of light transmission platelet aggregometry: results from a prospective study of individuals referred for bleeding disorder assessments.," J. Thromb. Haemost., vol. 7, no. 4, pp. 676–684, Apr. 2009, doi: 10.1111/j.1538-7836.2009.03273.x.

- [201] K. Koltai, G. Kesmarky, G. Feher, A. Tibold, and K. Toth, "Platelet Aggregometry Testing: Molecular Mechanisms, Techniques and Clinical Implications.," Int. J. Mol. Sci., vol. 18, no. 8, Aug. 2017, doi: 10.3390/ijms18081803.
- [202] U. Baber et al., "Prevalence and impact of high platelet reactivity in chronic kidney disease: results from the Assessment of Dual Antiplatelet Therapy with Drug-Eluting Stents registry.," Circ. Cardiovasc. Interv., vol. 8, no. 6, p. e001683, Jun. 2015, doi: 10.1161/CIRCINTERVENTIONS.115.001683.
- [203] C. C. F. M. J. Baaten et al., "Platelet Abnormalities in CKD and Their Implications for Antiplatelet Therapy.," Clin. J. Am. Soc. Nephrol., vol. 17, no. 1, pp. 155–170, Jan. 2022, doi: 10.2215/CJN.04100321.
- [204] L. B. Cofer et al., "Platelet Activity and Cardiovascular Risk in CKD and Peripheral Artery Disease," Kidney Int. Reports, vol. 7, no. 10, pp. 2242–2250, 2022, doi: https://doi.org/10.1016/j.ekir.2022.07.169.
- [205] L. G. N. et al., "2016 ACC/AHA Guideline Focused Update on Duration of Dual Antiplatelet Therapy in Patients With Coronary Artery Disease," JACC, vol. 68, no. 10, pp. 1082–1115, Sep. 2016, doi: 10.1016/j.jacc.2016.03.513.
- [206] S. V Ittaman, J. J. VanWormer, and S. H. Rezkalla, "The role of aspirin in the prevention of cardiovascular disease.," Clin. Med. Res., vol. 12, no. 3–4, pp. 147–154, Dec. 2014, doi: 10.3121/cmr.2013.1197.
- [207] J. Taliercio et al., "Aspirin for Primary and Secondary Prevention of Mortality, Cardiovascular Disease, and Kidney Failure in the Chronic Renal Insufficiency Cohort (CRIC) Study," Kidney Med., vol. 4, p. 100547, Oct. 2022, doi: 10.1016/j.xkme.2022.100547.
- [208] P. A. McCullough, K. R. Sandberg, S. Borzak, M. P. Hudson, M. Garg, and H. J. Manley, "Benefits of aspirin and beta-blockade after myocardial infarction in patients with chronic kidney disease.," Am. Heart J., vol. 144, no. 2, pp. 226–232, Aug. 2002, doi: 10.1067/mhj.2002.125513.
- [209] A. K. Berger, S. Duval, and H. M. Krumholz, "Aspirin, beta-blocker, and angiotensinconverting enzyme inhibitor therapy in patients with end-stage renal disease and an acute myocardial infarction.," J. Am. Coll. Cardiol., vol. 42, no. 2, pp. 201–208, Jul. 2003, doi: 10.1016/s0735-1097(03)00572-2.
- [210] R. S. Wright et al., "Acute myocardial infarction and renal dysfunction: a high-risk combination.," Ann. Intern. Med., vol. 137, no. 7, pp. 563–570, Oct. 2002, doi: 10.7326/0003-4819-137-7-200210010-00007.
- [211] D. M. Kelly and P. M. Rothwell, "Prevention and treatment of stroke in patients with chronic kidney disease: an overview of evidence and current guidelines.," Kidney Int., vol. 97, no. 2, pp. 266–278, Feb. 2020, doi: 10.1016/j.kint.2019.09.024.
- [212] H. Gallagher et al., "Aspirin to target arterial events in chronic kidney disease (ATTACK): study protocol for a multicentre, prospective, randomised, open-label, blinded endpoint, parallel group trial of low-dose aspirin vs. standard care for the primary prevention of cardio," Trials, vol. 23, no. 1, p. 331, Apr. 2022, doi: 10.1186/s13063-022-06132-z.

- [213] "KDIGO 2024 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease.," Kidney Int., vol. 105, no. 4S, pp. S117–S314, Apr. 2024, doi: 10.1016/j.kint.2023.10.018.
- [214] F. Verbeke et al., "A European Renal Best Practice (ERBP) position statement on the Kidney Disease: Improving Global Outcomes (KDIGO) clinical practice guideline for the management of blood pressure in non-dialysis-dependent chronic kidney disease: an endorsement with some ," Nephrol. Dial. Transplant. Off. Publ. Eur. Dial. Transpl. Assoc. - Eur. Ren. Assoc., vol. 29, no. 3, pp. 490–496, Mar. 2014, doi: 10.1093/ndt/gft321.
- [215] S. Abd ElHafeez, R. Hegazy, Y. Naga, I. Wahdan, and S. Sallam, "Non-steroidal antiinflammatory drugs among chronic kidney disease patients: an epidemiological study.," J. Egypt. Public Health Assoc., vol. 94, no. 1, p. 8, 2019, doi: 10.1186/s42506-018-0005-2.
- [216] M. L. Baker and M. A. Perazella, "Can NSAIDs Be Used Safely for Analgesia in Patients with CKD?: COMMENTARY.," Kidney360, vol. 1, no. 11, pp. 1192–1194, Nov. 2020, doi: 10.34067/KID.0004652020.
- [217] S. Sheth, S. Thakur, A. Thorat, and P. Gupta, "Safe and appropriate use of diclofenac in chronic kidney disease: An Indian perspective.," J. Fam. Med. Prim. care, vol. 10, no. 7, pp. 2450–2456, Jul. 2021, doi: 10.4103/jfmpc.jfmpc_2358_20.
- [218] P. C. Pham et al., "2017 update on pain management in patients with chronic kidney disease.," Clin. Kidney J., vol. 10, no. 5, pp. 688–697, Oct. 2017, doi: 10.1093/ckj/sfx080.
- [219] P. J. Roy, M. Weltman, L. M. Dember, J. Liebschutz, and M. Jhamb, "Pain management in patients with chronic kidney disease and end-stage kidney disease.," Current opinion in nephrology and hypertension, vol. 29, no. 6. England, pp. 671–680, Nov. 2020. doi: 10.1097/MNH.00000000000646.
- [220] N. Singhania, S. Bansal, S. Mohandas, D. P. Nimmatoori, A. A. Ejaz, and G. Singhania, "Role of renin-angiotensin-aldosterone system inhibitors in heart failure and chronic kidney disease.," Drugs Context, vol. 9, 2020, doi: 10.7573/dic.2020-7-3.
- [221] O. A. Safarini, C. Keshavamurthy, and P. Patel, "Calcineurin Inhibitors.," Treasure Island (FL), 2025.
- [222] D. M. Calvo Barbado, L. C. Saiz Fernández, L. Leache Alegría, M. C. Celaya Lecea, and M. Gutiérrez-Valencia, "Acute Kidney Injury associated with 'Triple whammy' combination: a protocol for a systematic review.," F1000Research, vol. 11, p. 496, 2022, doi: 10.12688/f1000research.109987.2.
- [223] K. K. Loboz and G. M. Shenfield, "Drug combinations and impaired renal function -- the 'triple whammy," Br. J. Clin. Pharmacol., vol. 59, no. 2, p. 239—243, Feb. 2005, doi: 10.1111/j.0306-5251.2004.2188.x.
- [224] H. Dahmke et al., "Evaluation of Triple Whammy Prescriptions After the Implementation of a Drug Safety Algorithm.," Drugs - real world outcomes, vol. 11, no. 1, pp. 125–135, Mar. 2024, doi: 10.1007/s40801-023-00405-y.
- [225] S. Jahangiri, S. H. Mousavi, M. R. Hatamnejad, M. Salimi, and H. Bazrafshan, "Prevalence of non-steroidal anti-inflammatory drugs (NSAIDs) use in patients with

hypertensive crisis.," Heal. Sci. reports, vol. 5, no. 1, p. e483, Mar. 2022, doi: 10.1002/hsr2.483.

- [226] Z. Varga, S. R. A. Sabzwari, and V. Vargova, "Cardiovascular Risk of Nonsteroidal Anti-Inflammatory Drugs: An Under-Recognized Public Health Issue.," Cureus, vol. 9, no. 4, p. e1144, Apr. 2017, doi: 10.7759/cureus.1144.
- [227] Z. Bouck et al., "Frequency and Associations of Prescription Nonsteroidal Antiinflammatory Drug Use Among Patients With a Musculoskeletal Disorder and Hypertension, Heart Failure, or Chronic Kidney Disease," JAMA Intern. Med., vol. 178, no. 11, pp. 1516–1525, Nov. 2018, doi: 10.1001/jamainternmed.2018.4273.
- [228] I. R. Ranasinghe, B. Sharma, and K. Bashir, "Hepatorenal Syndrome.," Treasure Island (FL), 2025.
- [229] A. O. Akinbamowo, D. J. Salzberg, and M. R. Weir, "Renal Consequences of Prostaglandin Inhibition in Heart Failure," Heart Fail. Clin., vol. 4, no. 4, pp. 505–510, 2008, doi: https://doi.org/10.1016/j.hfc.2008.03.002.
- [230] E. Pose, S. Piano, A. Juanola, and P. Ginès, "Hepatorenal Syndrome in Cirrhosis.," Gastroenterology, vol. 166, no. 4, pp. 588-604.e1, Apr. 2024, doi: 10.1053/j.gastro.2023.11.306.