FORMULATION AND EVALUATION OF ETHOSOMAL GEL FOR ANTI-AGING

Gupta Rajat Kumar Rajesh^{1*}, Dr. Arun Patel², Dr. Shailendra Patel³ Dr. Naveen Shiyayedi⁴

^{1*2,3,4}Shri Ram Group of Institutions, Department of Pharmacy, Jabalpur-482002 M.P., India. Rajiv Gandhi Proudyogiki Vishwavidyalaya, Bhopal-462033, Madhya Pradesh.

*Corresponding Author - Gupta Rajat Kumar Rajesh.

*Gmail: guptarajat1214@gmail.com Mobile No:8174865558.

Abstract

Ethosomes are soft, malleable, tiny bubble like lipid vesicles composed mainly of phospholipids, ethanol and water. The objective is problem of appearance of wrinkle before age and to prevent chronic effect of signs. The neem plant extract (1mg) was dissolved in methanol, added 1ml of 2 N HCl and filtered and estimated total alkaloids contents. Ethosomes were prepared by solvent dispersion method. The phospholipids (2 - 3 mg) and neem extract (1 mg) was dissolved in ethanol (30 to 40ml) and heated to 30° C \pm 1°C on water bath. We preparated ethosomes suspension containing ethanolic extract with eight ethosomal formulations. We evaluated physical evaluation, measurement of pH, spreadibility, consistency, homogeneity, viscosity, drug content, compatibility studies. We found that formulation FE8 had good values of spreadability, viscosity, pH, drug content and during the accelerated stability studies the appearance was clear and no significant variation in spreadability, pH and drug content was observed. • The formulated gels were subjected to stability studies. The pH of all formulations remained unchanged and was found to be within the range of 6.8-7.3. The viscosity and spreadability of all gels were also unaltered and found to be within the range. The drug content was found to be in the limit 70.21% to 77.52 %, for all gel formulation at all temperature conditions. All the studies performed provides a that novel topical ethosomal gel containing neem extract that can be used as an alternative remedy for management and treatment of anti aging related disorder and disease.

Key words: Ethosomal gel, Anti-aging, Cosmaceuticals, Topical drug delivery systems, Drug release kinetic modelling, Spreadibility.

Introduction

Cosmaceuticals has been used to describe the products that yield benefits traditionally, and active constituents thought to be cosmetic in nature, such as moisturization, as well as product that make marketing claims approaching those of drug products, such as reducing wrinkles, regenerates skin, firms, heal and penetrates into skin. Consumers seek "antiwrinkle" cosmetic products that treat or delay the visible signs of actual aging and weathered skin, such as wrinkles, lines, sagging, and hyper pigmentation and age spots. The present consumers are highly specific about choice of product; consumers prefer mainly products free of synthetic active constituents, synthetic preservatives and base free of animal derivatives. Unnatural, chemically- synthesized products may be perceived as being environmentally or personally unsafe^[1,2]. In contrast, natural products are perceived as pure, mild, and superior to chemically synthesized products. Natural based products extracted from plants or herbs are believed to contain antioxidant/free-radical scavenging agents that can neutralize the effects of free-radical damage^[3,4]. Additionally, they contain agents that stimulate the synthesis and restoration of damaged connective tissue structures in the dermis and barrier function in the epidermis^[5,6]. In spite of the various anti- aging cosmetic products on the market for the treatment of skin, there remains a need for effective topically applied cosmetic compositions that provide anti-aging or rejuvenating benefits to the skin, using natural ingredients as active components. They also expect the latest technology advances to be incorporated into innovative formulations^[7].

Organs of all the creatures including human grow from birth, and gradually decline with age, then, functions thereof are deactivated. When the deactivated part exceeds a certain extent, the creature dies. The process that the functions thereof are gradually declining is called aging. Skin is directly affected by surroundings and has important functions to maintain circumstance of the inner part of living bodies. Intrinsic aging, also known as the natural aging process, is a continuous process that normally begins in our mid-20s. Within the skin, collagen production slows, and elastin, the substance that enables skin to snap back into place, has a bit less spring. Dead skin cells do not shed as quickly and turnover of new skin cells may decreases slightly. While these changes usually begin in our 20s, the signs of intrinsic aging are typically not visible for decades^[8,9].

In the early 1980s, Mezei and his group described liposomes as the first topical lipid vesicular system for enhanced drug delivery to the skin. Since then, many works have shown that lipid vesicular systems are able to increase the accumulation of various molecules in the SC or other upper skin layers. Ethosomes are soft, malleable, tiny bubble like lipid vesicles composed mainly of phospholipids, ethanol (relatively high

concentration) and water. These "soft vesicles" represents smart vesicular carrier for enhanced delivery to/through skin. All components of the Ethosomal systems are considered as being safe for pharmaceutical and cosmetic use^[10,11].

Up till now work has been done over ethosomes to deliver one of herbal derivative that is ammonium salt of glycyrrhizic acid (antinflammatory agent) of Glycyrrhiza glabra. *In-vitro* and *in-vivo* experiments were carried out; Ethosome suspension showed very good skin tolerability in human volunteers, increases of in vitro percutaneous permeation and significantly enhanced anti-inflammatory activity of glycyrrhizic acid. Ethosomes even reported to deliver many synthetic drug for example Salbutamol Sulfates, Minoxidil, Cannabidol, Acyclovir, Anti-HIV Agents, zidovudine, Lamivudine, Trihexiphenidyl hydrochloride Insulin, Azelaic acid, Erythromycin, Bacitracin, DNA, Diclofenac, peptides^[12,13].

The main objective behind choosing this work is to treat world's today major problem that is appearance of wrinkle before age and to prevent chronic effect of signs which are results of various environmental factors like pollution, UV rays, toxic chemicals through effective and safe treatment (by complete natural product) within less period of time. Rationale behind choosing ethosomes as delivery system to fulfill objective of topical delivery system that is enhanced delivery of active agents at deeper level of skin, packed less tightly than conventional vesicles but has equivalent stability, allowing a more malleable structure and improves distribution in *stratum corneum* lipids, phospholipids present in ethosomes rejuvenate cell membrane and ethanol present in formulation act as preservative. Ethosomes containing extract via lotion are incorporated into cream. Even the cream is formulated by taking all natural components and no preservative was added in cream to make it safe. Cream is full of natural oils which nourishes and moisturizes skin effectively and are rich in omega oils which is highly required for collagen production [14,15].

Materials and Methods

Collection of plant material: Neem leaves were collected locally from Bhopal, a tree in college campus, required whole leaves were collected from same tree. Collected Neem leaves were washed by distilled water two times and dried under shade for one month.

Preparation of plant powder: The plant was dried under shade and then powdered coarsely with a mechanical grinder. The powder was passed through sieve No. 40 and stored in an airtight container for further use.

Preparation of extracts: The dried powder of plant leave was extracted with

Ethanol using Soxhlet apparatus. Accurate 250 gm of dried powder leave of plant was subjected to soxhlation. It was first defatted with petroleum ether then exhaustively extracted with solvent in a Soxhlet apparatus for 36 hours. The temperature was maintained at 40-50 degree centigrade.

Table No. 1: Formulations of Gel containing ethanolic extract of *neem* leaves

| Ingredient | F1 | F2 | F3 | F4 | F5 | F6 | F7 | F8 |
|----------------------------|------|------|------|------|------|------|-----------|------|
| Carbopol 934 (gm) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Sodium CMC (gm) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Ethosomes of neem (1% w/w) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Methyl Paraben (0.2%) | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 |
| Triethanolamine (ml) | q.s. | q.s. |
| Distilled water (100ml) | q.s. | q.s. |

Characterization and Evaluation of Gel Formulation

Physical Evaluation: Physical parameters such as color and appearance of the herbal gel were observed manually.

Measurement of pH: The pH of various gel formulations was determined by using digital pH meter. One gram of gel was dissolved in 100 ml distilled water and stored for two hours. The measurement of pH of each formulation was done in triplicate and average value was calculated.

Spreadibility: Spreadibility was determined by the apparatus which consists of a wooden block, provided with pulley at one end. By this method spreadibility was measured on the basis of slip and drag characteristics of gels.

$$S = M \times L / T$$

Where, S = Spreadibility, M = Weight in the pan (tied to the upper slide), L = Length moved by the glass slide and T = Time (in sec.) taken to separate the slide completely each other.

Homogeneity: All developed gels were tested for homogeneity by visual inspection after setting gels in the container. They were observed for their appearance and presence of any aggregates.

Viscosity: Viscosity of gel was measured by using Brookfield viscometer with spindle No. 7 at 50 rpm at room temperature.

Drug content: 1 g of the prepared gel was mixed with 100 ml of suitable solvent.

Compatibility studies: Fourier transformed infrared (FTIR) spectra technique has been used to study the physical and chemical interaction.

Drug release kinetic modelling: The kinetics of ethosomal gel FE release was determined using the release kinetics method of drug release into various kinetic equations: zero order release kinetics, first order release kinetics and Higuchi model.

Accelerated Stability Studies: The optimized formulations were subjected to a stability testing for six months as per ICH norms at a temperature and RH of 40° C \pm 2° C/75% RH \pm 5% RH respectively.

RESULT AND DISCUSSION

Preliminary Study Morphological Characterization of *Neem* leaves

Table No. 2: Morphology of *Neem* leaves

| S. No. Character | | Observation |
|------------------|---------|----------------------------------|
| 1 | Color | Green |
| 2 | Odor | Bitter |
| 3 | Taste | Bitter |
| 4 | Size | Length – 1.5-3cm, Width -1-1.5cm |
| 5 | Shape | Ovate |
| 6 | Texture | Rough outer periphery |

Physiochemical analysis of Neem leaves powder

Table No.3: Physiochemical analysis of powder of *Neem* leaves

| S. No. | Parameters | Observation (%) |
|--------|--------------------------|-----------------|
| 1 | Total ash value | 9 |
| 2 | Loss on drying | 1.2 |
| 3 | Acid insoluble ash value | 2.9 |
| 4 | Water soluble ash value | 1.6 |
| 5 | Foaming index | 6 (ml) |

Extract of Neem Leaves

Table No 4: Extractive values of extracts of Neem Leaves

| S. No. Solvents | | % Yield (w/w) | Color | Consistency | |
|-----------------|------------|---------------|------------|-------------|--|
| 1 | Pet. Ether | 2.91 % | Dark green | Greasy | |
| 2 | Methanol | 9.34 % | Dark green | Semi Solid | |

Total alkaloid content estimation (TAC)

Total alkaloid content was calculated as atropine equivalent mg/100mg using the equation based on the calibration curve: y = 0.008x + 0.010, $R^2 = 0.999$, where X is the Atropine equivalent (AE)and Y is the absorbance.

Table No. 5: Preparation of calibration curve of Atropine

| S. No. | Concentration (µg/ml) | Mean Absorbance |
|--------|-----------------------|-----------------|
| 1 | 0 | 0 |
| 2 | 40 | 0.352 |
| 3 | 60 | 0.514 |
| 4 | 80 | 0.679 |
| 5 | 100 | 0.845 |
| 6 | 120 | 0.997 |

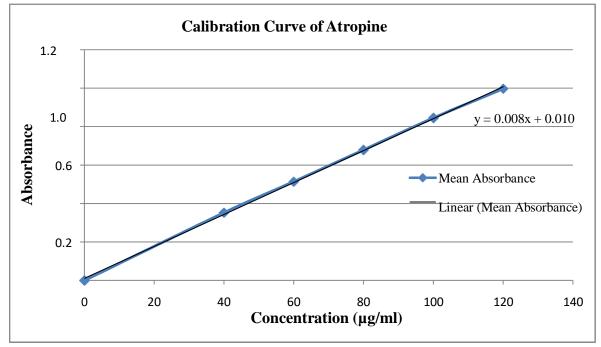


Figure 1: Graph of Calibration Curve of Atropine

Table 6: Estimation of total alkaloid content Neem Leaves extract

| | S. No. Extract | | Total alkaloid content (mg/ 100 mg of dried extract) |
|---|----------------|------------------|--|
| Ī | 1. | Methanol extract | 0.632 |

Formulation of Ethosomes

Table 7: Preparation of Ethosomes Suspension Containing Ethanolic Extract

| Ingredient | FE1 | FE2 | FE3 | FE4 | FE5 | FE6 | FE7 | FE8 |
|------------------------------|------|------|------|------|------|------|------|------|
| Ethanol extract of Neem (mg) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Phospholipid (mg) | 2 | 2 | 2 | 2 | 3 | 3 | 3 | 3 |
| Ethanol (ml) | 30 | 40 | 30 | 40 | 30 | 40 | 30 | 40 |
| Tween 20 (ml) | - | - | 2 | 2 | - | - | 2 | 2 |
| SLS (mg) | - | - | 500 | 500 | - | - | 500 | 500 |
| Distilled Water (upto 100ml) | q.s. |

Characterization of Neem Extract Loaded Ethosome Suspension

Image analysis of ethosomes by optical microscope: For the initial vesicle characterization of ethosome suspension were examined by compound microscope. The result revealed that formulation without added SLS and Tween 20 shown aggregation process among the

structure. Formulation in which SLS and Tween 20 was added shown spherical shaped vesicle structure without aggregation. Hence, Formulation containing SLS and Tween 20 in ehanolic extract (FE3, FE4, FE7 and FE8) were emphasized in further study.

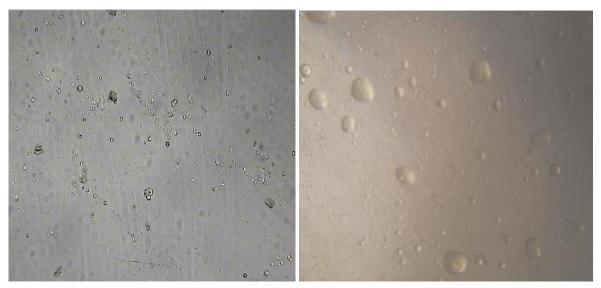


Figure 2: Microphotographs of Ethosome suspension with SLS and Tween (left) withoutSLS and Tween (right)

Vesicular shape and surface morphology by TEM: The vesicular shape and surface morphology of ethosomes of formulation FE8 examined by Transmission Electron Microscope (TEM). The TEM image showed that ethosomes were spherical shaped.

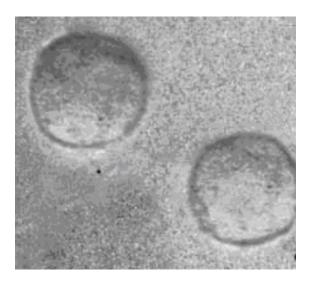


Figure 3: Visualization of ethosomes by TEM

Table No. 8: Characterization of Neem Extract Phytosome

| Batch | X7:-1.1 (0/) | Drug | Mean Particle | Encapsulation |
|-------|------------------|------------------|---------------|------------------|
| Code | Yield (%) | Content (%) | Size (nm) | Efficiency (%) |
| EF1 | 89.12 ± 0.05 | 70.21 ± 0.21 | 732 ± 13 | 78.67 ± 1.52 |
| EF2 | 88.32 ± 0.08 | 72.35 ± 0.76 | 694 ± 29 | 84.27 ± 0.81 |
| EF3 | 88.91 ± 0.03 | 77.52 ± 1.90 | 637 ± 17 | 81.34 ± 0.64 |
| EF4 | 89.45 ± 1.06 | 71.41 ± 1.63 | 713 ± 44 | 82.52 ± 1.30 |
| EF5 | 83.48 ± 1.95 | 70.44 ± 1.02 | 694 ± 21 | 72.19 ± 1.68 |
| EF6 | 81.32 ± 0.32 | 73.32 ± 0.08 | 765 ± 62 | 71.62 ± 0.37 |
| EF7 | 82.91 ± 0.31 | 72.91 ± 0.03 | 876 ± 32 | 90.62 ± 0.11 |
| EF8 | 87.45 ± 0.06 | 71.45 ± 1.06 | 954 ± 27 | 93.87 ± 0.47 |

Determination of Entrapment efficiency of ethosomes suspension: The entrapment efficiency of various ethosomes formulations are presented in Table. The entrapment efficiency of formulation FE8 was found to be highest (93.87%), EF7 (90.62%), EF4 (82.52%) while FE3 formulation showed entrapment efficiency (81.34%). However it has been observed the formulation containing phospholipid with ethanol has maximum entrapment efficiency.

In-vitro **drug release study:** Percentage drug release of hydrogel containing ethosomes (FE1- FE8) was observed at 360min 78.31%, 66.31%, 83.23%, 87.83%, 79.22%, 75.63%, 82.33% and 90.53% respectively. It was observed that addition of ethanol in formulation increase the release by increasing permeation properties of ethosomes. The ethosome (formulation FE8) showed maximum drug release as compared to others formulation.

Table No. 9: Cumulative % of drug release from Ethosome

| Table 110. 2. Cumulative 70 of drug release from Ethosome | | | | | | | | |
|---|-------|-------|-------|-------|-------|-------|-------|-------|
| Time (min) | FE1 | FE2 | FE3 | FE4 | FE5 | FE6 | FE7 | FE8 |
| 15 | 12.42 | 11.21 | 10.33 | 12.42 | 12.52 | 11.22 | 12.42 | 12.02 |
| 30 | 19.43 | 15.92 | 13.93 | 18.12 | 18.21 | 16.13 | 18.22 | 19.43 |
| 60 | 24.53 | 22.42 | 17.47 | 20.32 | 22.42 | 24.33 | 25.32 | 26.45 |
| 90 | 32.55 | 36.32 | 27.32 | 31.47 | 30.61 | 38.85 | 40.64 | 46.38 |
| 120 | 44.32 | 41.31 | 36.21 | 41.22 | 42.53 | 42.63 | 50.17 | 51.53 |
| 180 | 59.46 | 58.27 | 54.23 | 59.64 | 58.39 | 59.93 | 68.34 | 69.81 |
| 240 | 77.12 | 62.42 | 68.31 | 69.93 | 74.17 | 67.24 | 75.57 | 78.39 |
| 360 | 78.31 | 66.31 | 83.23 | 87.83 | 79.22 | 75.63 | 82.33 | 90.53 |

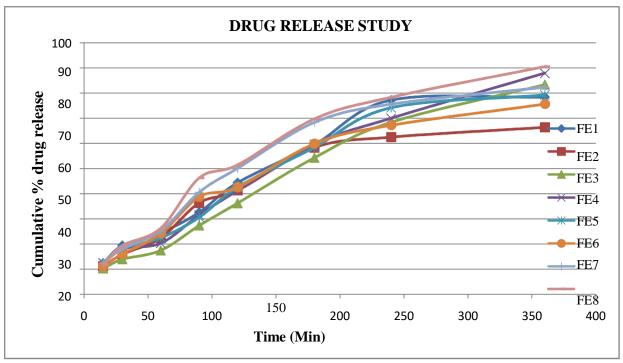


Figure 4: Drug release profile of ethosome suspension containing neem leaves extract

Formulation of topical gel of *Neem* extract loaded ethosome Table No. 10: Formulations of Gel containing ethanolic extract of *neem* leaves.

| Ingredient | F1 | F2 | F3 | F4 | F5 | F6 | F7 | F8 |
|-----------------------------------|------|------|------|------|------|-----------|-----------|------|
| Carbopol 934 (gm) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Sodium CMC (gm) | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| Ethosomes of <i>neem</i> (1% w/w) | FE1 | FE2 | FE3 | FE4 | FE5 | FE6 | FE7 | FE8 |
| Methyl Paraben (0.2%) | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 | 0.2 |
| Triethanolamine (ml) | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. |
| Distilled water (100ml) | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. | q.s. |

Evaluation of Neem Extract Loaded ethosomal gels Table No. 11: Physical Evaluation of Neem Extract Loaded ethosome gels

| Formulation code | Clarity | Odor | Phase Separation | Wash ability | Homogeneity | Grittiness |
|------------------|---------|------|---------------------|--------------|-------------|------------|
| F-1 | Clear | No | No | Washable | Homogeneous | No |
| F-2 | Clear | No | No | Washable | Homogeneous | No |
| F-3 | Clear | No | No | Washable | Homogeneous | No |
| F-4 | Clear | No | No | Washable | Homogeneous | No |
| F-5 | Clear | No | No | Washable | Homogeneous | No |
| F-6 | Clear | No | No | Washable | Homogeneous | No |
| F-7 | Clear | No | No | Washable | Homogeneous | No |
| F-8 | Clear | No | No | Washable | Homogeneous | No |

Table No. 12: Evaluation of Neem Extract Loaded Ethosome gels

| | | | | 0 |
|-------------|----|--------------------|---------------|--------------|
| Formulation | рH | Spreadability (cm) | Viscosity(cp) | % Permeation |

| code | - | | | |
|----------------|-----|---------------|---------------|-------|
| F-1 | 6.9 | 5.6 ± 0.3 | 100 ± 1.8 | 83.2% |
| F-2 | 6.8 | 6.8 ± 0.2 | 99 ± 2.0 | 86.3% |
| F-3 | 7.1 | 7.1 ± 0.6 | 114 ± 1.2 | 92.7% |
| F-4 | 7.3 | 7.6 ± 0.2 | 100 ± 0.8 | 91.0% |
| F-5 | 7.1 | 7.9 ± 0.6 | 118 ± 2.6 | 90.1% |
| . <u>.</u> F-6 | 7.0 | 6.9 ± 0.3 | 123 ± 2.6 | 90.7% |
| ·ig F-7 | 7.2 | 7.3 ± 0.1 | 105 ± 2.6 | 89.1% |
| Ę F-8 | 6.9 | 7.1 ± 0.7 | 113 ± 2.6 | 90.5% |
| <u>i</u> | | | | |

Table No. 13: In-Vitro Drug Release Profile of Phytosome gel from FE 8 Formulation

| Time (T) | S.R.T. | Log T. | %C.R | Log % C.R | Drug | Log% drug |
|----------|--------|--------|-------|-----------|-----------|-----------|
| (Hr.) | | | | | remaining | remaining |
| 0 | 0 | 0 | 0 | 0 | 100 | 2 |
| 15 | 3.873 | 1.176 | 12.02 | 1.080 | 87.02 | 1.939 |
| 30 | 5.477 | 1.477 | 19.43 | 1.288 | 80.57 | 1.906 |
| 60 | 7.746 | 1.778 | 26.45 | 1.778 | 73.55 | 1.867 |
| 90 | 9.487 | 1.954 | 46.38 | 1.954 | 53.38 | 1.727 |
| 120 | 10.954 | 2.079 | 51.53 | 2.079 | 48.47 | 1.685 |
| 180 | 13.416 | 2.255 | 69.81 | 2.255 | 30.81 | 1.489 |
| 240 | 15.492 | 2.380 | 78.39 | 2.380 | 21.39 | 1.330 |
| 360 | 18.974 | 2.556 | 90.53 | 2.519 | 09.47 | 0.976 |

Drug release kinetic modeling

On comparison of kinetic modeling and release profile data it was evident that ethosome FE-8 was found to release the drug in accordance to Higuchi kinetics, the regression coefficient was not found to be exactly near to 1, which could be due to influence of some other factors.

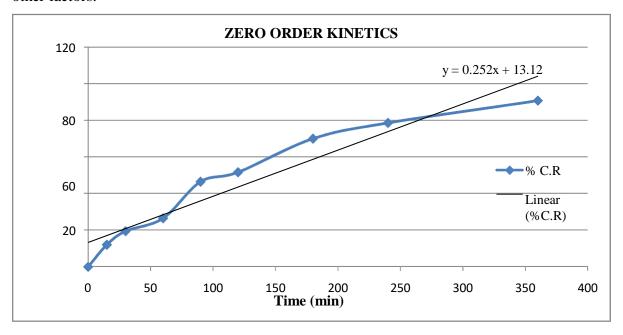


Figure 5: Zero order kinetics of formulation FE8



Figure 6: First order kinetics of formulation FE8

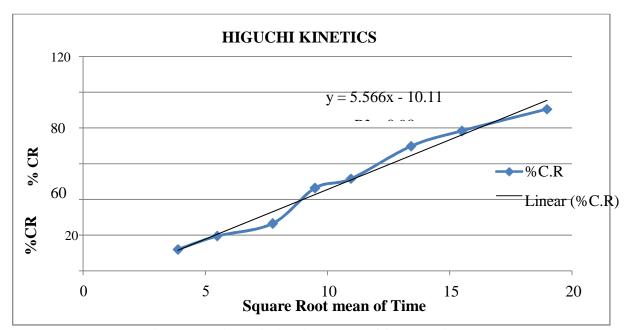


Figure 7: Higuchi kinetic Model of formulation FE8

Stability Study

The formulated gels were subjected to stability studies. No color fading was observed for all prepared gels. The pH of all formulations remained unchanged and was found to be within the range of 6.2-7.2. Viscosity and spreadability of all gels remained unaltered and found to be within the range. Drug content was found to be in the limit 90% -103% for all gel formulation.

Table 14: Accelerated Stability study of formulated gel

| Batch | Clarity/ | Phase | Spreadibility | Viscosity | ъU | Drug |
|-------|-------------|------------|---------------|---------------|------|------------------|
| | Homogeneity | Separation | (gm.cm/sec) | (cps) | pН | Content (%) |
| F-1 | Homogeneous | No | 4.9 ± 1.3 | 142 ± 1.3 | 7.0 | 69.01 ± 0.32 |
| F-2 | Homogeneous | No | 6.1 ± 0.3 | 134 ± 1.4 | 7.0 | 70.45 ± 1.06 |
| F-3 | Homogeneous | No | 7.0 ± 1.2 | 119 ± 1.2 | 7.2 | 76.12 ± 1.10 |
| F-4 | Homogeneous | No | 6.8 ± 0.3 | 123 ± 1.2 | 7. 1 | 71.14 ± 1.13 |
| F-5 | Homogeneous | No | 7.7 ± 0.1 | 152 ± 1.3 | 7. 3 | 72.63 ± 1.12 |
| F-6 | Homogeneous | No | 7.0 ± 0.4 | 149 ± 0.4 | 7.2 | 72.62 ± 1.03 |
| F-7 | Homogeneous | No | 7.3 ± 0.5 | 110 ± 0.3 | 7.2 | 70.97 ± 1.05 |
| F-8 | Homogeneous | No | 7.3 ± 0.2 | 117 ± 0.5 | 6.8 | 70.86 ± 1.71 |

Discussion

Ethosomes of neem leave extract has been successfully formulated in this procedure firstly Azadirachta indica leaves were collected locally from Bhopal then washed and dried. Neem leaves were morphological Characterized as green in color bitter in taste with ovate shape. Neem leave powder was subjected for various physiochemical analysis like total ash value (9), Loss on Drying (1.2), Acid Insoluble Ash Value (2.9), Water Soluble Ash Value (1.6) and Foaming Index (6 ml) and then extracted out methanolic fraction of neem leaves, yield of the extract which was 9.34%. Estimation of total alkaloid content in extract of neem leaves as Total alkaloid content was atropine equivalent 0.632 mg/100 mg using the equation based on the calibration curve: y = 0.008x + 0.010, R2 = 0.999, where X is the Atropine equivalent (AE) and Y is the absorbance.

Eight different types of Ethosome formulations were formed using fixed amount of ethanol extract of neem leaves, Phospholipid (2-3mg), ethanol (30-40ml) and surfactants e.g. tween20 and SLS. These formulations were characterized under various parameters like yield, drug content, particle size and Encapsulation Efficiency, also morphology like size and shape was observed via TEM. For all formulation yield was 81.32 % to 89.45%, drug content was 70.21% to 77.52 %, Mean Particle Size (nm) 637nm to 954nm and entrapment Efficiency was 71.62% to 93.87%. Percentage drug release of hydrogel containing ethosomes (FE1-FE8) was observed at 360min 78.31%, 66.31%, 83.23%, 87.83%, 79.22%, 75.63%, 82.33% and 90.53% respectively.

It was observed that addition of ethanol in formulation increase the release by increasing permeation properties of ethosomes. The ethosome (formulation FE8) showed maximum drug release as compared to others formulation. All eight Ethosome formulations were incorporated with gel and formed clear, odorless, washable, homogeneous, stable and free from grittiness gel was evaluated under the various parameter, pH of all formulations were observed between 6.8 to 7.3 and Spreadability between 5.6 to 7.9 cm, and viscosity

between 99 to 123 centi poice (cp) and % permeation between 83.2 % to 92.7 %. But on the basis of drug release profile F-8 formulation was very good because its % drug release was 90.53% followed First Order Kinetic Model. Three good formulation's drug releases were F-3 (83.23), F-4 (87.83) and F-7 (82.33). All gel formulations were F-1 to F-8 are stable at 4oC and 40oC showing homogeneity, no significant change in phase separation, Spreadibility, Viscosity, pH and Drug Content (%).

CONCLUSIONS

In the present study, an attempt was made to prepare, characterize and evaluate of topical therapeutic system of anti-aging system from Azadirachta indica herbal plant. Various formulations such as hydrogel, hydroalcohalic gel and ethosomal gel were designed and optimized. From the present study, the following conclusion can be drawn: The extraction of Azadirachta indica was carried out with ethanol by using soxhlet apparatus. The hydrogel and hydroalcohalic gel formulation of ethanolic acetate extract were designed by using varied concentration of carbopol and sodium CMC polymer. During the trial, the excipients concentrations of carbapol and Sodium CMC were gradually increased and then decreased as several problems like homogeneity, spreadibility and viscosity were encountered. These problems occured in some of the batches of polymer based gel containing Azadirachta indica. Hence, these batches were discarded and remaining batches were characterized for various parameter. The result showed that the developed herbal gel was greenish in color, translucent in appearance and showed good homogeneity with absence of lumps. Formulation FE8 had good values of spreadability, viscosity, pH, drug content and during the accelerated stability studies the appearance was clear and no significant variation in spreadability, pH and drug content was observed.

Percentage drug release of hydrogel containing ethosomes (FE1-FE8) was observed at 360 min 78.31%, 66.31%, 83.23%, 87.83%, 79.22%, 75.63%, 82.33% and 90.53% respectively. It was observed that addition of ethanol in formulation increase the release by increasing permeation properties of ethosomes. The ethosome (formulation FE8) showed maximum drug release as compared to others formulation. The formulated gels were subjected to stability studies. No color fading was observed for all prepared gels. The pH of all formulations remained unchanged and was found to be within the range of 6.8-7.3. The viscosity and spreadability of all gels were also unaltered and found to be within the range. The drug content was found to be in the limit 70.21% to 77.52 %, for all gel formulation at all temperature conditions. Stability studies revealed that all optimized formulations containing Azadirachta indica extract are stable and can be explored as a marketed product.

Natural remedies are more acceptable in the belief that they are safer with fewer side effects than the synthetic ones. Herbal formulations have growing demand in the world market. In the undertaken study an attempt has been made to establish that herbal gel containing Neem extract has promising anti-aging action. All the studies performed provides a that novel topical ethosomal gel containing neem extract that can be used as an alternative remedy for management and treatment of anti aging related disorder and disease.

REFERENCES

1) Kapoor S, Saraf S. Risk analysis tools for toxicological profile of cosmetics. Inter J Toxicol. 2008 (In press).

- 2) Kapoor S, Saraf S. Regulatory bodies and truth about cosmetic ingredients. Farmavita.net, 2007; 6 Nov.
- 3) Ashawat MS, Saraf S and Swarnlata Saraf. Biochemical and histopathological studies of herbal cream against UV radiation induced damage. Trends in Medical Research, 2007; 2(3): 135-141.
- 4) Ashawat MS, Saraf S and Swarnlata Saraf. Phytosomes: A novel approach towards functional cosmetics. J Plant Sci. 2007; 2(6): 644-649.
- 5) Ashawat MS, Saraf S and Swarnlata Saraf. Cosmetic Potentiality of plant extracts and natural oils. Bioscie Bitechnol Res Asia. 2006; 3: 181-188.
- **6**) Ashawat MS, Saraf S and Swarnlata Saraf. Antisolar activity of *R. Damnesia* and *T. Errecta*. Planta Indica. 2005; 2: 26-28.
- 7) Ashawat MS, Saraf S and Swarnlata Saraf. Preparation and characterization of herbal creams for improvement of skin viscoelastic properties. Int J Cos Sci. 2008; 30: 183-193.
- 8) Kaur CD, Saraf S, Novel approaches in herbal cosmetics. J Cos Dermatol. 2008;7: 89-95.
- 9) Shrivastava S, Kapoor S and Saraf S. Novel preparation and evaluation of lotion containing aloe gel beads. Ind J Pharm Edu Res. 2003; 42(2): 77-80.
- **10**) Kaloysky. ER, Zheng. M, Duggan.CM, Compositions and delivery methods for the treatment of wrinkles, fine lines and hyperhidrosis US Patent 6866856 Issued on March 15, 2005.
- **11**) Koh JS. Cigarette smoking associated with premature facial wrinkling: image analysis of facial skin replicas. Int J Dermatol. 2002; 41(1): 21-27.
- **12**) Patravale VB, Mandawgade SD. Novel cosmetic delivery systems: an application update. Int J Cos Sci. 2007; 30: 19–33.
- 13) Saraf S, Kapoor S. Cosmetic legislation. Pune, India: Niarli parkasahan 2008.
- 14) Cioca G, Calvo L. Liquid crystals and cosmetic applications. Cosmet Toil. 1990;105:57-62.
- **15**) Schmidt JB, Binder M and Macheines UV et. al. New treatment of atrophic acne scars by iontophoresis with estriol and tretinoin. Int J Dermatol. 1195; 34: 57–62.
- **16**) Honeywell-Nguyen PL, Bouwstra JA. The in vitro transport of pergolide from surfactant-based elastic vesicles through human skin: a suggested mechanism of action. J ControlRelease. 2003; 86: 145-156.
- **17**) Birman M, Lawrence N. Liposome stability via multi-walled delivery systems. Cosmet Toil. 2002; 117: 51-58.
- **18**) Bendas ER, Tadros MI. Enhanced transdermal delivery of Salbutamol Sulfate via ethosomes. AAPS Pharm Sci Tech. 2007; 8(4): Article 107.
- 19) Imbert D, Wickett R R. Topical delivery with liposomes. Cosmet Toil. 1995; 110: 32-45.
- **20**) American Academy of Dermatology. Turning Back the Hands of Time. February 21, 2005.

- 21) Fisher GJ. "The Pathophysiology of Photoaging of the Skin" Cutis, 2005; 75(2S):5-9.
- **22)** Moschella S, Hurley H. "Aging and Its Effects on the Skin." Dermatology: 3rd ed. Philadelphia: W.B. Saunders Company; 1992.
- 23) Oikarinen A. "Aging of the skin connective tissue: how to measure the biochemical and mechanical properties of aging dermis." Photodermatology, Photoimmunology and Photomedicine, 1994 Apr; 10 (2):47-52.
- **24**) Scimeca. JV, Zimmerman. CA, Metller FM, Kudo. A (Wo/2006/041526) composition comprising a *Rosmarinus officinalis* plant extract, an alpinia plant extract and a DNA repair enzyme.
- **25**) Touitou E, Godin B and Weiss C. Enhanced delivery of drugs into and across the skin by ethosomal carriers. Drug Development Research. 2000; 50: 406-415.
- **26**) Touitou et al, Ethosomes- efficiently delivering active agents to skin personal care, Jan.2005; 6(1): 71-74.
- **27**) P Sanjay, Ethosomes:A promising tool for transdermal delivery of drug. www.pharmainfonet.com
- **28**) Bendas ER, Tadros MI. Enhanced transdermal delivery of Salbutamol Sulfate via ethosomes. AAPS PharmSci Tech. 2007; 8(4): Article 107.
- **29**) Touitou E, Dayan N and Bergelson L. Ethosomes-novel vesicular carriers for enhanced delivery: characterization and skin penetration properties. J Control Release. 2000;65: 403-418.
- **30**) Cevc. G, lipid vesicles and other colloids as drug carriers on the skin; Advanced drug delivery Reviews 2004; 56:675-711.
- **31**) Dubey V, Mishra D, Dutta T et. al. Dermal and transdermal delivery of an anti-psoriatic agent via ethanolic liposomes. J Control Release. 2007; 123: 148-154.
- **32**) Jain S, Mishra D, Kuksal A et. al. Vesicular Approach for Drug Delivery into or Across the Skin: Current Status and Future Prospects. http://www.priory.com/pharmol/Manuscript-Jain.htm
- **33**) Merdan VM, Alhaique F, and Touitou E, Vesicular carriers for topical delivery. Acta Techno. LegisMedicament 1998;12: 1-6.
- **34**) Barry BW, Istransdermal drug delivery research still important today. Drug Delivery Tech..2001;6: 967-971.
- **35**) Assessed from URL: http://www.ntt-inc.com/default.asp, Company profile:Noveltherapeutic technologies: Ethosomes.
- **36)** Donatella P, Giuseppe L and Domenico M.. Ethosomes for skin delivery of ammonium glycyrrhizinate: In vitro percutaneous permeation through human skin and in vivo anti-inflammatory activity on human volunteers. J Control Release. 2005;106(1): 99-110.
- **37**) Kim JC, Lee MH and Rang MJ. Minodixil containing dosage forms: skin retention and after rising hair growth promotion, Drug Delivery, 2003;10(2): 119-123.

38) Lodzki M, Godin B, Rakou L et. al. Cannabidol transdermal delivery and anti-inflammatory effect in a marine model. J Control Release. 2003;93: 379-389.

- **39**) Horwitz E, Pisanty S,Czerninski R et al. A clinical evaluation of a novel liposomal carrier for acyclovir in the topical treatment of recurrentherpes labialis. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 1999;88: 700-705.
- **40**) Jain S, Umamaheshwari RB, Bhadra D et al. Ethosomes-A novel vesicular carrier for enhanced transdermal delivery of an anti-HIV agent. Indian J Pharm Sci. 2004;66(1):72-81.
- **41**) Dayan N, TouitouE. Carrier for skin delivery of trihexyphenidyl HCl: Ethosomes vs liposomes. Biomaterials. 2002;21:1879-1885.
- **42**) Touitou E, Godin B, Dayan N. et. al. Intracellular delivery mediated by an ethosomal carrier. Biomaterials. 2001;22: 3053-3059.
- **43**) Esposito E, Menegatti E and Cortesi R. Ethosomes and liposomes as topical vehicles for azelaic acid: a preformulation study. Int J Cos Sci. 2004; 26(5): 270.
- **44)** GodinB, Touitou E, Rubinstein E et. al, A new approach for treatment of deep skin infections by an ethosomal antibiotic preparation: an *in vivo*study, Journal of Antimicrobial Chemotherapy, 2005; 55(6): 989-994.
- **45**) Goldin B, TouitouE, Mechanism of bacitracin permeation enhancement through the skin and cellular membrane from an ethosomal carrier, J. Control. Release, 2004;94: 365-379.
- 46) Touitou E. Composition and methods for intracellular delivery. PCT/IL02/00516, 2002.
- **47**) Touitou E, Drug delivery across the skin, Expert Opin. Biol. Ther., 2002;2: 723-733.
- **48**) Cevc G, Schatzlein A and Blume G. Transdermal drug carriers: Basic properties, optimization and transfer efficiency in case of epicutaneously applied peptides. J Control Release. 1995; 36: 3-16.
- **49**) Nanominox-MS the innovative novelty on the world market of hair tonics, http://www.sinere.com/nanominox-ms_en.html.
- 50) S. Ganguli. Neem: A therapeutic for all seasons, Current Science. 2002;82(11), 1304
- 51) Rohan Rajkumar Patekar, Heena Bholaram Choudhary, Sachin Devidas Rede. Formulation and Evaluation of Ethosomal Gel and Non-ethosomal Gel of S.grandiflora Leaves. Research Journal of Pharmacy and Technology. 2022; 15(3):1029-6. doi: 10.52711/0974-360X.2022.00172
- **52)** Supandeep Singh Hallan, Maddalena Sguizzato, Paolo Mariani, Rita Cortesi, Nicolas Huang, Fanny Simelière, Nicola Marchetti, Markus Drechsler, Tautgirdas Ruzgas and Elisabetta Esposito. Design and Characterization of Ethosomes for Transdermal Delivery of Caffeic Acid, Pharmaceutics 2020;12:740; doi:10.3390/pharmaceutics12080740
- **53**) Mishra R, Shende S, Jain PK, Jain V. Formulation and evaluation of gel containing ethosomes entrapped with tretinoin. JDDT, 2018;.2022;8(5-s):315-21.
- **54**) Hakeeb Ahmed, Syed Sarim Imam, Ameeduz Zafar, Asgar Ali, Mohammed Aqil, Azka Gull. In vitro and preclinical assessment of factorial design based nanoethosomes transgel

- formulation of an opioid analgesic. Artificial Cells, Nanomedicine, and Biotechnology, 2016;44(8):1793-1802.
- 55) Parmar, Priyanka, Ashwani Kumar Mishra and Anupam Kumar Pathak. "Preparation and Evaluation of Ethosomal Gel of Clotrimazole for Fungal Infection by Mechanical Dispersion Method." (2016).
- **56**) Lamsal R, Geethalakshmi A and GubbalaS: Formulation and Evaluation of Gliclazide Ethosomes as a Novel Drug Carrier. Int J Pharm Sci Res 2015; 6(5): 2072-80.doi: 10.13040/IJPSR.0975-8232.6(5).2072-80.
- 57) Kadimpati, Kishore Kumar. Preparation, characterization and evaluation of finasteride ethosomes. International Journal of Drug Delivery. 2016;8:1-16.
- **58**) Sheo Datta Maurya, Sunil Kumar Prajapati, Anish Kumar Gupta Gyanendra Kumar Saxena and Ram Chand Dhakar. Formulation Development and Evaluation of Ethosome of Stavudine. Indian J.Pharm. Educ. Res. 2010;44(1):102-08.
- 59) Barupal AK, Gupta V, Ramteke S. Preparation and Characterization of Ethosomes for Topical delivery of Aceclofenac. Indian J Pharm Sci. 2010 Sep;72(5):582-6. doi: 10.4103/0250-474X.78524. PMID: 21694989; PMCID: PMC3116302.
- **60)** Jain S, Tiwary AK, Sapra B, Jain NK. Formulation and evaluation of ethosomes for transdermal delivery of lamivudine. AAPS PharmSciTech. 2007 Dec 21;8(4):E111. doi: 10.1208/pt0804111. PMID: 18181532; PMCID: PMC2750697.
- **61)** Ciocac. G, lipid vesicles and other colloids as drug carriers on the skin; Advanced drug delivery Reviews 2004; 56:675-711.
- 62) S. Ganguli. Neem: A therapeutic for all seasons, Current Science. 2002;82(11), 1304
- 63) Kanungo D. In Neem., 2nd ed, Randhawa and Parmar, B. S. 1996 77–110.
- 64) Chatterjee A, Pakrashi S. The Treatise on Indian Medicinal Plants, 1994; 3: 76.
- **65**) Bhargava KP, Gupta MB and Gupta GP. Et al. Biological activities of medicinal properties of neem. Indian J. Med. Res., 1970, 58: 724–730.
- **66)** R. Banerjee. Biological activities and medicinal properties of neem (*Azadirachta indica*), current science. 2002; 82. (11), 10.
- **67**) De Jussieu, A., Mem. Mus. Hist. Nat., Paris, 1830, 19: 220
- 68) Pennington TD., Flora Neotropica, New York Botanical Garden, NY, Monogr. 1981.
- **69)** Acessed at URL: http://www.neemfoundation.org/neem-articles/chemistry of neem.
- 70) Chatterjee A, Pakrashi S. The Treatise on Indian Medicinal Plants, 1994; 3: 76.
- 71) Mitra CR. Neem, Indian Central Oilseeds Committee, Hyderabad, 1963: 69–94.
- **72**) Govindachari TR. Chemical and Biological investigations of *Azadirachata indica*. Curr. Sci., 1992, 63: 117–122.
- 73) Anonymous, Doctors Find Neem Good for Skin Diseases, New Delhi Evening News, 1985.
- 74) Kanungo D. In Neem., , 2nd ed, Randhawa and Parmar, B. S. 1996 77–110.
- 75) Bhargava KP, Gupta MB and Gupta GP. Et al. Biological activities of medicinal properties

- of neem. Indian J. Med. Res., 1970, 58: 724-730.
- **76**) Pillai NR, Santhakumari G. Antiinflammatory activity of saponins and other natural products. Planta Med., 1981, 43: 59–63.
- 77) Sithisarn P, Supabphol R, Gritsanapan W Comparison of free radical scavenging activity of Siamese neem tree (Azadirachta indica A. Juss var. siamensis Valeton) leaf extracts prepared by different methods of extraction. Med. Princ. Pract. 2006; 15(3): 219-22.
- **78**) Mukherjee Pulak K. (2001).Quality Control Herbal Drug, Published by Business Horizon, Ist edition, 380.
- 79) Agarwal S.S. (2007). Herbal Drug Technology, Universities Press Pvt.Ltd., Ist edition, 3-7.
- **80**) Harbone J.B., Methods of Plant Analysis Chapter II In: Phytochemical methods: A guide to modern techniques of plant analysis Toppan Company Ltd, Japan., 1973, (1), pp 4 5.
- **81**) A Gupta *et al.*, Formulation and evaluation of topical gel of diclofenac sodium using different polymers, Drug Invention Today 2010, 2(5),250-253
- **82**) Das K, Dang R, Machale UM, Fatepuri S, Formulation and evaluation of herbal gel containing stevia leaves extract, The Pharma Review, 2010, 8(44), 112-118...
- **83**) Prakash RP, Rao R. NG, Soujanya C, Formulation, evaluation and anti-inflammatory activity of topical etoricoxib gel, Asian J. pharmaceutical and clinical research, 2010, 3(2), 126-129.
- **84**) P. Anitha et. al, Ethosomes A noninvasive vesicular carrier for transdermal drug delivery, Anitha *et al.*, Int. J. Rev. Life. Sci., 1(1), 2011, 17-24
- **85**) K Pavan Kumar *et al*, Ethosomes-A Priority in Transdermal Drug Delivery, International Journal of Advances in Pharmaceutical Sciences, 1 (2010) 111-121
- **86**) Jain S, Tiwary AK, Sapra B, Jain NK., Formulation and Evaluation of Ethosomes for Transdermal Delivery of Lamivudine, *AAPS PharmSciTech*. 2007, 8(4): Article 11.