

# Automated scoring as numbers and shape measurement of bacterial colony by using Cell Profiler tool

**Paushali Pal<sup>1\*</sup> N.Sridhar<sup>2</sup> S.Tamilarasi<sup>3</sup>**

*<sup>1,2</sup>Department of Microbiology*

*Sri Shakthi Institute of Engineering and Technology*

*Coimbatore, Tamil Nadu, India*

*<sup>3</sup>Department of Electronics and Communications Engineering*

*Sri Shakthi Institute of Engineering and Technology*

*Coimbatore, Tamil Nadu, India*

*\*Corresponding Author: Dr. Paushali Pal, Ph.D.; Assistant Professor; Phone: 9572631577.....; email: palpaushali@gmail.com...*

## ABSTRACT

The present study examined gray image of bacterial colony by using CP tool (version 2.1.0) for obtaining numbers and area shape (mean radius). An image analysis for bacterial colony (Gray scale) was processed by using CP tool (version 2.1.0). For the input data, CP program and an example “pipeline” file were also downloaded from this tool and incorporated in the present tool. All the output data were obtained through the incorporated image as per several computational simulations and saved as .csv file. In the present study, a total 255 colonies were identified in the image of bacterial colony. Among these colonies, 80 nos. of tiny colonies, 152 nos. of small colonies and 23 nos. of large colonies as per mean radius were scored. It is concluded that scoring and shape measurement of bacterial colony might be performed easily from an image, which is an automated image processing scoring technique.

**Keywords:** Cell Profiler tool, Image processing, Bacterial colony, Automated analysis

## Introduction

An image analysis software namely Cell Profiler (CP), is a freely available, could be capable of handling 100 nos. of images of any cell types from yeast colony to mammalian cells (Carpenter et al., 2006; Bray et al., 2015).

Many experiments in a biology laboratory involve visual inspection, such as examining yeast colonies or growth patches on agar plates, or examining live or stained cells samples by microscopy (Carpenter et al., 2006; Bray et al., 2015).

The image analysis can easily be done after acquiring images and analyzed automatically by using image analysis tool as several advantages such as less tedious, more objective and quantitative, and, no visual error, proper measurement of shape of objects viz. cells, cytoplasm, nuclei, colonies, etc. with a proper input set up (Carpenter et al., 2006; Bray et al., 2015; Talapatra et al., 2016; 2021).

This unit outlines a protocol for the automated counting and analysis of yeast colonies growing on agar plates; however, the methods described can be adapted to a wide variety of biological “objects” and can be used to measure a wide variety of features for each object studied earlier in CP (Carpenter et al., 2006; Lamprecht et al., 2007; Kametsky et al. 2011; Bray et al., 2015; Talapatra et al., 2016; 2021).

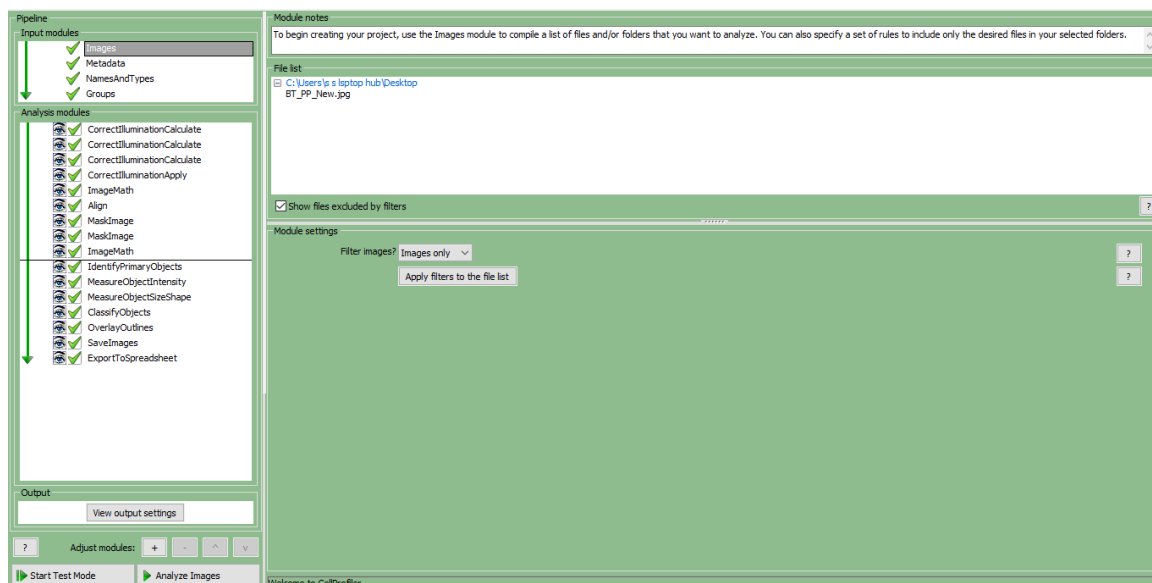
In an earlier study, Bray et al. (2015) evaluated through a pipeline of colour image of yeast colony while the present study examined gray image of bacterial colony by using CP tool (version 2.1.0).

## Materials and Methods

In the present study, an image of bacterial colony (Gray coloured) was processed by using CP (version 2.1.0) tool (Fig 1). This software was downloaded from the designated website (<http://www.cellprofiler.org/download.shtml>). For the input data, CP program and an example “pipeline” file were also downloaded from this tool as per Bray et al. (2015) and incorporated in the present tool. The input interface is depicted in Fig 2. The step of image processing is exhibited in Fig 3A-H. All the output data were obtained through the incorporated image as per several computational simulations and saved as .csv file.



**Fig 1: An input image of bacterial colony used in the present study**

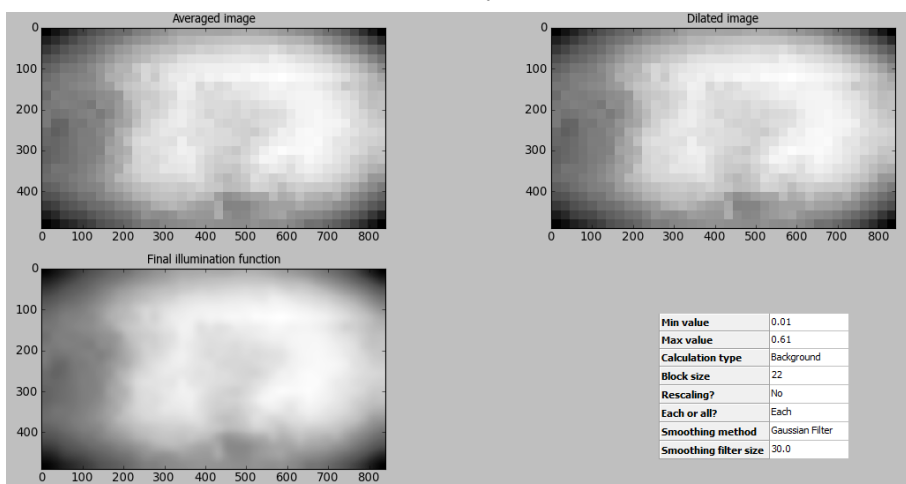


**Fig 2: Input interface of CP tool used for present study**

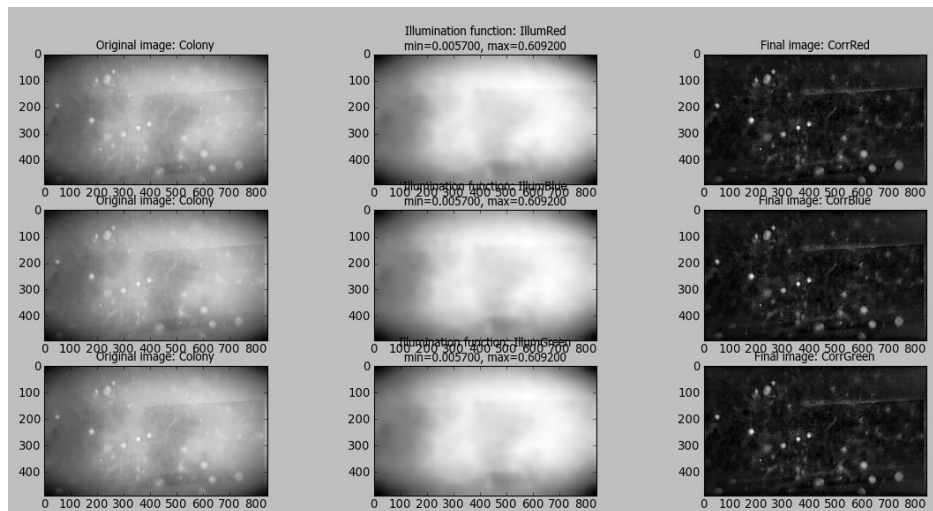


**A. Original image (Gray coloured) retrieved from CP tool**

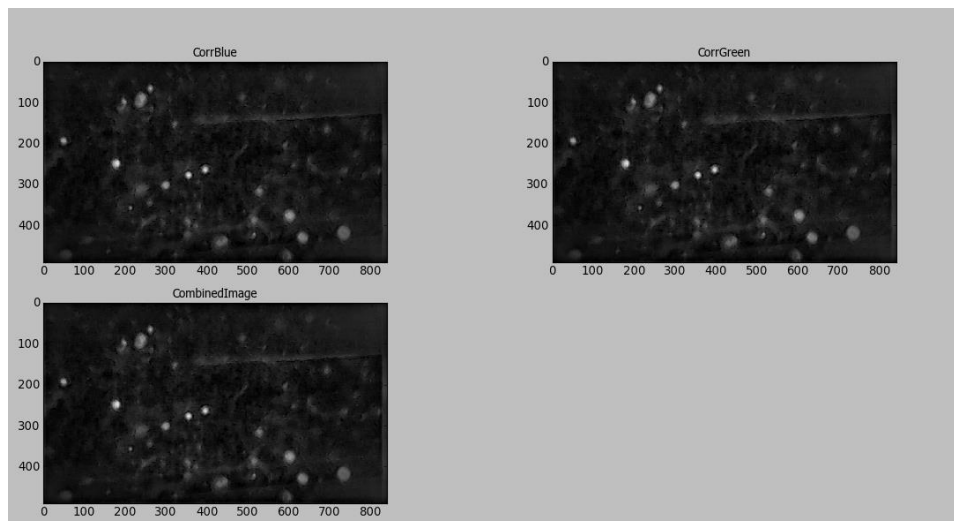
**B.**



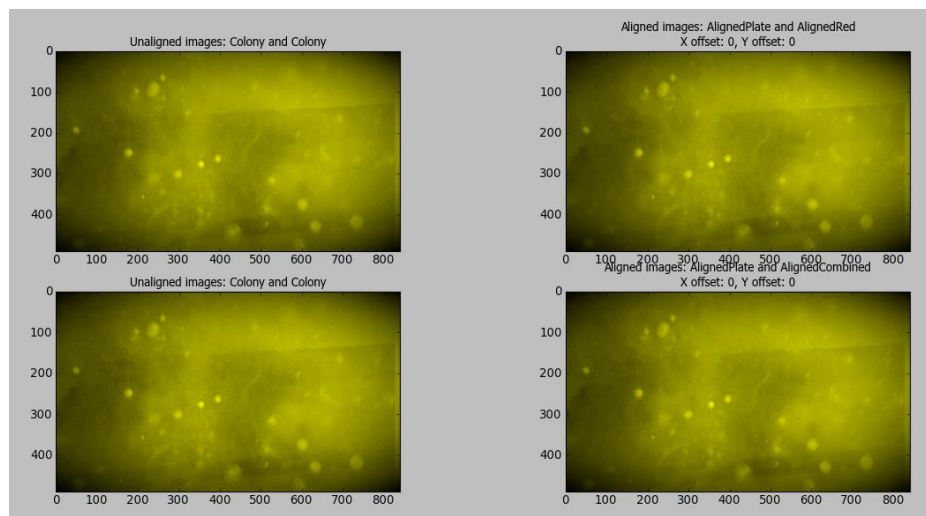
**B. Calculation of correct illumination of image in CP tool**



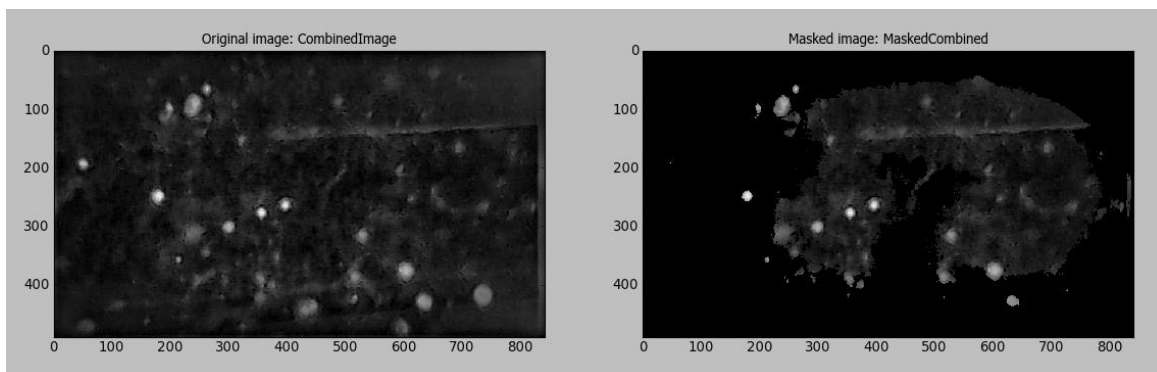
**C. Application of correct illumination of image in CP tool**



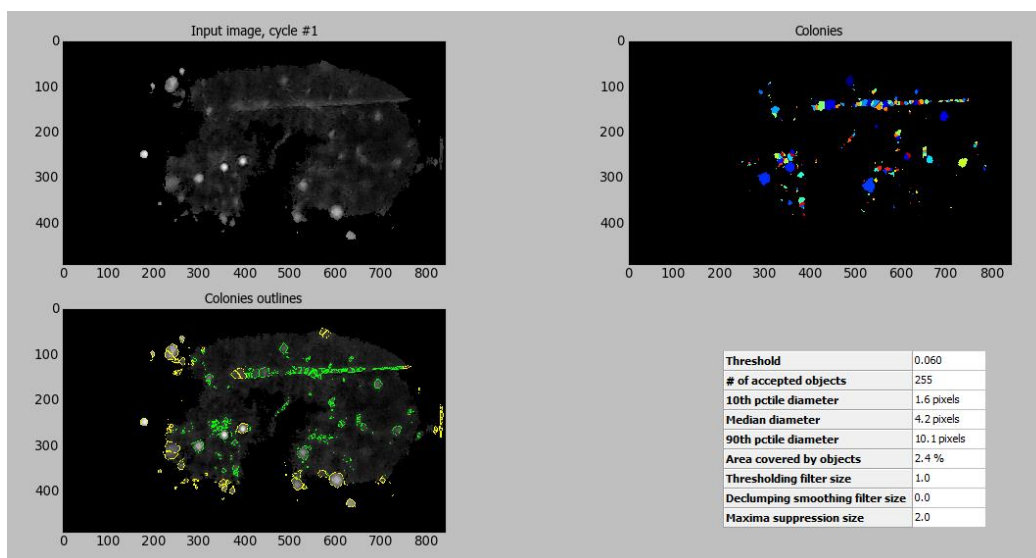
**D. Mathematical calculation of image in CP tool**



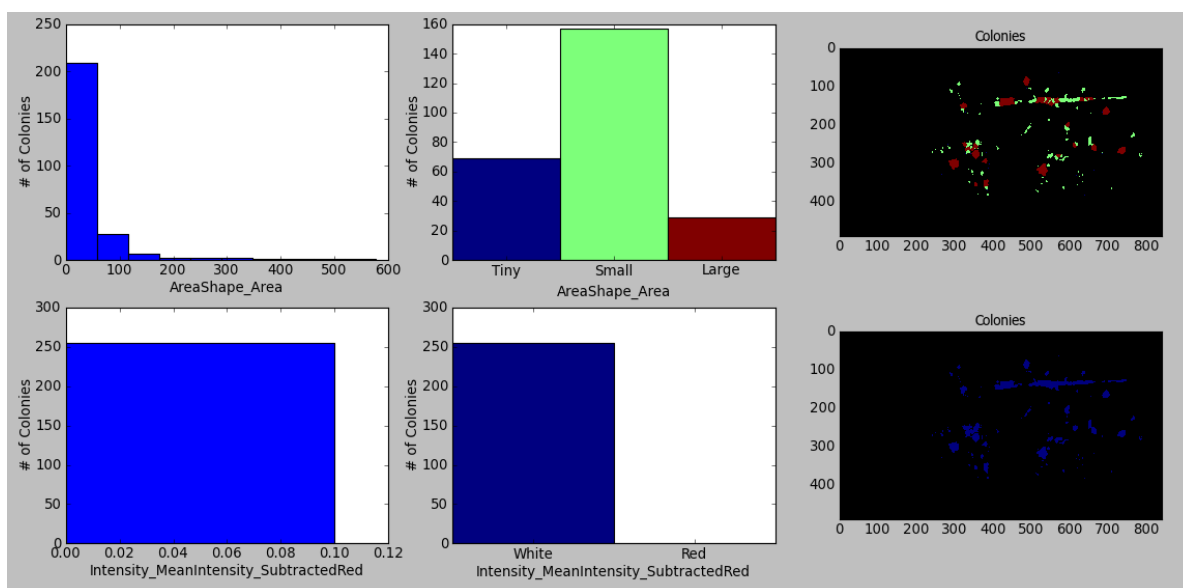
**E. Alignment of image in CP tool**



**F. Masking of image in CP tool**



**G. Identification of primary object of image in CP tool**

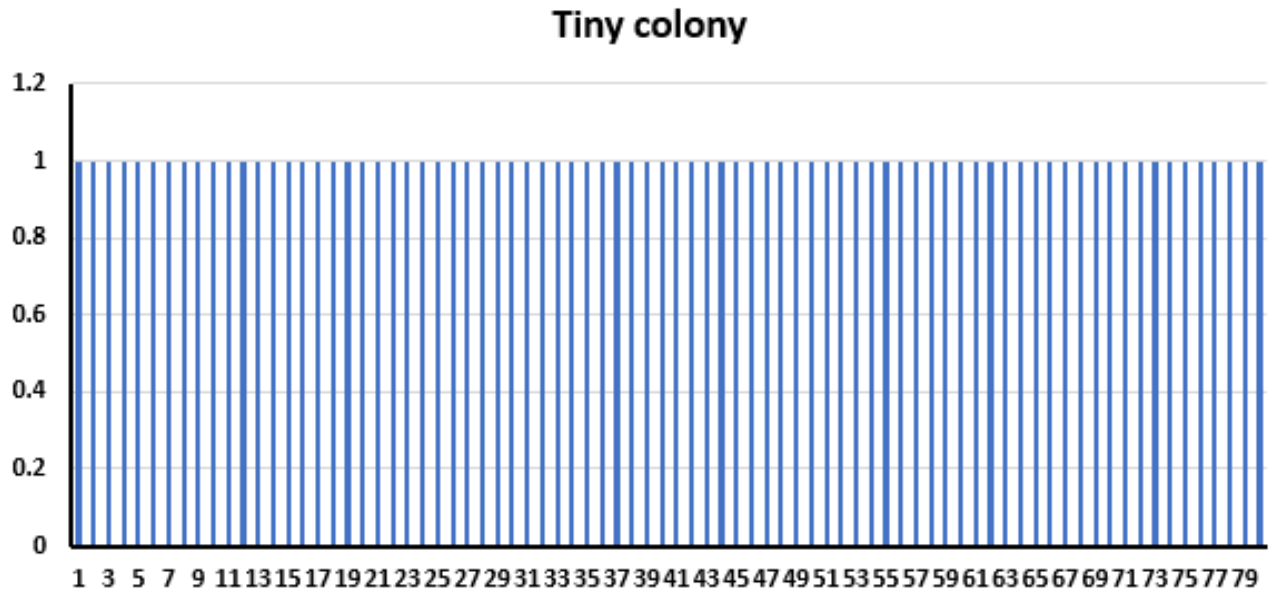


**H. Area shape of object of image in CP tool**

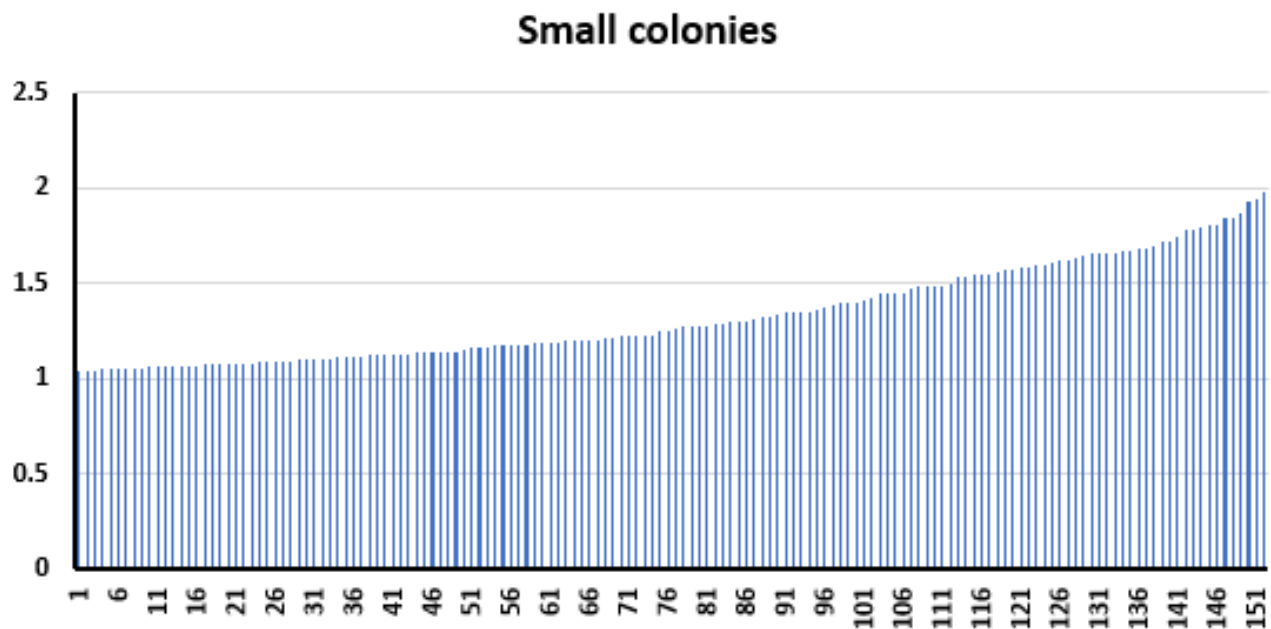
**Fig 3A-H: Different steps of image processing through CP tool**

## Results and Discussion

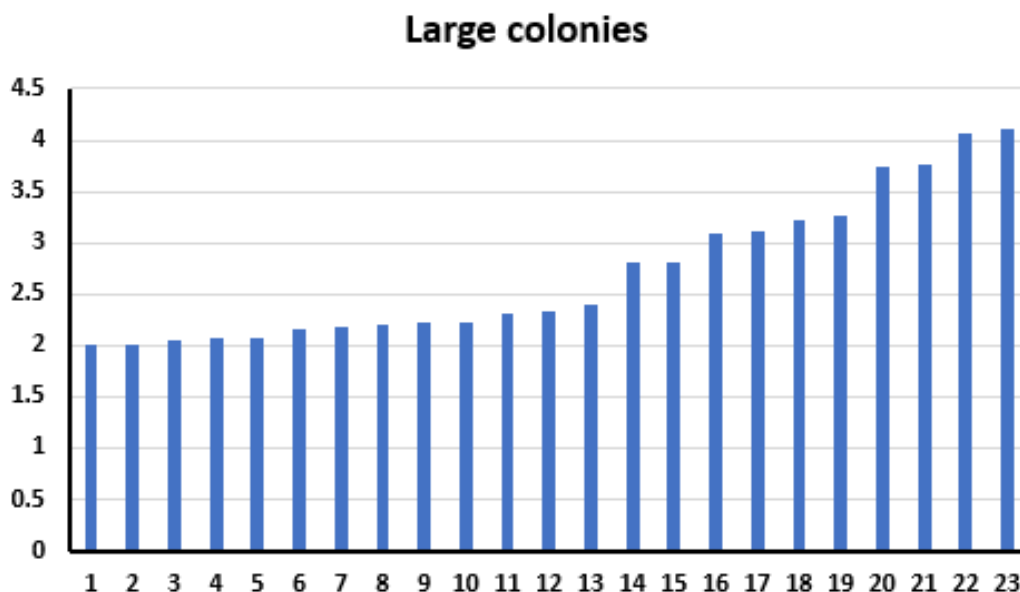
In the present study, a total 255 colonies were identified in the image of bacterial colony. Among these colonies, 80 nos. of tiny colonies, 152 nos. of small colonies and 23 nos. of large colonies as per mean radius were scored (Fig 4-6).



**Fig 4: Area (mean radius) of tiny colonies**



**Fig 5: Area (mean radius) of small colonies**



**Fig 6: Area (mean radius) of large colonies**

In worldwide context, CP tool has already been used by many investigators for identifying a variety of biological processes in different cell types and organisms, viz. yeast colony, different cell types, etc. (Carpenter et al., 2006; Moffat et al., 2006; Bray et al., 2015; Talapatra et al., 2016; 2021). Lamprecht et al. (2007) documented that CP tool has also modified for the measurement of yeast colonies, yeast growth patches, wounds healing assays and tumours quantification, which is supported the present study. According to Chiang et al. (2015), manual counting of bacterial colonies can be time-consuming and imprecise while automated screening is suitable for obtaining faster results.

## Conclusion

In conclusion, present study attempted to screen image (Gray scale) of bacterial colony on the basis of object identification especially numbers and shape (mean radius) by using CP tool, an image-based analysis software. However, the previous study has been studied on colour image of yeast colony, but we performed with the image of bacterial colony (Gray scale). This study can be a beneficial finding in microbiological research to extract the rich information especially numbers and shape of colonies in an image.

## Acknowledgement

Authors convey thanks to the developer of this tool.

## Conflict of interest

Authors declare no conflict of interest.

## References

- M.-A. Bray, M.S. Vokes, A.E. Carpenter, Using CellProfiler for automatic identification and measurement of biological objects in images, *Curr. Protoc. Mol. Biol.* 109 (2015) 14.17.1-14.17.13.
- A.E. Carpenter, T.R. Jones, M.R. Lamprecht, C. Clarke, I.H. Kang, O. Friman, D.A. Guertin, J.H. Chang, R.A. Lindquist, J. Moffat, P. Golland, D.M. Sabatini, CellProfiler: image analysis software for identifying and quantifying cell phenotypes, *Genome Biol.* 7 (2006) R100.
- P-JChiang, M-JTseng, Z-SHe, C-H. Li. Automated counting of bacterial colonies by image analysis. *Journal of Microbiological Methods*, 108(2015), 74-82.
- M. R. Lamprecht, D.M. Sabatini, A.E. Carpenter, CellProfiler: free, versatile software for automated biological image analysis, *Biotechniques* 42 (2007) 71-75.
- J. Moffat, D.A. Grueneberg, X. Yang, S.Y. Kim, A.M. Kloepper, G. Hinkle, B. Piqani, T. M. Eisenhaure, B. Luo, J.K. Grenier, A.E. Carpenter, S.Y. Foo, S.A. Stewart, B.R. Stockwell, N. Hacohen, W.C. Hahn, E.S. Lander, D.M. Sabatini, D.E. Root, A lentiviral RNAi library for human and mouse genes applied to an arrayed viral high-content screen, *Cell* 124 (2006) 1283-1298.
- Talapatra SN, Mitra P, Swarnakar S, Morphology and phenotype of peripheral erythrocytes of fish: A rapid screening of images by using software. *International Letters of Natural Sciences* 54 (2016) 27-41
- S. N. Talapatra, R. Chaudhuri, S. Ghosh, CellProfiler and WEKA tools: Image analysis for fish erythrocytes shape and machine learning model algorithm accuracy prediction of dataset. *World Scientific News*, 154,(2021) 101-116.