

Evaluation of the Potential Cardio Protective Effects of *Sechium edule* (Chayote Fruit) Extract on Male Wistar Rats Subjected to Cadmium-Induced Toxicity

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Abstract:

Cardio toxicity, a major health concern linked to heavy metal accumulation, particularly Cadmium (Cd) chloride, induces oxidative stress and disrupts heart function. This study investigated the potential ameliorative effects of ethanol extracts from *Sechium edule* (S.E.) fruits on Cd-induced oxidative stress in male Wistar rats. Daily administration of Cd (4mg/kg) for 5 weeks resulted in oxidative stress, evidenced by elevated lipid peroxidation and altered antioxidant status. Treatment with S.E. extract (200mg/kg and 400mg/kg) demonstrated significant reductions in serum total cholesterol, triglycerides, and low-density lipoprotein cholesterol, coupled with increased high-density lipoprotein cholesterol. The extract reversed Cd-induced elevations in serum ALT, lipid peroxidation, & improved enzymatic and non-enzymatic antioxidant levels in the heart. S.E. extract also regulated serum Na⁺ and K⁺ ion levels and improved abnormal ECG parameters induced by Cd, indicating enhanced heart function. Histopathological analysis revealed marked improvements in the structural features of the heart, kidney, and liver in S.E. extract-treated groups compared to the Cd control group. These findings suggest that *Sechium edule* may hold promise as a novel treatment for cardio toxicity resulting from cadmium exposure.

Keywords: Cardiotoxicity, Cadmium, *Sechium edule*, cholesterol, triglycerides.

1. Introduction:

Cardio toxicity, often induced by heavy metal exposure like cadmium (Cd), poses severe health risks Cd, ^[1] a non-essential heavy metal, is highly toxic and associated with various diseases. ^[2-5] Its mechanism involves oxidative tissue damage, reactive oxygen species production, and disruption of cellular functions. Herbal medicines, with known cardiovascular benefits, offer a potential safer alternative. ^[6-8]

The study focuses on *Sechium edule* (S.E.), a plant widely used in Ayurvedic medicine, renowned for its diverse phytoconstituents. With significant diuretic, nephroprotective and anti-inflammatory etc. ^[9-12] properties, S.E. is investigated for its potential in countering cadmium (Cd)-induced cardiotoxicity in Wistar rats. Cd, a major public health concern, infiltrates ecosystems through various sources, posing severe health risks. Cd toxicity, ^[14,15] associated with oxidative damage, impacts organs and tissues, leading to cardiomyopathy and heart failure. Synthetic drugs carry risks, emphasizing the need for safer alternatives. Herbal medicines, including S.E., gain importance for cardiovascular health. This study explores S.E.'s cardioprotective effects, providing insights into its therapeutic potential against Cd-induced cardiovascular harm. ^[16-19]

2. Materials & Methods:

2.1. Plant material:

Fruits of *Sechium edule* (S.E.) were sourced from Spencer, Haldia, and the local market in Haldia, West Bengal. Taxonomic authentication at the Regional Research Institute (Ayurveda), Bangalore, assigned the voucher specimen reference number (RRCBI/MCW/7/2008) for future use.

2.2. Preparation of the extract:

Sechium edule (S.E.) fruit extract preparation involved cleaning, drying, and powdering. The coarse powder underwent petroleum ether treatment, maceration in ethanol, and distillation. The resulting ethanolic extract, a dark mass, was stored for further use, ensuring removal of impurities and preservation of bioactive compounds. ^[20-23]

2.3. Phytochemical investigation:

Preliminary phytochemical screening of the ethanolic extract of *Sechium edule* (S.E.) fruits done for identification of the presence of alkaloids, phenolics, flavonoids, saponins, carotenoids, carbohydrates, and glycosides.

2.4. Experimental animals:

Male albino Wistar rats (180-220g) will be used, housed in controlled conditions with a 12h light-dark cycle, sanitized cages, and standard diet access. After a week of acclimatization, experiments following CPCSEA guidelines and IAEC approval will commence. ^[24]

2.5. Drugs and Chemicals:

Cadmium chloride, vitamin C, ketamine HCl, and assay kits for total cholesterol, triglycerides, SGOT, SGPT, ALP, total protein, serum urea, sodium, potassium ions, and creatinine will be purchased. All other chemicals used will be of analytical reagents grade from commercial sources. ^[25, 26]

2.6. Oral Toxicity Test:

According to OECD 423 guidelines, Wistar rats, after overnight fasting, will receive a single oral dose (2000 mg/kg b.w.) of S.E. fruit extract. Food will be withheld for 3-4 hrs post-administration. Rats will be observed for 30 min initially, periodically for the first 24 hrs (with emphasis on the first 4 hrs), and daily for 14 days. Observations will cover skin, fur, eyes, mucous membranes, respiratory and circulatory functions, autonomic responses, and central nervous system parameters. Mortality, if any, will be recorded over the two-week period. [27,28]

2.7. Induction of Cardiotoxicity:

Male Wistar rats, rendered cardiotoxic, received oral Cd (4mg/kg) for 5 weeks. Cd (90 mg), dissolved in sterile water (30 ml), was administered orally (0.25 ml) via an intragastric tube throughout the 5-week period.

2.8. Experimental design:

Male Wistar rats, post one-week acclimatization, were divided into five groups (n=6). Pre-treatment with S.E. fruit extract (200 and 400 mg/kg; p.o.) was done for 3 weeks, followed by Cd administration (4 mg/kg; p.o.) for 5 weeks. Groups received clean water, Cd, Vit-C + Cd, S.E. (200 mg/kg) + Cd, and S.E. (400 mg/kg) + Cd, respectively. Treatments were administered orally using an intra-gastric tube for 56 days. On the 56th day, after a 24-hour fast, blood was collected for biochemical analysis, and animals were euthanized for further assessments. [29]

3. Result:

3.1. Preliminary phytochemical investigation:

In preliminary phytochemical analysis of an alcoholic fruit extract S.E showed the presence of carbohydrates, proteins, flavonoids, saponins and tannins.

3.2. Acute oral toxicity test:

In toxicity study, no mortality occurred during 48 hours of observation with the selected dose 2000mg/kg p.o. As the ethanolic extract of S.E fruits were found to be tolerated up to a dose level of 2000mg/kg p.o. The extract was considered to be safe and dose ranges 400mg/kg p.o. (1/5th of the extract) and 200mg/kg p.o. (1/10th of the extract) was selected for the present study.

3.3. In Vivo Activity:

3.3.1. Effect of alcoholic fruit extract of S.E on lipid profile in Cd- induced cardiotoxic rats.

A. Effect of administration of Cd (4mg/kg; p.o.) on lipid profiles in rats.

Total cholesterol, Triglyceride, high density lipoprotein (HDL-C), low density lipoprotein (LDL-C) and very low-density lipoprotein (VLDL-C) levels in vehicle treated rats (Group-I) were found to be 64.09 ± 2.46 , 46.81 ± 1.10 , 36.79 ± 1.68 , 20.67 ± 0.43 and 8.56 ± 0.22 respectively. In Cd (4mg/kg) treated (Group II) the serum level of total cholesterol, Triglyceride, HDL-C, LDL-C and VLDL-C were found to be 79.55 ± 1.41 , 117.59 ± 3.83 , 15.75 ± 0.73 , 38.04 ± 0.82 and 23.53 ± 0.76 respectively. Thus, Cd significantly increased the levels of total cholesterol, Triglyceride, LDL-C and VLDL-C and decreased HDL-C level.

B. Effect of administration of Vit-C (50mg/kg; p.o.) on lipid profiles in Cd-induced cardiotoxic rats.

Total cholesterol, triglyceride, HDL-C, LDL-C and VLDL-C levels in Vit-C (50mg/kg) treated rats (Group III) were found to be 66.59 ± 1.26 , 63.03 ± 1.74 , 28.53 ± 0.68 , 25.13 ± 0.23 and 12.61 ± 0.35 respectively. The above values have significantly decreased the levels of total cholesterol, triglyceride, LDL-C, and very VLDL-C and HDL-C value have increased significantly in group-III when compared to group-II.

C. Effect of administration of lower dose of alcoholic fruit extract of S.E (200mg/kg; p.o.) on lipid profiles in Cd-induced cardiotoxic rats.

Total cholesterol, triglyceride, HDL-C, LDL-C, and VLDL-C levels in lower dose of S.E. (200mg/kg) treated rats (Group IV) were found to be 73.33 ± 1.94 , 68.59 ± 1.88 , 26.76 ± 2.19 , 32.23 ± 0.09 and 114.12 ± 0.37 respectively. The above values have significantly decreased the levels of total cholesterol, triglyceride, LDL-C and VLDL-C values and HDL-C value have increased significantly when compared to group-II.

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) on lipid profiles in Cd-induced cardiotoxic rats.

Total cholesterol, triglyceride, HDL-C, LDL-C and VLDL-C levels in higher dose of S.E. (400mg/kg) treated rats (Group V) were found to be 71.37 ± 1.46 , 64.49 ± 2.80 , 32.51 ± 2.29 , 26.61 ± 0.51 and 13.49 ± 0.55 respectively. The above values have significantly decreased the levels of total cholesterol, triglyceride, LDL-C and VLDL-C values and HDL-Cs value have increased significantly when compared to group-II.

Table 1: Effect of alcoholic fruit extract of S.E on lipid profile Cd-induced cardiotoxic rats.

Groups	Treatment	Total cholesterol (mg/dL)	Triglyceride (mg/dL) s	HDL-C (mg/dL)	LDL-C (mg/dL)	VLDL-C (mg/dL)
I	Control	64.09±2.46	46.81±1.10	36.79±1.67	20.67±0.43	8.562±0.22
II	Cd (4mg/kg; p.o.)	79.55±1.41 ^{##}	117.59±3.83 ^{##}	15.75±0.73 ^{##}	38.04±0.82 ^{##}	23.53±0.76 ^{##}
III	Vit-C (50 mg/kg; p.o.) +Cd	66.59±1.26 ^{**}	63.03±1.74 ^{**}	28.53±0.68 ^{**}	25.13±0.23 ^{**}	12.61±0.35 ^{**}
IV	S .E. (200 mg/kg;p.o.) +Cd	73.33±1.94 ^{**}	68.59±1.88 ^{**}	26.76±2.19 ^{**}	32.23±0.09 ^{**}	114.12±0.37 [*]
V	S .E. (400 mg/kg; p.o.) +Cd	71.37±1.46 ^{**}	64.49±2.80 ^{**}	32.51±2.29 ^{**}	26.61±0.15 [*]	13.49±0.55 ^{**}

Values are expressed as Mean ± SEM, n=6; ^{##} P<0.01 considered statistically significant as compare to normal control group; ^{***} P<0.001, ^{**} P<0.01 and ^{*}P<0.05 considered statistically significant when compared to Cd treated group.

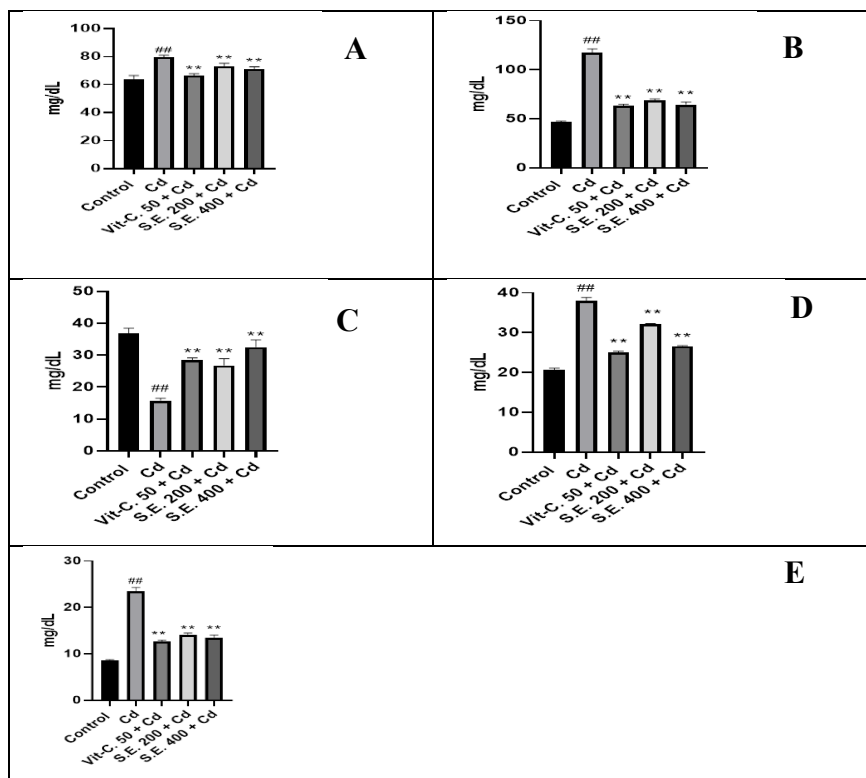


Figure 1: Effect of alcoholic fruit extract of S.E. on A. cholesterol B. Triglyceride C. HDL-C D. LDL-C E. VLDL-C level in Cd-induced cardiotoxic rats.

3.3.2. Effect of alcoholic fruit extract of S.E. on serum lactate dehydrogenase (LDH), creatine phosphokinase (CPK), Serum glutamic oxaloacetic transaminase (SGOT), SGPT, and total protein levels pyruvic transaminase (SGPT) and total protein levels in Cd- induced cardiotoxic rats.

A. Effect of administration of Cd (4mg/kg; p.o.) on serum cardiac markers in rats.

Serum LDH, CPK, SGOT, SGPT and total protein levels in vehicle treated rats (Group-I) were found to be 139.39±6.412, 32.98±3.08, 25.09±2.55, 26.31±0.99, and 7.12±0.39 respectively. In Cd (4mg/kg) treated rats (Group II) the LDH, CPK, SGOT, SGPT and total protein were found to be 455.60±41.11, 62.08±3.25, 180.20±11.72, 96.40±3.94 and 5.04±0.133 respectively. Thus, in Cd treated rats (Group-II) significantly increased the levels of serum LDH, CPK, SGOT and SGPT and decreased total protein level.

B. Effect of administration of Vitamin-C (50mg/kg; p.o.) on serum cardiac marker in Cd-induced cardiotoxic rats.

Serum LDH, CPK, SGOT, SGPT and total protein levels in Vit-C (50mg/kg) treated rats (Group-III) were found to be 150.36±12.01, 36.29±1.64, 117.88±2.75, 58.96±1.94 and 6.81±0.12 respectively. The above values have significantly decreased the level of LDH, CPK, SGOT and SGPT and increased total protein level when compared to group-II.

C. Effect of administration of lower dose of alcoholic fruit extract of S.E (200mg/kg; p.o.) on serum cardiac marker in Cd-induced cardiotoxic rats.

Serum LDH, CPK, SGOT, SGPT and total protein levels in rats treated with alcoholic fruit extract of S.E. (200mg/kg) treated rats (Group-IV) were found to be 246.09±5.91, 39.81±2.49, 128.76±4.95, 77.47±1.05 and 6.20±0.11 respectively. The above values have significantly decreased the level of LDH, CPK, SGOT and SGPT and increased total protein level when compared to group-II.

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) on serum cardiac marker in Cd-induced cardiotoxic rats.

Serum LDH, CPK, SGOT, SGPT and total protein levels in alcoholic fruit extract of S.E. (400mg/kg) treated rats (Group-V) were found to be 196.64±4.48, 38.56±2.58, 124.01±2.99, 64.30±2.23 and 6.23±0.09 respectively. The above values have significantly decreased Serum LDH, CPK, SGOT and SGPT and increased total protein level when compared to group-II.

Table 2: Effect of alcoholic fruit extract of S.E. on biochemical parameter LDH, CPK, SGOT, SGPT and total protein in Cd-induced cardiotoxic rats.

Groups	Treatment	LDH (U/L)	CPK (U/L)	SGOT (IU/L)	SGPT (IU/L)	Total protein (gm/dL)
I	Control	139.39±6.41	32.98±3.08	25.09±2.55	26.31±0.99	7.12±0.39
II	Cd (4mg/kg;p.o)	455.60±41.11 ^{##}	62.08±3.25 ^{##}	180.20±11.72 ^{##}	96.40±3.94 ^{##}	5.04±0.13 [#]
III	Vit-C (50mg/kg; p.o) +Cd	150.36±12.01 ^{**}	36.29±1.64 ^{**}	117.88±2.75 [*]	58.96±1.94 ^{**}	6.81±0.12 [*]
IV	S.E. (200mg/kg;p.o) +Cd	246.09±5.91 ^{**}	39.81±2.49 ^{**}	128.76±4.95 [*]	77.47±1.05 ^{**}	6.20±0.11 [*]
V	S.E. (400mg/kg;p.o) +Cd	196.64±4.48 ^{**}	38.56±2.58 ^{**}	124.01±2.99 [*]	64.30±2.23 ^{**}	6.23±0.09 [*]

Values are expressed as mean \pm SEM, n=6; $^{###}P < 0.01$ considered statistically significant as compare to normal control group; $^{***}P < 0.001$, $^{**}P < 0.01$ and $^{*}P < 0.05$ considered statistically significant when compared to Cd group.

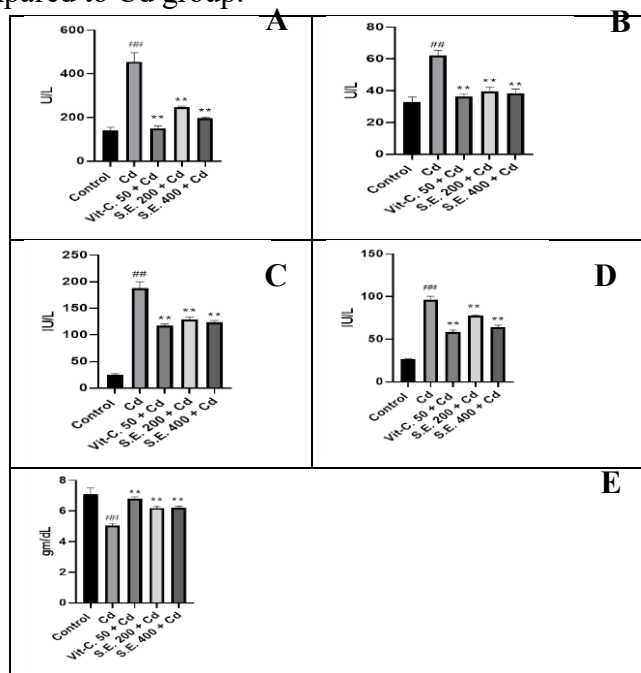


Figure 2: Effect of alcoholic fruit extract of S.E. on A. LDH, B. CPK, C. SGOT, D. SGPT, E. Total protein level in Cd-induced cardiotoxic rats.

3.3.3. Effect of alcoholic fruit extract of S.E. on blood urea, creatinine, alkaline phosphatase (ALP), sodium (Na^+) and potassium (K^+) levels in Cd-induced cardiotoxic rats.

A. Effect of administration of Cd (4mg/kg; p.o.) on serum level of cardiotoxic rats.

Blood urea, creatinine, ALP, Na^+ and K^+ levels in vehicle treated rats (Group-I) were found to be 24.06 ± 1.03 , 0.174 ± 0.007 , 14.28 ± 1.40 , 45.32 ± 1.90 and 3.95 ± 0.13 respectively. In Cd (4mg/kg) treated rats (Group II) the serum levels of urea, creatinine, ALP, Na^+ and K^+ were found to be 45.16 ± 2.04 , 0.752 ± 0.03 , 75.84 ± 6.42 , 141.69 ± 15.62 and 6.06 ± 0.61 respectively. Thus, in Cd treated rats (Group-II) significantly increased the levels of blood urea, serum creatinine, ALP, Na^+ and K^+ .

B. Effect of administration of Vit-C (50mg/kg; p.o.) on serum levels in Cd-induced cardiotoxic rats.

Blood urea, creatinine, ALP, Na^+ and K^+ levels in Vit-C (50mg/kg) treated rats (Group-III) were found to be 28.43 ± 1.55 , 0.401 ± 0.018 , 16.95 ± 1.63 , 65.08 ± 1.24 and 3.77 ± 0.51 respectively. The above values have significantly decreased blood urea, creatinine, ALP, Na^+ and K^+ level when compared to group II.

C. Effect of administration of lower dose of alcoholic fruit extract of S.E. (200mg/kg; p.o.) on serum levels in Cd-induced cardiotoxic rats.

Blood urea, creatinine, ALP, Na^+ and K^+ levels in lower dose of ethanolic extract of fruits S.E. (200mg/kg) treated rats (Group-IV) were found to be 38.38 ± 1.67 , 0.324 ± 0.037 , 19.03 ± 1.51 , 87.27 ± 2.12 and 4.26 ± 0.26 respectively. The above values have significantly decreased for blood urea, creatinine, ALP, Na^+ and K^+ level when compared to group-II.

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) on serum levels in Cd-induced cardiotoxic rats.

Blood urea, creatinine, ALP, Na⁺ and K⁺ levels in higher dose of ethanolic extract of fruits S.E. (400mg/kg) treated rats (Group-IV) were found to be 35.63±1.31, 0.314±0.019, 17.25±1.05, 82.98±2.52 and 3.53±0.18 respectively. The above values have significantly decreased for blood urea, creatinine, ALP, Na⁺ and K⁺ level when compared to group-II.

Table 3: Effect of alcoholic fruit extract of S.E. on biochemical parameter Blood urea, serum creatinine, alkaline phosphatase, sodium (Na⁺) and potassium (K⁺) in Cd- induced cardiotoxic rats.

Groups	Treatment	Urea (mg/dL)	Creatinine (mg/dL)	ALP (KA/dL)	Na ⁺ (mmol/L)	K ⁺ (mmol/L)
I	Control	24.06±1.03	0.174±0.007	14.28±1.40	45.32±1.90	3.95±0.13
II	Cd (4mg/kg; p.o.)	45.16±2.04 ^{##}	0.752±0.029 ^{##}	75.84±6.42 ^{##}	141.69±15.62 ^{##}	6.06±0.61 ^{##}
III	Vit-C (50mg/kg; p.o.) +Cd	28.43±1.55 ^{**}	0.401±0.018 ^{**}	16.95±1.63 ^{**}	65.08±1.24 ^{**}	3.77±0.51 ^{**}
IV	S.E. (200mg/kg; p.o.) +Cd	38.38±1.67 [*]	0.324±0.037 ^{**}	19.03±1.51 ^{**}	87.27±2.12 ^{**}	4.26±0.26 ^{**}
V	S.E. (400mg/kg; p.o.) +Cd	35.63±1.31 ^{**}	0.314±0.019 ^{**}	17.25±1.05 ^{**}	82.98±2.52 ^{**}	3.53±0.18 ^{**}

Values are expressed as mean ± SEM, n=6; ^{##} P<0.01 considered statistically significant as compare to normal control group; ^{***} P<0.001, ^{**} P<0.01 and ^{*} P<0.05 considered statistically significant when compared to Cd group.

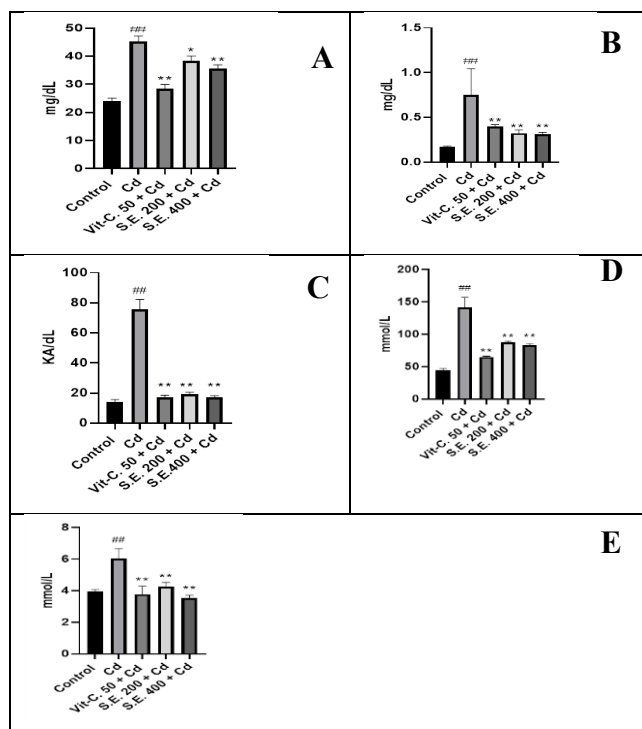


Figure 3: Effect of alcoholic fruit extract of S.E. on A. Urea, B. Creatinine, C. ALP, D. Na⁺, E. K⁺ level in Cd-induced cardiotoxic rats.

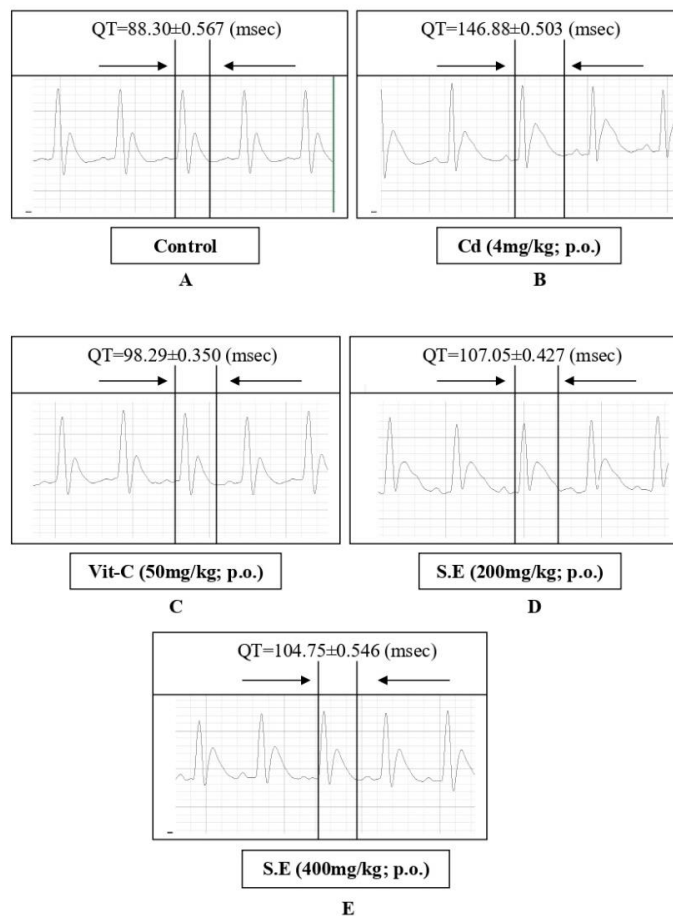


Figure 4: A-E Effect of S.E. on ECG in QT interval in Cd-induced cardiotoxic rats.

3.3.4. Effect of alcoholic fruit extract of S.E. on ECG in QT interval of heart in Cd-induced cardiotoxic rats.

A. Effect of administration of Cd (4mg/kg; p.o.) on QT interval in rats.

QT interval in vehicle treated rats (Group-I) were found to be 82.93 ± 0.392 , 85.26 ± 0.445 and 88.30 ± 0.567 respectively on day 1st, 30th and 56th day. In Cd (4mg/kg) treated rat (Group-II) the QT interval was found to be 87.33 ± 0.567 , 122.45 ± 0.627 and 146.88 ± 0.503 respectively on day 1st, 30th and 56th day. Thus, in Cd treated rats (Group-II) the QT interval was significantly increased over a period of 56th days when compared to (Group-I).

B. Effect of administration of Vit-C (50mg/kg; p.o.) on QT interval in Cd-induced cardiotoxic rats.

QT interval in rats, treated with Vit-C (50mg/kg) in (group-III) were found to be 86.34 ± 0.336 , 91.40 ± 0.533 and 98.29 ± 0.0350 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

C. Effect of administration of lower dose of alcoholic fruit extract of S.E. (200mg/kg; p.o.) in Cd-induced cardiotoxic rats.

QT interval in rats treated with lower dose of alcoholic fruit extract of S.E. (200mg/kg) in (Group-IV) were found to be 82.64 ± 0.455 , 97.22 ± 0.456 and 107.05 ± 0.427 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) in Cd-induced cardiotoxic rats.

QT interval in rats treated with higher dose of alcoholic fruit extract of S.E. (400mg/kg) in (Group-V) were found to be 87.51 ± 0.321 , 96.50 ± 0.601 and 104.75 ± 0.546 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

Table 4: Effect of alcoholic fruit extract of S.E. on ECG in QT interval of heart in Cd-induced cardiotoxic rats.

Groups	Treatment	QT interval (msec)		
		Day 1	Day 30	Day 56
I	Control	82.93±0.392	85.26±0.445	88.30±0.567
II	Cd (4mg/kg; p.o.)	87.33±0.567 [#]	122.45±0.627 ^{##}	146.88±0.503 ^{##}
III	Vit-C (50mg/kg; p.o.) + Cd	86.34±0.336	91.40±0.533 [*]	98.29±0.350 [*]
IV	S.E (200 mg /kg; p.o.) + Cd	82.64±0.455 [*]	97.22±0.456 [*]	107.05±0.427 ^{**}
V	S.E (400 mg /kg; p.o.) + Cd	87.51±0.321	96.50±0.601 [*]	104.75±0.546 ^{**}

Values are expressed as mean ± SEM, n=6; ^{##} P<0.01 considered statistically significant as compare to normal control group; ^{***} P<0.001, ^{**} P<0.01 and ^{*} P<0.05 considered statistically significant when compared to Cd group.

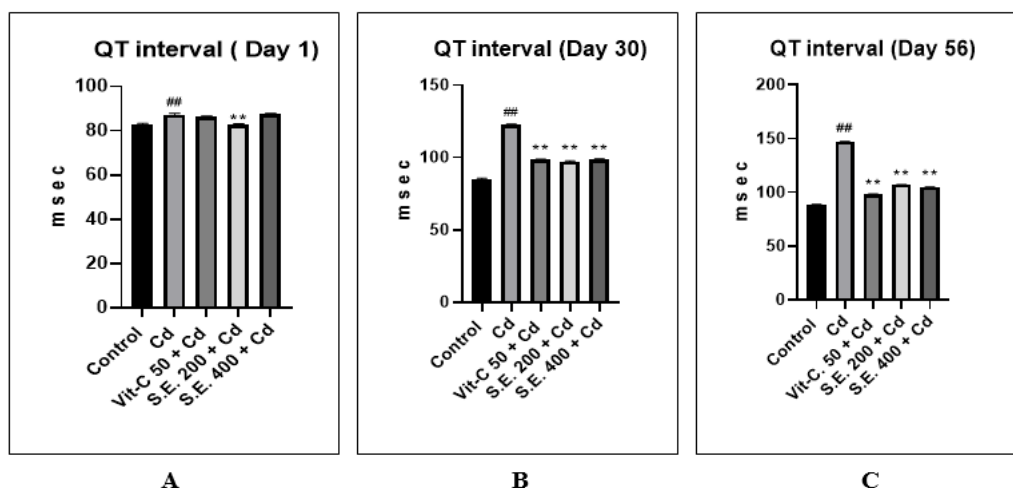


Figure 4: A-C Effect of alcoholic fruit extract of S.E. on ECG in QT interval of heart in Cd-induced cardiotoxic rats.

3.3.5. Effect of alcoholic fruit extract of S.E. on ECG in QTc interval in Cd- induced cardiotoxic rats:

A. Effect of administration of cadmium (4mg/kg; p.o.) on QTc interval in rats.

QTc interval in vehicle treated rats (Group-I) were found to be 71.61 ± 0.15 , 71.90 ± 0.20 and 72.60 ± 0.36 respectively on day 1st, 30th and 56th day. In Cd (4mg/kg) treated rat (Group-II) the QTc interval were found to be 73.88 ± 0.35 , 81.98 ± 0.51 and 88.36 ± 0.21 respectively on day 1st, 30th and 56th day. Thus, in Cd treated rats (Group-II) the QTc interval was significantly increase over a period of 56th days when compared to (Group-I).

B. Effect of administration of Vit-C (50mg/kg; p.o.) on QTc interval in Cd- induced cardiotoxic rats: QTc interval in rats treated with Vit-C (50mg/kg) in (group-III) were found to be 73.37 ± 1.6 , 72.23 ± 0.99 and 74.62 ± 0.25 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

C. Effect of administration of lower dose of alcoholic fruit extract of S.E. (200mg/kg; p.o.) in Cd-induced cardiotoxic rats.

QTc interval in rats treated with lower dose of alcoholic fruit extract of S.E. (200mg/kg) in (Group-IV) were found to be 70.86 ± 0.16 , 75.25 ± 0.23 and 78.81 ± 0.37 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) in Cd-induced cardiotoxic rats.

QTc interval in rats treated with higher dose of alcoholic fruit extract of S.E. (400mg/kg) in (Group-V) were found to be 73.95 ± 1.4 , 76.78 ± 0.27 and 77.42 ± 0.59 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

Table 5: Effect of alcoholic fruit extract of S.E. on ECG in QTc interval of heart in Cd-induced cardiotoxic rats.

Groups	Treatment	QTc (msec)		
		1 st day	30 th day	56 th day
I	Control	71.61 ± 0.15 5	71.90 ± 0.20	72.60 ± 0.36
II	Cd (4mg/kg; p.o)	73.88 ± 0.35 5 ^{##}	81.98 ± 0.51 [#]	88.36 ± 0.21 [#]
III	Vit-C (50mg/kg; p.o) + Cd	73.37 ± 0.1 6	72.23 ± 0.99 [*]	74.62 ± 0.25 [*]
IV	S.E (200 mg/kg; p.o) + Cd	70.86 ± 0.1 6 ^{**}	75.25 ± 0.23 [*]	78.81 ± 0.37 [*]
V	S.E (400 mg/kg; p.o) + Cd	73.95 ± 0.1 4	76.78 ± 0.27 [*]	77.42 ± 0.59 [*]

Values are expressed as mean \pm SEM, n=6; $##$ $P < 0.01$ considered statistically significant as compare to normal control group; $***$ $P < 0.001$, $**$ $P < 0.01$ and $*$ $P < 0.05$ considered statistically significant when compared to cadmium group.

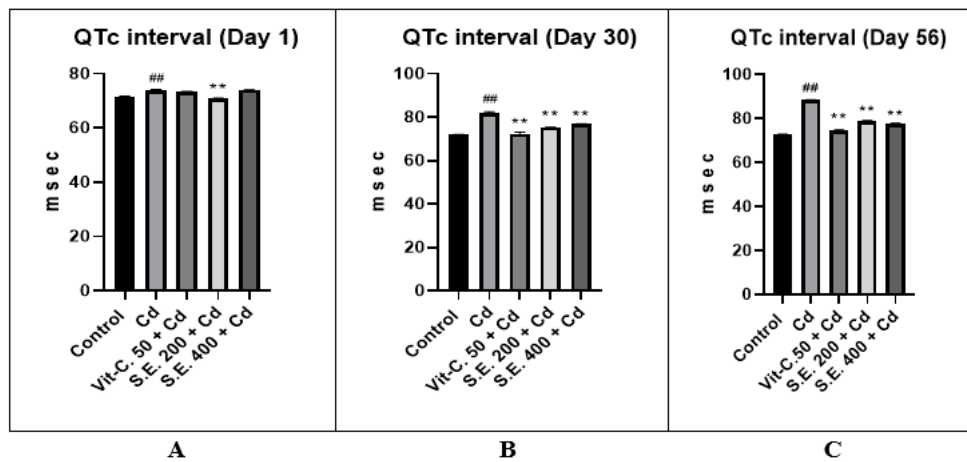


Figure 5: A-C Effect of alcoholic fruit extract of S.E. on ECG in QTc of heart in Cd-induced cardiotoxic rats.

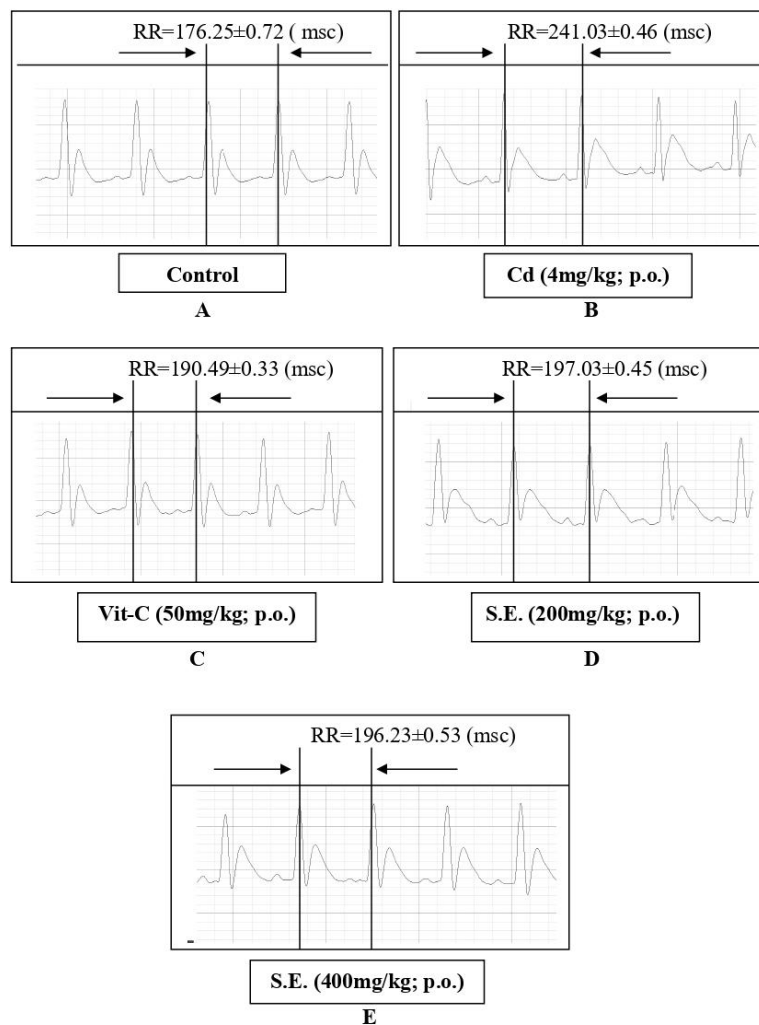


Figure 6: A-E Outcome alcoholic fruit extract of S.E. on ECG in RR interval of heart in Cd-induced Cardiotoxicity in rats.

3.3.6. Outcome alcoholic fruit extract of S.E. on ECG in RR interval of heart in Cd-induced cardiotoxic rats:

A. Effect of administration of cadmium (4mg/kg; p.o) on RR interval in rats.

RR interval in vehicle treated rats (Group-I) were found to be 168.14 ± 0.37 , 172.31 ± 0.48 and 176.25 ± 0.72 respectively on day 1st, 30th and 56th day. In Cd (4mg/kg) treated rat (Group-II) the RR interval was found to be 171.41 ± 0.34 , 216.80 ± 0.46 and 241.03 ± 0.46 respectively on day 1st, 30th and 56th day. Thus, in cadmium treated rats (Group-II) the RR interval was increase significantly over a period of 56th days when compared to (Group-I).

B. Effect of administration of Vitamin-C (50mg/kg; p.o.) on RR interval in Cd-induced cardiotoxic rats.

RR interval in rats treated with Vitamin-C (50mg/kg) in (group-III) were found to be 170.19 ± 0.46 , 184.01 ± 2.11 and 190.49 ± 0.327 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

C. Effect of administration of lower dose of alcoholic fruit extract of S.E. (200mg/kg; p.o.) in Cd-induced cardiotoxic rats.

RR interval in rats treated with lower dose of alcoholic fruit extract of S.E. (200mg/kg) in (Group-IV) were found to be 169.24 ± 0.55 , 187.15 ± 0.562 and 197.03 ± 0.46 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) in Cd-induced cardiotoxic rats.

RR interval in rats treated with higher dose of alcoholic fruit extract of S.E. (400mg/kg) in (Group-V) were found to be 171.35 ± 0.73 , 186.10 ± 0.49 and 196.23 ± 0.53 respectively on day 1st, 30th and 56th day. The above values were significantly decreased from day 30th to 56th day when compared to (Group-II).

Table 6: Effect of alcoholic fruit extract of S.E. on ECG in RR interval of heart in Cd-induced cardiotoxic rats.

Group s	Treatment	RR interval (msec)		
		Day 1	Day 30	Day 56
I	Control	168.14±0.3 7	172.31±0.4 8	176.25±0.72
II	Cd (4mg/kg; p.o)	171.41±0.3 4 ^{##}	216.80±0.4 6 ^{##}	241.03 ^{##} ± 0.46
III	Vit-C (50mg/kg; p.o) + Cd	170.19±0.4 5	184.01±2.1 1 ^{**}	190.49±0.33 ^{**}
IV	S.E (200 mg /kg; p.o) + Cd	169.24±0.5 5 [*]	187.15±0.5 6 ^{**}	197.03±0.45 ^{**}
V	S.E (400 mg /kg; p.o) + Cd	171.35±0.7 3	186.10±0.4 9 ^{**}	196.23±0.53 ^{**}

Values are expressed as mean ± SEM, n=6; ^{##} P<0.01 considered statistically significant as compare to normal control group; ^{***}P<0.001, ^{**} P<0.01 and ^{*} P<0.05 considered statistically significant when compared to Cd group.

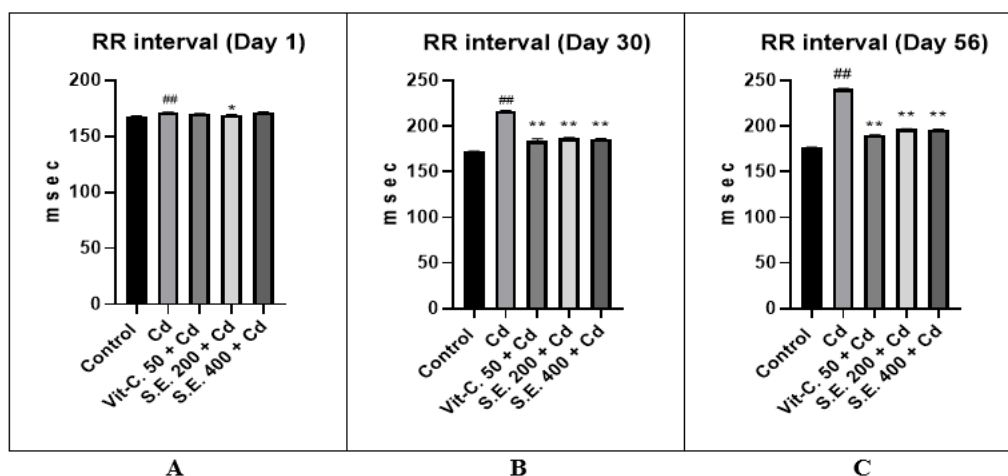


Figure 7: A-C Effect of alcoholic fruit extract of S.E. on ECG in RR interval of heart in Cd-induced cardiotoxic in rats.

3.3.7. Outcome alcoholic fruit extract of S.E. on heart rate in Cd-induced cardiotoxic in rats:

A. Effect of administration of Cd (4mg/kg; p.o.) on heart rate in rats.

Heart rate in vehicle treated rats (Group-I) were found to be 356.84 ± 0.80 , 348.42 ± 1.13 and 340.44 ± 1.40 respectively on day 1st, 30th and 56th day. In Cd (4mg/kg;) treated rat (Group-II) the heart rate was found to be 350.03 ± 0.70 , 276.74 ± 0.60 and 248.92 ± 0.48 respectively on day 1st, 30th and 56th day. Thus, in Cd treated rats (Group-II) the heart rate was significantly decreased over a period of 56th days when compared to (Group-I).

B. Effect of administration of Vitamin-C (50mg/kg; p.o.) on heart rate in Cd-induced cardiotoxic rats.

Heart rate in rats treated with Vitamin-C (50mg/kg) in (group-III) were found to be 352.52 ± 0.96 , 326.22 ± 3.72 and 315.77 ± 0.76 respectively on day 1st, 30th and 56th day. The above values were significantly increased from day 30th to 56th day when compared to (Group-II).

C. Effect of administration of lower dose of alcoholic fruit extract of S.E. (200mg/kg; p.o.) in Cd-induced cardiotoxic rats.

Heart rate in rats treated with lower dose of alcoholic fruit extract of S.E. (200mg/kg) in (Group-IV) were found to be 344.33 ± 0.99 , 320.58 ± 0.96 and 304.72 ± 0.71 respectively on day 1st, 30th and 56th day. The above values were significantly increased from day 30th to 56th day when compared to (Group-II).

D. Effect of administration of higher dose of alcoholic fruit extract of S.E. (400mg/kg; p.o.) in Cd-induced cardiotoxic rats.

Heart rate in rats treated with higher dose of alcoholic fruit extract of S.E. (400mg/kg) in (Group-V) were found to be 350.19 ± 2.10 , 322.40 ± 0.85 and 305.76 ± 0.82 respectively on day 1st, 30th and 56th day. The above values were significantly increased from day 30th to 56th day when compared to (Group-II).

Table 7: Effect of alcoholic fruit extract of S.E. on heart rate in Cd-induced cardiotoxic rats.

Groups	Treatment	Heart rate (BPM)		
		1 st day	30 th day	56 th day
I	Control	356.84 ± 0.80	348.42 ± 1.13	340.44 ± 1.39
II	Cd (4mg/kg; p.o)	350.03 ± 0.70 ^{##}	276.74 ± 0.59 [#]	248.92 ± 0.48 ^{##}
III	Vit-C (50mg/kg; p.o) + Cd	352.52 ± 0.96	326.22 ± 3.72 [*]	315.77 ± 0.76 ^{**}
IV	S.E (200 mg/kg; p.o) + Cd	344.33 ± 0.99	320.58 ± 0.96 [*]	304.72 ± 0.71 ^{**}
V	S.E (400 mg/kg; p.o) + Cd	350.19 ± 2.10	322.40 ± 0.85 [*]	305.76 ± 0.82 ^{**}

Values are expressed as mean \pm SEM, n=6; ## P<0.01 considered statistically significant as compare to normal control group; ***P<0.001, ** P<0.01 and * P<0.05 considered statistically significant when compared to Cd group. [BPM=Beats per minute]

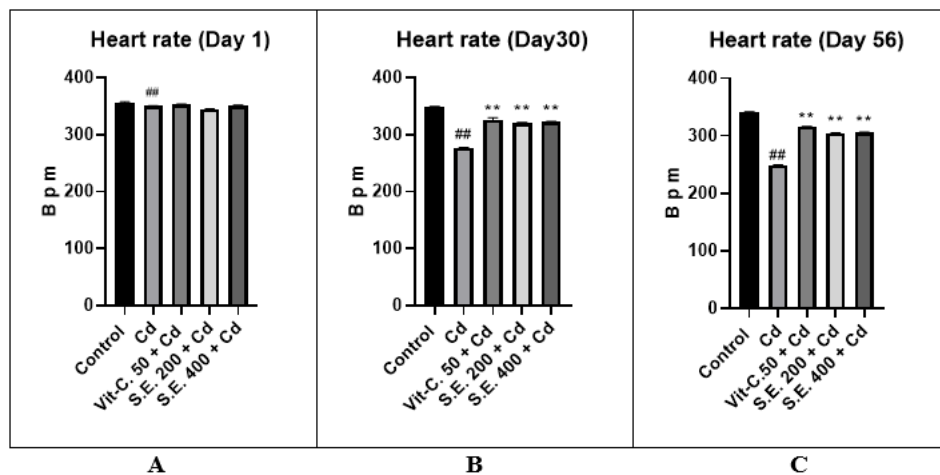


Figure 8: A-C Effect of alcoholic fruit extract of S.E. on heart rate in Cd-induced Cardiotoxic in rats.

3.3.8. Histopathological Observation:

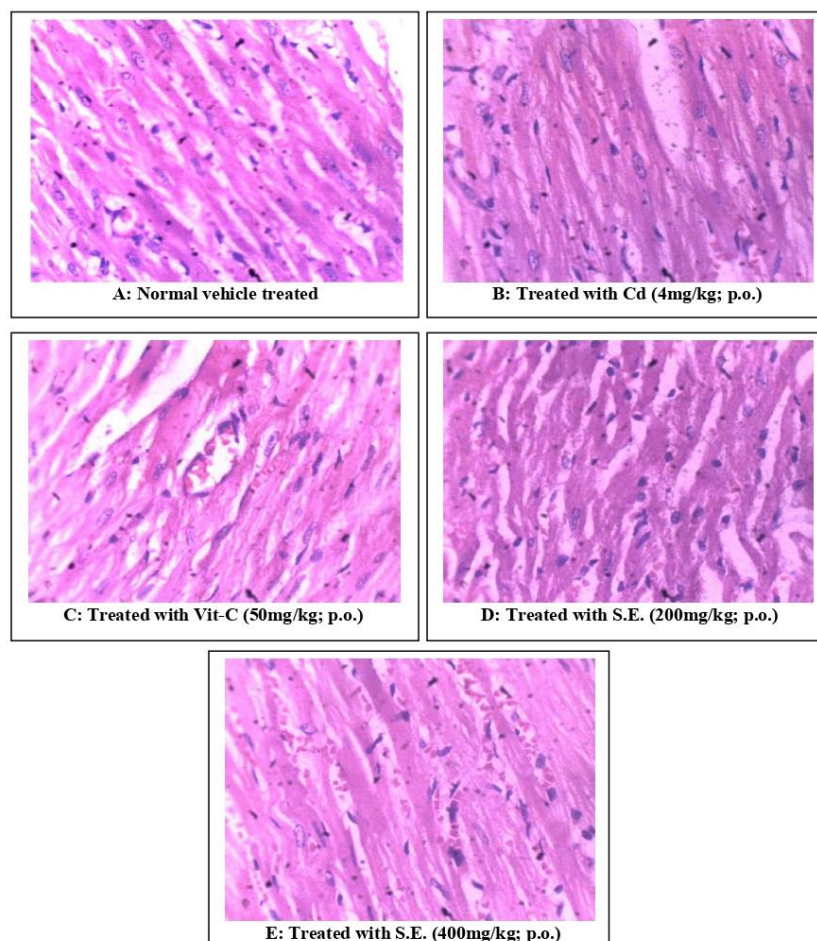


Figure 9: A-E Effect of alcoholic fruit extract of S.E. on H & E staining of heart in Cd-induced cardiotoxic rats.

In the Cd-induced cardiotoxicity model, the hearts of normal rats exhibited intact muscle fibres, while Cd-exposed hearts showed degenerative changes, fiber necrosis, and vascular damage. Treatment with *Sechium edule* extract restored normal cardiac architecture, indicating its potential cardioprotective effects.

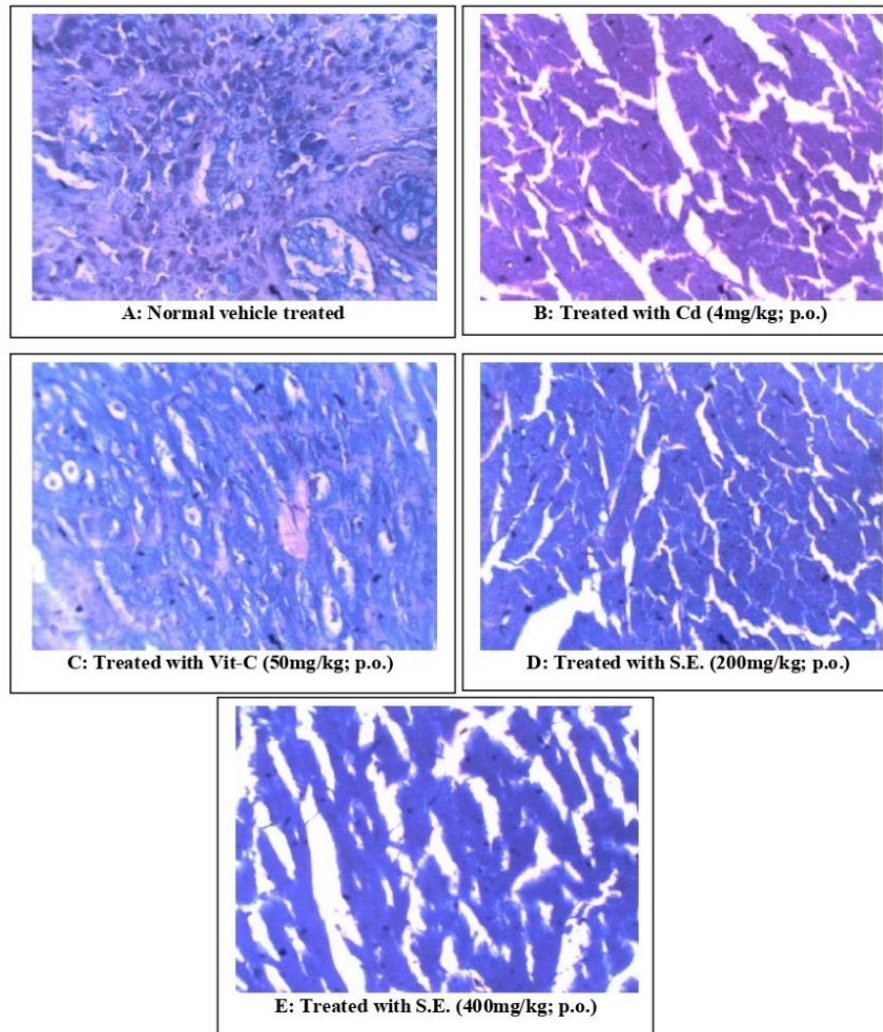


Figure 10: A-E Effect of alcoholic fruit extract of S.E. on Masson trichrome staining of heart in Cd-induced cardiotoxic rats.

In Masson trichrome staining, normal rat hearts exhibited normal collagen distribution, while Cd-induced cardiotoxic rats showed increased collagen bundles, indicative of fibrosis. Treatment with *Sechium edule* extract and Vit-C restored normal collagen appearance and cardiac architecture, suggesting potential anti-fibrotic effects.

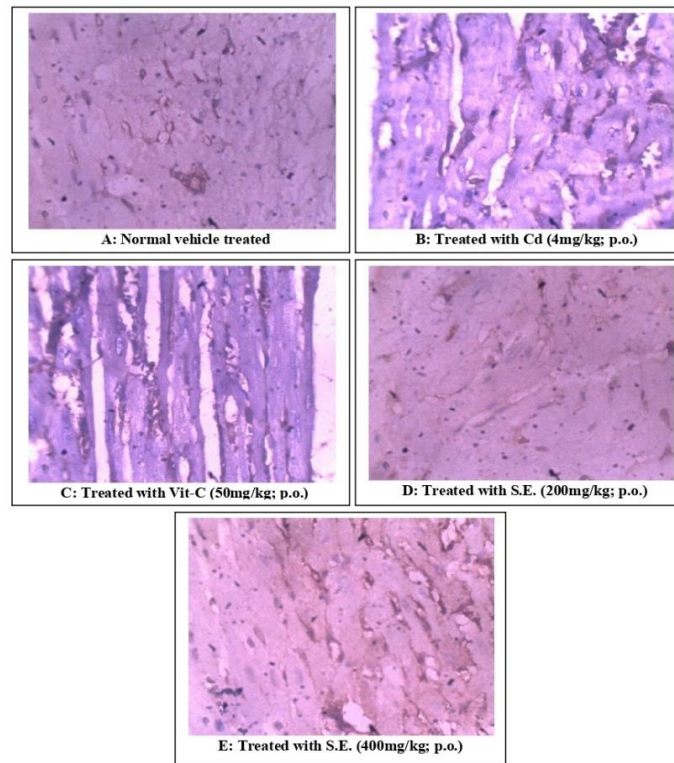


Figure 11: A-E Effect of alcoholic fruit extract of S.E. on staining with Bcl₂ of heart in Cd-induced cardio toxic rats.

Bcl₂ staining in myocardial tissue sections revealed moderate expression in normal control rats. Cd-induced group exhibited lower Bcl₂ expression, while Vit-C and *Sechium edule* (S.E.) treatments showed improved expression. S.E. treatment notably increased Bcl₂ expression, suggesting its potential anti-apoptotic effects in myocardial fibres compared to the Cd group.

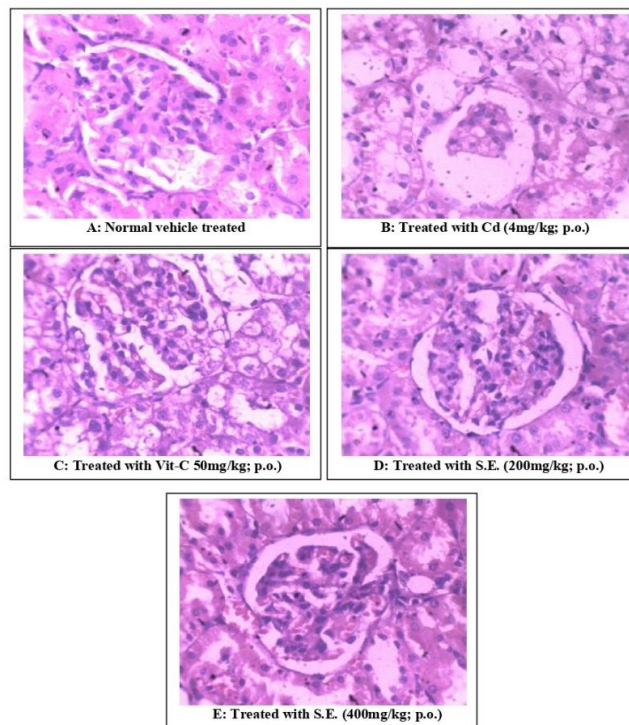


Figure 12: A-E Effect of alcoholic fruit extract of S.E. on H & E staining of kidney in Cd-induced cardiotoxic rats.

In the Cd-induced cardiotoxic model, normal rat kidneys had intact tubules and glomeruli, while Cd-exposed kidneys showed degenerative tubules, desquamated epithelial cells, misshapen tubules, and glomerular congestion. Treatment with Vit-C and *Sechium edule* extract restored almost normal cellular architecture, indicating potential renal protective effects.

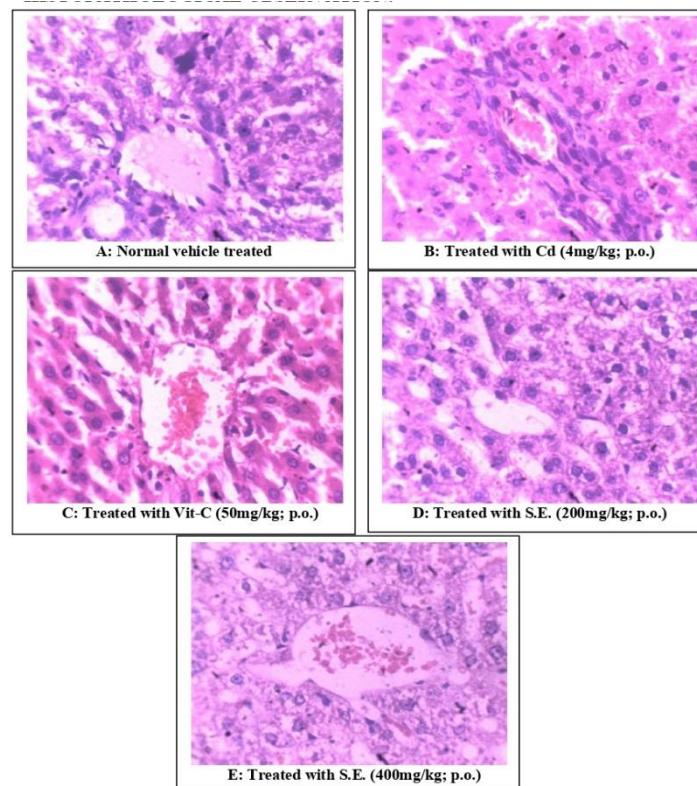


Figure 13: A-E Effect of alcoholic fruit extract of S.E. on H & E staining of liver in Cd-induced cardiotoxic rats.

In normal rat liver, hepatic lobules with portal triads were intact. Cd exposure led to severe damage, including fatty changes, focal necrosis, Kupffer cell proliferation, and bile duct loss. Vit-C and *Sechium edule* extract treatment partially improved hepatocyte structure, reduced bile duct loss, and alleviated focal necrotic areas, indicating potential hepatoprotective effects.

4. Discussion:

The presence of heavy metal ions, even at low concentrations, poses significant health risks due to their accumulation in tissues, causing metabolic, histological, and pathological changes in humans and mammals. Occupational exposure to cadmium (Cd) induces oxidative stress, generating reactive oxygen species that can lead to damage in organs, the nervous system, and even death. Cd accumulation in the heart causes peroxidative disorders, significantly increasing membrane lipid peroxidation markers like TBARS and protein carbonyls. Cd replaces redox-active metals, increasing free iron availability and triggering the Fenton reaction, resulting in damaging hydroxyl radical production and increased membrane lipid peroxidation.

Cd's thiol-affinitive nature primarily targets cellular glutathione (GSH), depleting the GSH pool and disrupting cellular redox balance, causing oxidative stress. Sub-chronic Cd treatment in the study significantly reduced GSH concentration in the heart. Cd-induced cardiotoxicity is associated with decreased superoxide dismutase (SOD) activity, causing detrimental effects on cells, such as mitochondrial enzyme inactivation, lipid peroxidation, and reactive nitrogen species production. Glutathione reductase (GR) activity, crucial for maintaining GSH levels, was also significantly decreased in the heart.

Cd exposure elevated serum markers indicating lipid abnormalities, cardiac damage, and impaired renal function. Treatment with *Sechium edule* (S.E.) extract significantly ameliorated these serum markers, suggesting its potential in protecting against cardiac damage and modulating lipid abnormalities.

Markers of cardiac function, including LDH, CPK, SGOT, and SGPT, were markedly elevated in Cd-induced cardiotoxic rats, indicating cardiomyocyte necrosis. S.E. extract treatment reduced these enzyme levels, suggesting protection against cardiac muscle damage.

ECG analysis revealed increased CVD risk in Cd-induced cardiotoxic rats, with prolonged QT, QTc, and RR intervals. S.E. extract treatment significantly decreased these intervals, indicating cardiac protection. Cd-induced bradycardia was also alleviated by S.E. extract treatment. Elevated Na⁺ and K⁺ ions in Cd-induced cardiotoxic rats suggested electrolyte imbalances and potential renal dysfunction, which were significantly decreased by S.E. extract treatment, indicating protection against cardiopathy and electrolyte imbalances.

Cd-induced renal damage, evidenced by increased serum urea and creatinine levels, was prevented by S.E. extract treatment. Histochemical analysis demonstrated severe histopathological changes in Cd-induced cardiotoxic rats, partially mitigated by S.E. extract treatment. Masson trichrome staining indicated fibrosis in Cd-treated hearts, partially alleviated by S.E. extract treatment.

Cd-induced liver damage, including fatty changes, focal necrosis, Kupffer cell proliferation, and bile duct proliferation, was partially improved by S.E. extract treatment.

5. Conclusion:

The ethanolic extract of *Sechium edule* (S.E.) significantly improved lipid profiles, decreased cardiac enzymes (LDH, CPK, SGOT, SGPT), lowered blood urea, serum creatinine, ALP levels, and regulated Na⁺, K⁺ ions in Cd-induced cardiotoxic rats. ECG parameters were normalized, and histopathological improvements were observed in the heart, liver, and kidney. S.E. exhibits potential antioxidant effects against Cd-induced cardiotoxicity.

References:

1. Satarug S. Long-term exposure to cadmium in food and cigarette smoke, liver effects and hepatocellular carcinoma. *Current Drug Metabolism*. 2012 Mar 1;13(3):257-71.
2. Sarwar N, Saifullah, Malhi SS, Zia MH, Naeem A, Bibi S, Farid G. Role of mineral nutrition in minimizing cadmium accumulation by plants. *Journal of the Science of Food and Agriculture*. 2010 Apr 30;90(6):925-37.
3. Satarug S, Garrett SH, Sens MA, Sens DA. Cadmium, environmental exposure, and health outcomes. *Environmental health perspectives*. 2010 Feb;118(2):182-90.
4. Satarug S, Haswell-Elkins MR, Moore MR. Safe levels of cadmium intake to prevent renal toxicity in human subjects. *British Journal of Nutrition*. 2000 Dec;84(6):791-802.
5. Goering PL, Waalkes MP, Klaassen CD. Toxicology of cadmium. In *Toxicology of metals: biochemical aspects 1995* (pp. 189-214). Berlin, Heidelberg: Springer Berlin Heidelberg.
6. ATSDR. Agency for toxic substance and disease registry, US toxicological profile for cadmium.
7. Urani C, Melchiorretto P, Canevali C, Morazzoni F, Gribaldo L. Metallothionein and hsp70 expression in HepG2 cells after prolonged cadmium exposure. *Toxicology in Vitro*. 2007 Mar 1;21(2):314-9.
8. Ziegler DM. Role of reversible oxidation-reduction of enzyme thiols-disulfides in metabolic regulation. *Annual review of biochemistry*. 1985 Jul;54(1):305-29.
9. Satarug S, Moore MR. Adverse health effects of chronic exposure to low-level cadmium in foodstuffs and cigarette smoke. *Environmental health perspectives*. 2004 Jul;112(10):1099-103.
10. El-Demerdash FM, Yousef MI, Kedwany FS, Baghdadi HH. Cadmium-induced changes in lipid peroxidation, blood hematology, biochemical parameters and semen quality of male rats: protective role of vitamin E and β -carotene. *Food and chemical toxicology*. 2004 Oct 1;42(10):1563-71.
11. Ercal N, Gurer-Orhan H, Aykin-Burns N. Toxic metals and oxidative stress part I: mechanisms involved in metal-induced oxidative damage. *Current topics in medicinal chemistry*. 2001 Dec 1;1(6):529-39.
12. Watanabe M, Henmi K, Ogawa KI, Suzuki T. Cadmium-dependent generation of reactive oxygen species and mitochondrial DNA breaks in photosynthetic and non-photosynthetic strains of *Euglena gracilis*. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*. 2003 Feb 1;134(2):227-34.
13. Casalino E, Sblano C, Landriscina C. Enzyme activity alteration by cadmium administration to rats: the possibility of iron involvement in lipid peroxidation. *Archives of biochemistry and biophysics*. 1997 Oct 15;346(2):171-9.
14. Waisberg M, Joseph P, Hale B, Beyersmann D. Molecular and cellular mechanisms of cadmium carcinogenesis. *Toxicology*. 2003 Nov 5;192(2-3):95-117.
15. Shaikh ZA, Vu TT, Zaman K. Oxidative stress as a mechanism of chronic cadmium-induced hepatotoxicity and renal toxicity and protection by antioxidants. *Toxicology and applied pharmacology*. 1999 Feb 1;154(3):256-63.

16. Akyolcu MC, Ozcelik D, Dursun S, Toplan S, Kahraman R. Accumulation of cadmium in tissue and its effect on live performance. In *Journal de Physique IV (Proceedings)* 2003 May 1 (Vol. 107, pp. 33-36). EDP sciences.
17. Zhang GH, Yamaguchi M, Kimura S, Higham S, Kraus-Friedmann N. Effects of heavy metal on rat liver microsomal Ca²⁺ (+)-ATPase and Ca²⁺ sequestering. Relation to SH groups. *Journal of Biological Chemistry*. 1990 Feb 5;265(4):2184-9.
18. Li W, Zhao Y, Cou IN. Alterations in cytoskeletal protein sulfhydryls and cellular glutathione in cultured cells exposed to cadmium and nickel ions. *Toxicology*. 1993 Jan 29;77(1-2):65-79.
19. Waisberg M, Joseph P, Hale B, Beyersmann D. Molecular and cellular mechanisms of cadmium carcinogenesis. *Toxicology*. 2003 Nov 5;192(2-3):95-117.
20. Mogensen CE. Microalbuminuria and hypertension with focus on type 1 and type 2 diabetes. *Journal of internal medicine*. 2003 Jul;254(1):45-66.
21. Maity S, Firdous SM, Debnath R. Evaluation of antidiabetic activity of ethanolic extract of *Sechium edule* fruits in alloxan-induced diabetic rats. *World J Pharm Pharm Sci*. 2013 Aug 7;2(5):3612-21..
22. Maity S, Firdous SM, Debnath R. Evaluation of antidiabetic activity of ethanolic extract of *Sechium edule* fruits in alloxan-induced diabetic rats. *World J Pharm Pharm Sci*. 2013 Aug 7;2(5):3612-21.
23. Firdous SM, Ahmed S, Dey S. Antiepileptic and central nervous system depressant activity of *Sechium edule* fruit extract. ||| *Bangladesh Journal of Pharmacology*|||. 2012 Sep 25;7(3):199-202.
24. Mumtaz SF, Paul S, Bag AK. Effect of *Sechium edule* on chemical induced kidney damage in experimental animals. *Bangladesh Journal of Pharmacology*. 2013;8(1):28-35.
25. Tsay HS, Agrawal DC. Tissue culture technology of Chinese medicinal plant resources in Taiwan and their sustainable utilization. *International Journal of Applied Science and Engineering*. 2005 Dec;3(3):215-23.
26. Thévenod F. Cadmium and cellular signaling cascades: to be or not to be?. *Toxicology and applied pharmacology*. 2009 Aug 1;238(3):221-39.
27. Kroemer G. Mitochondrial control of apoptosis. *Bulletin de L'academie Nationale de Medecine*. 2001 Jan 1;185(6):1135-42.
28. Chen CY, Zhang SL, Liu ZY, Tian Y, Sun Q. Cadmium toxicity induces ER stress and apoptosis via impairing energy homeostasis in cardiomyocytes. *Bioscience reports*. 2015 Jun 22;35(3):e00214.
29. Thévenod F, Lee WK. Toxicology of cadmium and its damage to mammalian organs. *Cadmium: from toxicity to essentiality*. 2013:415-90.